

THERAPEUTIC POTENTIAL OF “EXOSOMES DERIVED MULTIPLE ALLOGENEIC PROTEINS PARACRINE SIGNALING: EXOSOMES D-MAPPS” IS BASED ON THE EFFECTS OF EXOSOMES, IMMUNOSUPPRESSIVE AND TROPHIC FACTORS

Carl Randall Harrell¹, Crissy Fellabaum¹, Bojana Simovic Markovic², Aleksandar Arsenijevic² and Vladislav Volarevic²

¹Regenerative Processing Plant-RPP, LLC, 34176 US Highway 19 N Palm Harbor, Palm Harbor, Florida, United States of America

²Center for Molecular Medicine and Stem Cell Research, Faculty of Medical Sciences University of Kragujevac, Kragujevac, Serbia

TERAPIJSKI POTENCIJAL “EGZOSOMALNIH MULTIPLIH ALOGENIH PROTEINA ZA PARAKRINU SIGNALIZACIJU, EGZOSOM D-MAPPS” JE ZASNOVAN NA EFEKTIMA EGZOSOMA, IMUNOSUPRESIVNIH I TROFIČKIH FAKTORA

C. Randall Harrell¹, Crissy Fellabaum¹, Bojana Simović Marković², Aleksandar Arsenijević² i Vladislav Volarević²

¹Postrojenje za regenerativnu preradu - RPP, LLC, 34176 US Highway 19 N Palm Harbor, Palm Harbor, Florida, Sjedinjene Američke Države

²Centar za molekulska medicinu i istraživanje matičnih ćelija, Fakultet medicinskih nauka Univerziteta u Kragujevcu, Kragujevac, Srbija

Received / Priljen: 03. 09. 2018.

Accepted / Prihvaćen: 04. 09. 2018.

ABSTRACT

Due to their differentiation capacity and potent immunosuppressive and pro-angiogenic properties, mesenchymal stem cells (MSCs) have been considered as new therapeutic agents in regenerative medicine. Since most of MSC-mediated beneficial effects are a consequence of their paracrine action, we designed MSC-based product “Exosomes Derived Multiple Allogeneic Proteins Paracrine Signaling (Exosomes d-MAPPS)”, which activity is based on MSCs-derived growth factors and immunomodulatory cytokines capable to attenuate inflammation and to promote regeneration of injured tissues. Interleukin 1 receptor antagonist (IL-1Ra) and IL-27 were found in high concentrations in Exosomes d-MAPPS samples indicating strong anti-inflammatory and immunosuppressive potential of Exosomes d-MAPPS. Additionally, high concentrations of vascular endothelial growth factor receptor (VEGFR1) and chemokines (CXCL16, CCL21, CXCL14) were noticed at Exosomes d-MAPPS samples suggesting their potential to promote generation of new blood vessels and migration of CXCR6, CCR7 and CXCR4 expressing cells. Since all proteins which were found in high concentration in Exosomes d-MAPPS samples (IL-1Ra, CXCL16, CXCL14, CCL21, IL-27 and VEGFR1) are involved in modulation of lung, eye, and synovial inflammation, Exosomes d-MAPPS samples were prepared as inhalation and ophthalmic solutions in addition to injection formulations; their application in several patients suffering from chronic obstructive pulmonary disease, osteoarthritis, and dry eye syndrome resulted with significant improvement of biochemical and functional parameters. In conclusion, Exosomes d-MAPPS, due to the presence of important anti-inflammatory, immunomodulatory, and pro-angiogenic factors, represents potentially new therapeutic agent in regenerative medicine that should be further tested in large clinical studies.

Keywords: mesenchymal stem cells, therapy, regeneration, immunosuppression, differentiation

SAŽETAK

Mezenhimalne matične ćelije (MSCs), se zbog svojih imunomodulatornih i proangiogenih karakteristika, primenjuju u regenerativnoj medicini. Kako MSCs parakrinim mehanizmom ostvaruju svoje imunomodulatorne i proangiogene efekte, dizajnirali smo produkt „Egzosomalni multipli alogeni proteini za parakrinu signalizaciju (Egzosom d-MAPPS)“, koji sadrži egzosome, faktore rasta i citokine koje proizvode MSCs i njima smanjuju inflamaciju i pospešuju regeneraciju oštećenog tkiva.

Antagonist receptora IL-1 (IL-1 Ra) i interleukin (IL)-27, su pronađeni u visokim koncentracijama u ovom produktu, što je ukazivalo na snažan antiinflamacijski i imunosupresivni potencijal Egzosom d-MAPPS. Uz to, u Egzosom d-MAPPS je zabeležena i visoka koncentracija receptora za vaskularni endotelijalni faktor rasta (engl. vascular endothelial growth factor receptor, VEGFR1), kao i hemokina (CXCL16, CCL21, CXCL14), što ukazuje na potencijal Egzosom d-MAPPS da indukuje neo-angiogenezu i pospeši migraciju ćelija koje ekspimiraju CXCR6, CCR7 i CXCR4.

Pošto su sve komponente Egzosom d-MAPPS (IL-1Ra, CXCL16, CXCL14, CCL21, IL-27 i VEGFR1) uključene u modulaciju zapaljenja pluća, oka i zglobova, Egzosom d-MAPPS smo davali, u vidu inhalacionih rastvora, kapi za oči, ili intraartikularnih injekcija, pacijentima obolelih od hronične opstruktivne bolesti pluća, osteoartritisa i sindroma suvog oka. Preliminarni rezultati su pokazali značajno poboljšanje kako funkcionalnih tako i biohemijskih parametara nakon primene Egzosom d-MAPPS.

Egzosom D-MAPPS, zbog egzozoma, antiinflamacijskih, imunomodulatornih i proangiogenih faktora može da predstavlja nov terapijski agens u regenerativnoj medicini i njegov terapijski potencijal treba da se detaljnije ispita u kliničkim studijama sa velikim brojem pacijenata.

Ključne reči: mezenhimalne matične ćelije, terapija, regeneracija, imunosupresija, diferencijacija

Corresponding author:

Prof. dr. Vladislav Volarevic

Postal address: 69 Svetozar Markovic Street,

34000 Kragujevac, Serbia;

e-mail: drvolarevic@yahoo.com

tel./fax.: +38134306800



UDK: 602.9

Ser J Exp Clin Res 2019; 20 (3): 189-197

DOI: 10.2478/SJECR-2018-0032



INTRODUCTION

Mesenchymal stem cells (MSCs) are adult, self-renewable stem cells which are, due to their differentiation capacity and immunomodulatory characteristics, used as new therapeutic agents in regenerative medicine (1-3). MSCs are fibroblast-like cells that express: CD105 (endoglin, also identified as SH2, a component of the receptor complex of transforming growth factor- β (TGF- β) involved in proliferation, differentiation, and migration), CD73 (SH3/4, ectoenzyme that regulates the purinergic signaling through the hydrolysis of adenosine triphosphate (ATP)), CD44 (hyaluronan receptor involved in migration), CD90 (Thy-1, regulates differentiation of MSCs) [4]. Importantly, MSCs do not express CD14 (marker of monocytes), CD34 (marker of hematopoietic cells), CD45 (pan-leukocyte marker), CD79a and CD19 (marker of B lymphocytes) and lack expression of major histocompatibility complex (MHC) class II and co-stimulatory molecules, CD80 (B7-1), CD86 (B7-2), and CD40, suggesting a low immunogenicity *in vitro* and *in vivo* and their potential for safe allogeneic transplantation (5, 6).

MSCs have substantial differentiation potential. In addition to the cells of mesodermal origin (osteoblasts, chondroblasts, and adipocytes), MSCs are capable of generating neural cells, hepatocytes, alveolar epithelial cells, insulin-producing cells, cardiomyocytes, indicating their clinical application (7-14). Several lines of evidence suggested that MSCs have capacity to differentiate into functional cardiomyocytes (9), hepatocytes (10, 11), alveolar epithelial cells and lung precursor cells (13) contributing to the regeneration of injured myocardium, liver and lungs (12, 15-17). Additionally, in paracrine manner, through the production of immunomodulatory factors (indoleamine 2,3-dioxygenase (IDO), prostaglandin E2 (PGE2), nitric oxide (NO), transforming growth factor beta (TGF- β), interleukin (IL)-10, interleukin 1 receptor antagonist (IL-1Ra) and growth related oncogene (GRO), MSCs are able to suppress detrimental autoimmune response and to attenuate autoimmune and chronic, inflammatory diseases (3, 18, 19).

In addition to their immunomodulatory characteristics [3, 18, 19], MSCs may promote angiogenesis, as well. Through the production of several pro-angiogenic factors (vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF), TGF- β , platelet-derived growth factor (PDGF), angiopoietin-1, placental growth factor (PGF), IL-6, monocyte chemoattractant protein-1 (MCP-1), epidermal growth factor (EGF)), MSCs induce generation of new blood vessels having beneficial effects in the therapy of degenerative and ischemic cardiovascular and neurodegenerative diseases (20).

It was recently revealed that immunomodulatory and pro-angiogenic paracrine effects of MSCs are, at least partially, mediated by MSC-derived exosomes: nano-sized extracellular vesicles that deliver proteins, lipids, DNA fragments, mRNA to the target cells: immune cells, endothelial cells (ECs), pericytes and other tissue-resident cells (21).

MSC-derived exosomes, released into the extracellular milieu, can be either taken up by neighboring cells, (residing in the microenvironment of engrafted MSCs) or may be carried to distant sites via biological fluids where, in endocrine manner, modulate function of target cells (21, 22).

MSCs reside in perivascular niches of many diverse tissues and organs (bone marrow (BM), adipose tissue (AT), peripheral blood (PB), lungs, bone, heart, dental pulp (DP), amniotic fluid (AF), placenta (PL), chorion membrane (CM), chorion vili (CV), umbilical cord (UC), Wharton's jelly (WJ)) (23). Differences in extracellular milieu (influence of neighboring cells and their products, hypoxia) as well as intracellular conditions (expression of certain micro RNAs) significantly affect function and therapeutic potential of MSCs (23).

Several lines of evidence suggest that MSCs derived from placental tissues have superior cell biological properties such as improved proliferative capacity, life span and differentiation potential than MSCs derived from adult tissues. PL-MSCs have a higher expansion and engraftment capacity than BM-MSC [23]. Moreover, clonal subpopulations of PL-MSCs have been attributed with the potential to differentiate into tissues from all three germ layers. Accordingly, due to their capacity for neuronal differentiation, PL-MSCs have been proposed as one of the main candidates for stem cell therapy of multiple sclerosis, nerve injuries, and sensorineural hearing loss (23-26).

Ethical concerns related to the derivation of PL-MSCs should be disregarded by the fact that placental tissues are normally considered medical waste and can be recovered without harm to the donor or fetus (27). Bearing in mind the simplicity of the harvesting procedure for isolation of PL-MSCs and their huge therapeutic potential (28, 29), we recently developed: "Exosomes Derived Multiple Allogeneic Proteins Paracrine Signaling, Exosomes d-MAPPS", biological product which activity is based on placental derived biomaterials, growth factors, and immunomodulatory cytokines capable to attenuate inflammation and to promote regeneration of injured tissues. Herewith, we analyzed and discussed in detail concentrations of bio-active molecules in Exosomes d-MAPPS emphasizing its therapeutic potential in regenerative medicine.

MATERIAL AND METHODS

Exosomes d-MAPPS sample acquisition

Sterile Exosomes d-MAPPS is an engineered biologic product obtained from placental tissue, previously collected from healthy human donors. Blood samples were given by the donor prior to or at the time of collection and were tested by laboratories certified under the Clinical Laboratory Improvement Amendments (CLIA) and were found negative using United States (U.S) Food and Drug Administration (FDA) licensed tests for detection of at minimum: Hepatitis B Virus, Hepatitis C Virus, Human Immunodeficiency Virus Types 1/2, Treponema Pallidum.



Placental tissue samples were obtained with patient consent as well as institutional ethical approval and kept at 4°C until processed.

Exosomes d-MAPPS sample was engineered as a sterile product, manufactured under current Good Manufacturing Practices (cGMP) regulated and reviewed by the FDA. Sterile Exosomes d-MAPPS sample incorporate Regenerative Processing Plant's (RPP) proprietary patented sterilization process to provide safe sterile product.

Exosomes d-MAPPS samples, used in this study, were manufactured under specific conditions in order to be applicable for bioavailability testing and for different therapeutic use.

Determination of cytokines, chemokines, growth factors and their receptors in Exosomes d-MAPPS samples

Concentrations of cytokines, chemokines, growth factors and their receptors in Exosomes d-MAPPS samples were determined as previously described (30). Briefly, about fifty milliliters of sample was concentrated to 1.0-ml protein with trichloroacetic acid. The acetone-washed protein pellet was resolubilized in urea, and proteins were processed with dithiothreitol and iodoacetamide and digested with trypsin. Tryptic peptides were quantified and 10 µg was loaded through pressure cell onto a biphasic column for online two-dimensional high-performance liquid chromatography (HPLC) separation (strong-cation exchange and reversed-phase) and concurrent analysis by nanospray using a hybrid mass spectrometer. Three salt cuts of 50, 100, and 500 mM ammonium acetate were performed per sample run, with each followed by a 120-min organic gradient to separate the peptides.

Resultant peptide fragmentation spectra were compared with proteome database concatenated with common contaminants and reversed sequences to control false discovery rates. Peptide spectrum matches (PSMs) were filtered and assigned matched-ion intensities (MITs) based on observed peptide fragment peaks. PSM MITs were summed on a per-peptide basis, and only those uniquely and specifically matching a particular protein were moved onto subsequent analysis. Briefly, peptide intensity distributions were log-transformed, normalized across biological replicates by LOESS, and standardized by median absolute deviation and mean centering across samples as suggested. Peptides were then filtered to maintain at least two hits in one replicate set, and missing values were imputed using a random distribution of low-level values. Peptide abundance trends for each protein were scaled to a specific, well-sampled reference peptide. Sample-to-sample variation was visualized by PCA, Pearson's correlation and hierarchically clustered using the Ward agglomeration method to generate a heat map of protein abundance trends normalized by z-score (30).

RESULTS

Exosomes d-MAPPS has strong anti-inflammatory and immunomodulatory potential

Since MSCs produce immunosuppressive and anti-inflammatory factors (3, 18, 19), we analyzed concentration of major MSC-derived immunomodulatory molecules in Exosomes d-MAPPS sample (Figure 1). For this purpose, levels of IDO, IL-1ra, IL-10, IL-4, IL-13, IL-18 binding protein (IL-18 Bpa), TGFβ1 and Latency associated peptide of TGFβ1 (LAP (TGFβ1), were measured (Figure 1A). Among

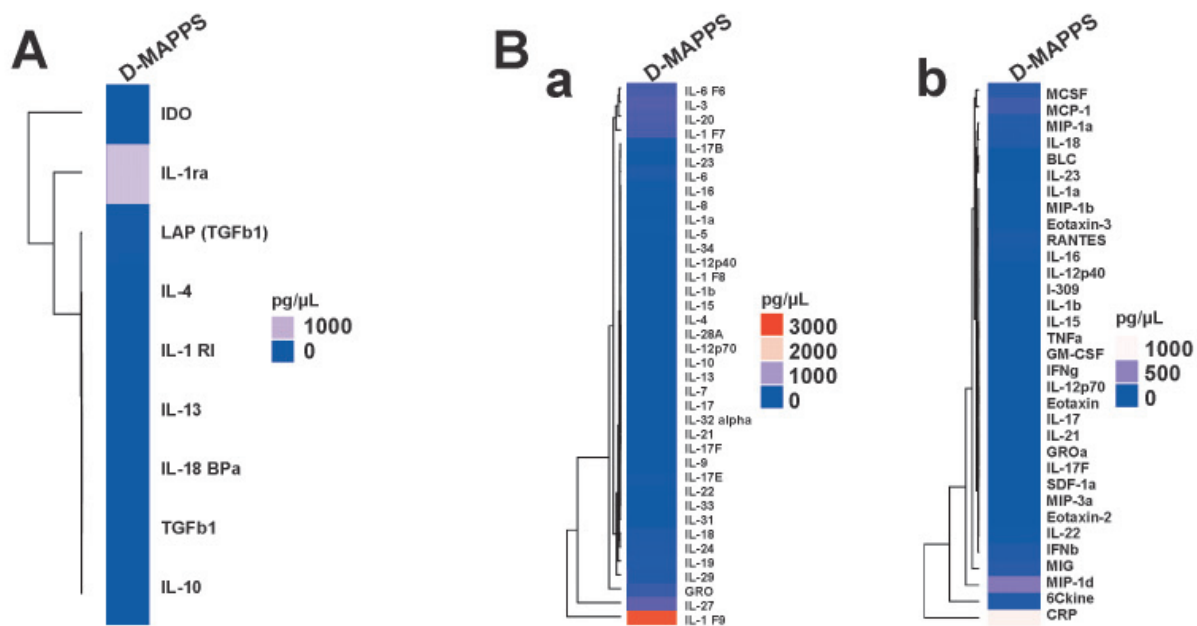


Figure 1: Inflammatory and immunomodulatory biomarkers in Exosomes d-MAPPS samples. (A) Different concentrations of 9 anti-inflammatory and immunomodulatory molecules are presented at heatmap. (B) Heatmap shows concentrations of 39 interleukins (a) and 33 inflammatory biomarkers (b) determined at Exosomes d-MAPPS sample.



measured immunoregulatory factors, IL-1Ra was found in high concentrations (1000 pg/μl). MSC-derived IL-1Ra is a naturally occurring cytokine which acts as an inhibitor of inflammatory cytokine IL-1. When IL-1Ra binds to the IL-1 receptor (IL-1R), binding of IL-1 is blocked and pro-inflammatory signal from IL-1 receptor is stopped. Accordingly, various pro-inflammatory events, initiated by IL-1:IL-1R binding, including the synthesis and releases of chemokines and enhanced influx of neutrophils, macrophages, and lymphocytes in inflamed tissues, are inhibited by IL-1Ra [4]. In line with these findings, high concentration of IL-1Ra, noticed in Exosomes d-MAPPS (Figure 1), indicates strong anti-inflammatory and immunomodulatory potential of this product.

In order to confirm strong anti-inflammatory properties of Exosomes d-MAPPS and to demonstrate that inflammatory mediators are not present in significant concentration in Exosomes d-MAPPS, levels of major inflammatory interleukins of innate and acquired immunity were evaluated (Figure 1B). As it is shown in Figure 1B, the main inflammatory cytokines of innate immunity (TNF-α, IL-1β, IL-12, IL-18) were not detected in Exosomes d-MAPPS sample. Similarly, Th1 (IFN-γ), Th2 (IL-4, IL-5, IL-10, IL-13) and Th17 (IL-17 and IL-22) cytokines were present in non-detectable concentrations indicating that neither one of T cell-dependent inflammatory pathways could not be elicited by Exosomes d-MAPPS. Among interleukins which might have dual (pro and anti-inflammatory) role (IL-6, IL-8, IL-27), only IL-27 was measured in

Exosomes d-MAPPS sample (1000 pg/μl). IL-27 promotes regulatory and immunosuppressive effects of MSCs by enhancing MSC-dependent generation of IL-10 producing CD4+T cells within the population of activated helper T cells. Additionally, capacity of MSCs to induce apoptosis of inflammatory Th1 and Th17 cells in programmed death ligand 1 (PDL1) dependent manner is significantly enhanced by IL-27 (31). Accordingly, presence of IL-27 could be considered as an additional indicator of immunosuppressive properties and therapeutic potential of Exosomes d-MAPPS sample.

In addition to IL-27, IL-1F9 was noticed in high concentration in Exosomes d-MAPPS sample (3000 pg/μl; Figure 1Ba). Since currently there is no available information regarding the role of IL-1F9 in MSC-based immunomodulation, the role of this cytokine in Exosomes d-MAPPS-based effects should be explored and analyzed in further studies.

Exosomes d-MAPPS can promote migration of CXCR6, CCR7 and CXCR4 expressing cells

One of the main properties of MSCs is their homing capacity towards the site of the injury or inflammation where they, in juxtacrine and/or paracrine manner, suppress detrimental immune response and ongoing inflammation [3]. MSCs expressed chemokine-specific receptors (CXCR4, CX3CR1, CXCR6, CCR1, and CCR7) and are attracted by chemokines (CXCL12, CXCL14, CX3CL1, CXCL16, CCL3, CCL19, and CCL21) released from damaged tissues and inflammatory immune cells (32). Interestingly, MSCs are also able to produce chemokines which, in autocrine manner, enable migration of MSCs towards the site of injury or inflammation (32). In line with these observations, we measured high concentration of MSCs-derived chemokine CXCL16 in Exosomes d-MAPPS sample (1500 pg/μl) (Figure 2A). Since CXCR6, ligand for CXCL16, is highly expressed on MSCs and immune cells (memory/effector T cells, NK, NKT cells and plasma cells) (32), high concentration of this chemokine in Exosomes d-MAPPS sample strongly indicates that Exosomes d-MAPPS can be used as chemoattractant enabling migration of CXCR6 expressing cells into the inflamed or injured tissues.

Similarly, 6Ckine (CCL21) (ligand for CCR7 receptor) is measured at Exosomes d-MAPPS sample (500 pg/μl) (Figure 2A). Having in mind that CCL21:CCR7 axis is important for migration of MSCs in wounds, homing of naïve T cells in peripheral lymph nodes and for migration of antigen processing, activated DCs into peripheral lymph nodes and T cell-rich fields within injured lungs, synovia and eyes (33-38), high levels of CCL21 in Exosomes d-MAPPS, could be used for recruitment of CCR7 expressing MSCs and immune cells during Exosomes d-MAPPS-based modulation of skin/joint/eye/lung inflammatory diseases. In line with these findings, high concentration (2000pg/ml; Figure 2A) of platelet factor 4 (PF4), which is involved in tissue regeneration and wound repair (39), was noticed in Exosomes d-MAPPS sample confirming its potential therapeutic use in regenerative medicine.

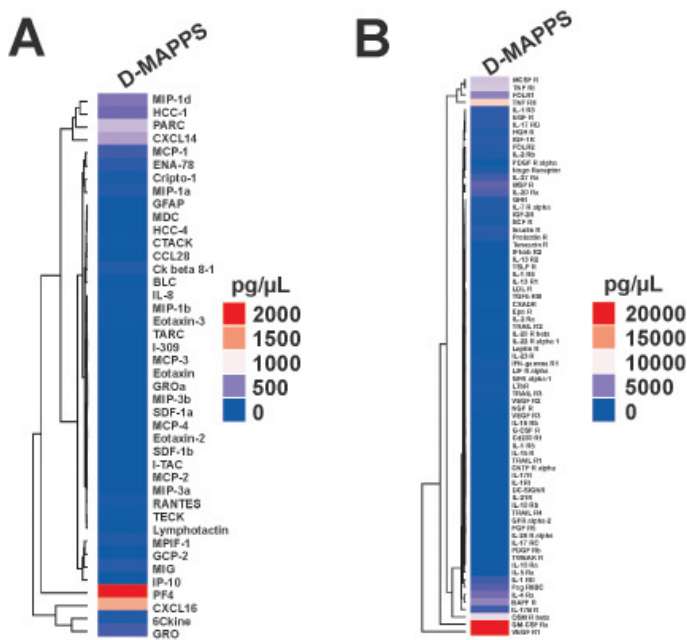


Figure 2: Chemokines and soluble receptors in Exosomes d-MAPPS sample. (A) Concentrations of 42 chemokines in Exosomes d-MAPPS samples are presented at heatmap. (B) Heatmap represents concentrations of 75 growth factor-related receptors and biomarkers in Exosomes d-MAPPS sample.



CXCL14 was also detected in Exosomes d-MAPPS sample (500 pg/ μ l; Figure 2A). CXCL14 specifically binds to CXCR4 and, in a similar manner as CXCL12, is involved in CXCR4-dependant migration of MSCs into injured or inflamed tissues [40].

In addition to elevated levels of CXCL16, CCL21, PF4 and CXCL14, GRO- γ well known MSC-derived chemokine with strong immunosuppressive properties [41], has been detected in Exosomes d-MAPPS sample (500 pg/ μ l; Figure 2A). Human MSCs secrete GRO- γ which, accompanied with GRO- α , promote conversion of monocyte derived DCs (MDDCs) towards myeloid suppressive phenotype enabling generation of tolerogenic myeloid derived suppressor cells (MDSCs) (42). In line with these findings, presence of GRO in Exosomes d-MAPPS sample, strongly indicates its potential for *in vitro* generation of MDSCs and MDSCs-based cell therapy of autoimmune and chronic inflammatory diseases.

Exosomes d-MAPPS has the capacity to induce neo-vascularization in VEGF-dependent manner

Having in mind that generation of new blood vessels and re-vascularization are mainly responsible for MSC-dependent regeneration of ischemic tissues (20), we evaluated presence of angiogenesis-related growth factor receptors in Exosomes d-MAPPS sample in order to explore capacity of Exosomes d-MAPPS to induce neo-angiogenesis-based tissue regeneration. As it is shown in Figure 2B, high concentrations of VEGFR1 (20000 pg/ μ l) was determined in Exosomes d-MAPPS sample. VEGFR1 plays critical role in migration of MSCs and MSCs-based neo-angiogenesis [43]. VEGFR1 binds VEGF and is expressed by multiple bone marrow-derived cell types, including endothelial progenitor cells and MSCs. BM-derived endothelial progenitor cells and MSCs are mobilized into peripheral blood and recruited to the sites of ischemia in VEGFR1-dependent manner, where they participate in tissue repair and revascularization (42). Based on these results, it is highly expected that Exosomes d-MAPPS can modulate generation and maturation of BM-derived cells. In line with these observations are high concentrations of granulocyte-macrophage colony-stimulating factor receptor (GM-CSFR) which was also noticed in Exosomes d-MAPPS sample (20000 pg/ μ l; Figure 2B). Since signaling from GM-CSFR can promote an astonishing variety of cellular functions, including protection from apoptosis, progression through the cell cycle, early commitment to myelopoiesis, differentiation/maturation of committed progenitors, and multiple activation and motility functions in mature immune cells (44), Exosomes d-MAPPS can be used for controlled differentiation of BM-derived, GM-CSFR expressing cells.

DISCUSSION

Due to their differentiation capacity and potent immunosuppressive and pro-angiogenic properties, MSCs have been considered as new therapeutic agents in regen-

erative medicine (45, 46). Nevertheless, safety issues of MSCs-based therapy are still a matter of debate, especially in the long-term follow up (47). Several studies reported that transplanted MSCs, in response to the growth factors produced in the local microenvironment, differentiated into undesired tissues, mainly bone and cartilage (48, 49). Multiple areas of ossifications or calcifications were observed in infarcted myocardium after transplantation of MSCs (48, 49). Since most of MSC-mediated beneficial effects are consequence of their paracrine action, we designed Exosomes d-MAPPS, soluble product, which contains a broad number of MSC-derived immunomodulatory and pro-angiogenic factors (Figures 1-2). Among anti-inflammatory mediators IL-1Ra was presented in the highest concentrations in Exosomes d-MAPPS (Figure 1A). Recently, a well-characterized subpopulation of IL-1Ra expressing MSCs have been described (50). MSCs, in IL-1Ra dependent manner, were able to suppress inflammation and fibrosis in the lungs. Interestingly, therapeutic and anti-inflammatory effects of MSCs-overexpressing IL-1Ra were more effective than effects of recombinant IL-1Ra (50), indicating that MSC-derived IL-1Ra acts synergistically with other immunomodulatory cytokines and chemokines in suppression of immune response.

Among chemokines, CXCL16, CCL21 and CXCL14 were present in high concentrations in Exosomes d-MAPPS sample (Figure 2A). Each of these molecules is crucially involved in the pathogenesis of lung, synovial, and eye inflammation (51-57). Bronchial epithelium is an important source of CXCL16 (51) while its receptor CXCR6 is highly expressed on lung-infiltrated T cells (51). Moreover, an increased expression of CXCL16 was noticed in the lungs of bleomycin-treated mice while CXCR6 expression was markedly increased in the lung in patients with interstitial lung diseases (53), indicating the importance of CXCL16: CXCR6 axis in the pathogenesis of lung injury and inflammation. Similarly, CXCL16 plays an important role in recruitment of T cells in inflamed synovium of patients suffering from Rheumatoid arthritis, suggesting CXCL16 as a target molecule in biological therapy of Rheumatoid arthritis [54].

In similar manner as CXCL16, CCL21 significantly contributes to the recruitment of CCR7 expressing immune cells in inflamed synovia (55). Additionally, both CCL21 and CCR7 are significantly up-regulated in inflamed corneas [56]. CCL21 facilitate migration of inflammatory, antigen presenting dendritic cells (DCs) in CCR7 dependent manner from the cornea to draining lymph nodes, while local administration of anti-CCL21 may reduce recruitment of DCs resulting with the attenuation of corneal inflammation (56).

Smoking-induced expression of CXCL14 in the airway epithelium represents a novel potential molecular link between smoking-associated airway epithelial injury, chronic obstructive pulmonary disease (COPD) and lung cancer. Airway epithelium responds to cigarette smoking with altered CXCL14 gene expression, contributing to the dis-



ease-relevant phenotype that result with the development of COPD and lung cancer (57).

Since bone repair and regeneration depend on vasculogenesis and osteogenesis, both of these processes are essential for successful bone remodeling (58). Several lines of evidence suggest that pro-angiogenic VEGF-A and VEGFR1 play crucially important role in bone regeneration (59, 60). Despite the fact that VEGF-A: VEGFR1 axis could contribute to the regeneration of the bone, VEGFR1 mediated signaling contributes to the development of complications of ischemic retinopathies, including retinopathy of prematurity (ROP), age-related macular degeneration (AMD), and diabetic retinopathy (DR) (61). VEGFR1 expression was up-regulated during pathogenesis of choroidal neovascularization (CNV), a model of AMD. Accordingly, blockade of VEGFR1 suppresses pathological angiogenesis and vascular leakage in the eye (61).

IL-27, measured in high concentrations in Exosomes d-MAPPS sample (Figure 1B) has dual: immunomodulatory and angiostatic effect in the injured eye. It inhibits pathophysiological intraocular neovascularization by reducing VEGF production in macrophages (62) and at the same time attenuate ongoing inflammation by suppressing proliferation of IL-17 producing Th17 cells (63). In similar manner, PDF4, which was also present in high concentration in Exosomes d-MAPPS sample (Figure 2A), limits generation of Th17 cells and is crucially involved in suppression of Th17 immune response (64). In this way, we assume that Exosomes d-MAPPS, in IL-27 and PDF4-dependant manner, may attenuate potentially detrimental effects of VEGFR1 signaling in Th17 cell driven eye injury and inflammation.

Since all proteins which were found in high concentration in Exosomes d-MAPPS samples (IL-1Ra, CXCL16, CXCL14, CCL21, IL-27, PDF4 and VEGFR1) are involved in modulation of lung, eye and synovial inflammation, we analyzed effects of Exosomes d-MAPPS-based therapy in patients suffering from chronic inflammatory diseases (COPD, osteoarthritis and dry eye syndrome). Our preliminary results, obtained in several pilot trials, indicated that Exosomes d-MAPPS was well tolerated since none of undesired, side effects were observed in Exosomes d-MAPPS-treated patients. Importantly, in patients that received Exosomes d-MAPPS, we noticed significant attenuation of inflammation, accompanied with an improvement of biochemical and functional parameters of injured lungs, knees and eyes.

Having in mind that therapeutic effects of Exosomes d-MAPPS are attributed to the MSC-derived soluble factors which could be found within MSC-derived exosomes (65), we believe that, at least some of Exosomes d-MAPPS-mediated beneficent effects were related to the function of exosomes. Accumulating evidence has suggested that MSCs, via exosomes, suppress detrimental immune response, attenuate inflammation and promote tissue repair and regeneration (21). Compared with cells, exosomes have no risk of aneuploidy and are well tolerated by the immune system

without the risk of rejection by immune cells after allogeneic transplantation (65). Additionally, due to their membrane-based structure, exosomes are able to cross the plasma membrane and to deliver their cargo into target cells throughout the body, indicating their potential to act in paracrine as well as endocrine manner (65). Accordingly, encouraging therapeutic effects of MSC-derived exosomes were observed in several animal models of organ specific and systemic inflammatory diseases, including acute and chronic injury of the eye and lungs (21, 66). It is well known that in early stage of corneal damage, injured epithelial cells produce IL-1 in order to elicit strong innate immune response (67). MSC-derived IL-1Ra, which was found in high concentration in Exosomes d-MAPPS sample (Figure 1A), binds to the IL-1R and prevents ongoing inflammation in the injured corneas. Similarly, it is well known that, in IL-1Ra dependent manner, MSCs are able to efficiently attenuate lung injury and inflammation (5). In line with these observations, we believe that IL-1Ra containing exosomes were responsible for the attenuated inflammation in Exosomes d-MAPPS-treated patients suffering from corneal injury and COPD.

In conclusion, Exosomes d-MAPPS, due to the presence of several important anti-inflammatory, immunomodulatory and pro-angiogenic factors, represents potentially a new therapeutic agent in regenerative medicine that should be further tested in large clinical studies.

ACKNOWLEDGMENTS

This study was supported by Macroproject of Faculty of Medical Sciences University of Kragujevac (MP 01/18).

CONFLICT OF INTEREST

Dr. C. Randall Harrell and Dr. Crissy Fellabaum are employed at RPP.

REFERENCES

1. Gazdic M, Volarevic V, Arsenijevic N, Stojkovic M. Mesenchymal stem cells: a friend or foe in immune-mediated diseases. *Stem Cell Rev* 2015;11:280-287.
2. Volarevic V, Ljubic B, Stojkovic P, Lukic A, Arsenijevic N, Stojkovic M. Human stem cell research and regenerative medicine-present and future. *Br Med Bull* 2011;99:155-168.
3. Volarevic V, Gazdic M, Simovic Markovic B, Jovicic N, Djonov V, Arsenijevic N. Mesenchymal stem cell-derived factors: Immunomodulatory effects and therapeutic potential. *Biofactors* 2017;43:633-644.
4. Volarevic V, Al-Qahtani A, Arsenijevic N, Pajovic S, Lukic ML. Interleukin-1 receptor antagonist (IL-1Ra) and IL-1Ra producing mesenchymal stem cells as modulators of diabetogenesis. *Autoimmunity* 2010;43:255-263.



5. Carraro G, Perin L, Sedrakyan S, Giuliani S, Tiozzo C, Lee J, Turcatel G, De Langhe SP, Driscoll B, Bellusci S, Minoo P, Atala A, De Filippo RE, Warburton D. Human amniotic fluid stem cells can integrate and differentiate into epithelial lung lineages. *Stem Cells* 2008;26:2902-2911.
6. Moorefield EC, McKee EE, Solchaga L, Orlando G, Yoo JJ, Walker S, Furth ME, Bishop CE. Cloned, CD117 selected human amniotic fluid stem cells are capable of modulating the immune response. *PLoS One* 2011;6:e26535.
7. Deng J, Petersen BE, Steindler DA, Jorgensen ML, Laywell ED. Mesenchymal stem cells spontaneously express neural proteins in culture and are neurogenic after transplantation. *Stem Cells* 2006; 24:1054-1064.
8. Yan ZJ, Hu YQ, Zhang HT, Zhang P, Xiao ZY, Sun XL, Cai YQ, Hu CC, Xu RX. Comparison of the neural differentiation potential of human mesenchymal stem cells from amniotic fluid and adult bone marrow. *Cell Mol Neurobiol* 2013;33:465-475.
9. Farzaneh M, Rahimi F, Alishahi M, Khoshnam SE. Paracrine mechanisms involved in mesenchymal stem cell differentiation into cardiomyocytes. *Curr Stem Cell Res Ther* 2018 Aug 21. doi: 10.2174/1574888X13666180821160421.
10. Zheng YB, Gao ZL, Xie C, Zhu HP, Peng L, Chen JH, Chong YT. Characterization and hepatogenic differentiation of mesenchymal stem cells from human amniotic fluid and human bone marrow: A comparative study. *Cell Biol Int* 2008;32:143-1448.
11. Wu Q, Tang J, Li Y, Li L, Wang Y, Bao J, Bu H. Hepatic differentiation of mouse bone marrow-derived mesenchymal stem cells using a novel 3D culture system. *Mol Med Rep* 2017;16:9473-9479.
12. Huang K, Kang X, Wang X, Wu S, Xiao J, Li Z, Wu X, Zhang W. Conversion of bone marrow mesenchymal stem cells into type II alveolar epithelial cells reduces pulmonary fibrosis by decreasing oxidative stress in rats. *Mol Med Rep* 2015;11:1685-1692.
13. Gong X, Sun Z, Cui D, Xu X, Zhu H, Wang L, Qian W, Han X. Isolation and characterization of lung resident mesenchymal stem cells capable of differentiating into alveolar epithelial type II cells. *Cell Biol Int* 2014;38:405-411.
14. Xie QP, Huang H, Xu B, Dong X, Gao SL, Zhang B, Wu YL. Human bone marrow mesenchymal stem cells differentiate into insulin-producing cells upon microenvironmental manipulation in vitro. *Differentiation* 2009;77:483-491.
15. Li YW, Zhang C, Sheng QJ, Bai H, Ding Y, Dou XG. Mesenchymal stem cells rescue acute hepatic failure by polarizing M2 macrophages. *World J Gastroenterol* 2017;23:7978-7988.
16. Guan XJ, Song L, Han FF, Cui ZL, Chen X, Guo XJ, Xu WG. Mesenchymal stem cells protect cigarette smoke-damaged lung and pulmonary function partly via VEGF-VEGF receptors. *J Cell Biochem* 2013;114:323-335.
17. Pandey AC, Lancaster JJ, Harris DT, Goldman S, Juneman E. Cellular Therapeutics for Heart Failure: Focus on Mesenchymal Stem Cells. *Stem Cells Int* 2017;2017:9640108.
18. Di Trapani M, Bassi G, Ricciardi M, Fontana E, Bifari F, Pacelli L, Giacomello L, Pozzobon M, Féron F, De Coppi P, Anversa P, Fumagalli G, Decimo I, Menard C, Tarte K, Krampera M. Comparative study of immune regulatory properties of stem cells derived from different tissues. *Stem Cells Dev* 2013;22:2990e3002.
19. Kode JA, Mukherjee S, Joglekar MV, Hardikar AA. Mesenchymal stem cells: immunobiology and role in immunomodulation and tissue regeneration. *Cytotherapy* 2009;11:377e91.
20. Tao H, Han Z, Han ZC, Li Z. Proangiogenic Features of Mesenchymal Stem Cells and Their Therapeutic Applications. *Stem Cells Int* 2016;2016:1314709.
21. Harrell CR, Simovic Markovic B, Fellabaum C, Arsenijevic A, Djonov V, Arsenijevic N, Volarevic V. Therapeutic Potential of Mesenchymal Stem Cell-Derived Exosomes in the Treatment of Eye Diseases. *Adv Exp Med Biol* 2018 May 18. doi: 10.1007/5584_2018_219.
22. Hyenne V, Apaydin A, Rodriguez D, Spiegelhalter C, Hoff-Yoessle S, Diem M, Tak S, Lefebvre O, Schwab Y, Goetz JG, Labouesse M. RAL-1 controls multivesicular body biogenesis and exosome secretion. *J Cell Biol* 2015;211:27-37.
23. Hass R, Kasper C, Böhm S, Jacobs R. Different populations and sources of human mesenchymal stem cells (MSC): A comparison of adult and neonatal tissue-derived MSC. *Cell Commun Signal* 2011;9:12.
24. Kil K, Choi MY, Kong JS, Kim WJ, Park KH. Regenerative efficacy of mesenchymal stromal cells from human placenta in sensorineural hearing loss. *Int J Pediatr Otorhinolaryngol* 2016;91:72-81.
25. Cho JS, Lee J, Jeong DU, Kim HW, Chang WS, Moon J, Chang JW. Effect of Placenta-Derived Mesenchymal Stem Cells in a Dementia Rat Model via Microglial Mediation: a Comparison between Stem Cell Transplant Methods. *Yonsei Med J* 2018;59:406-415.
26. Jiang H, Zhang Y, Tian K, Wang B, Han S. Amelioration of experimental autoimmune encephalomyelitis through transplantation of placental derived mesenchymal stem cells. *Sci Rep* 2017;7:41837.
27. Moore MC, Van De Walle A, Chang J, Juran C, McFetridge PS. Human Perinatal-Derived Biomaterials. *Adv Healthc Mater* 2017;6(18).
28. Farmer D. Placental stem cells: The promise of curing diseases before birth. *Placenta* 2017;59:113-115.
29. Abumaree MH, Abomaray FM, Alshabibi MA, AlAskar AS, Kalionis B. Immunomodulatory properties of human placental mesenchymal stem/stromal cells. *Placenta* 2017;59:87-95.
30. Xu Q, Resch MG, Podkaminer K, Yang S, Baker JO, Donohoe BS, Wilson C, Klingeman DM, Olson DG, Decker SR, Giannone RJ, Hettich RL, Brown SD, Lynd LR, Bayer EA, Himmel ME, Bomble YJ. Dramatic performance of *Clostridium thermocellum* explained by its wide range of cellulase modalities. *Sci Adv* 2016;2:e1501254.



31. Xu F, Yi J, Wang Z, Hu Y, Han C, Xue Q, Zhang X, Luan X. IL-27 regulates the adherence, proliferation, and migration of MSCs and enhances their regulatory effects on Th1 and Th2 subset generations. *Immunol Res* 2017;65:903-912.
32. Sordi V, Malosio ML, Marchesi F, Mercalli A, Melzi R, Giordano T, Belmonte N, Ferrari G, Leone BE, Bertuzzi F, Zerbini G, Allavena P, Bonifacio E, Piemonti L. Bone marrow mesenchymal stem cells express a restricted set of functionally active chemokine receptors capable of promoting migration to pancreatic islets. *Blood* 2005;106:419-427.
33. Yoshida R, Nagira M, Kitaura M, Imagawa N, Imai T, Yoshie O. Secondary lymphoid-tissue chemokine is a functional ligand for the CC chemokine receptor CCR7. *J Biol Chem* 1998;273:7118-71122.
34. Hocking AM. The Role of Chemokines in Mesenchymal Stem Cell Homing to Wounds. *Adv Wound Care (New Rochelle)* 2015;4:623-630.
35. Sasaki M, Abe R, Fujita Y, Ando S, Inokuma D, Shimizu H. Mesenchymal stem cells are recruited into wounded skin and contribute to wound repair by trans-differentiation into multiple skin cell type. *J Immunol* 2008;180:2581.
36. Xu B, Aoyama K, Kusumoto M, Matsuzawa A, Butcher EC, Michie SA, Matsuyama T, Takeuchi T. Lack of lymphoid chemokines CCL19 and CCL21 enhances allergic airway inflammation in mice. *Int Immunol* 2007;19:775-84.
37. Liu J, Wei Y, Luo Q, Xu F, Zhao Z, Zhang H, Lu L, Sun J, Liu F, Du X, Li M, Wei K, Dong J. Baicalin attenuates inflammation in mice with OVA-induced asthma by inhibiting NF- κ B and suppressing CCR7/CCL19/CCL21. *Int J Mol Med* 2016;38:1541-1548.
38. Jin Y, Shen L, Chong EM, Hamrah P, Zhang Q, Chen L, Dana MR. The chemokine receptor CCR7 mediates corneal antigen-presenting cell trafficking. *Mol Vis* 2007;13:626-34.
39. De Pascale MR, Sommese L, Casamassimi A, Napoli C. Platelet derivatives in regenerative medicine: an update. *Transfus Med Rev* 2015;29:52-61.
40. Hayashi Y, Murakami M, Kawamura R, Ishizaka R, Fukuta O, Nakashima M. CXCL14 and MCP1 are potent trophic factors associated with cell migration and angiogenesis leading to higher regenerative potential of dental pulp side population cells. *Stem Cell Res Ther* 2015;6:111.
41. Kuhn EN, Wu SM. Origin of cardiac progenitor cells in the developing and postnatal heart. *J Cell Physiol*
42. Chen HW, Chen HY, Wang LT, Wang FH, Fang LW, Lai HY, Chen HH, Lu J, Hung MS, Cheng Y, Chen MY, Liu SJ, Chong P, Lee OK, Hsu SC. Mesenchymal stem cells tune the development of monocyte-derived dendritic cells toward a myeloid-derived suppressive phenotype through growth-regulated oncogene chemokines. *J Immunol* 2013;190:5065-77.
43. Okuyama H, Krishnamachary B, Zhou YF, Nagasawa H, Bosch-Marce M, Semenza GL. Expression of vascular endothelial growth factor receptor 1 in bone marrow-derived mesenchymal cells is dependent on hypoxia-inducible factor 1. *J Biol Chem* 2006;281:15554-15563.
44. Hercus TR, Thomas D, Guthridge MA, Ekert PG, King-Scott J, Parker MW, Lopez AF. The granulocyte-macrophage colony-stimulating factor receptor: linking its structure to cell signaling and its role in disease. *Blood* 2009;114:1289-1298.
45. Srivastava M, Ahlawat N, Srivastava A. Amniotic Fluid Stem Cells: A New Era in Regenerative Medicine. *J Obstet Gynaecol India* 2018;68:15-19.
46. Loukogeorgakis SP, De Coppi P. Stem cells from amniotic fluid--Potential for regenerative medicine. *Best*
47. Volarevic V, Markovic BS, Gazdic M, Volarevic A, Jovicic N, Arsenijevic N, Armstrong L, Djonov V, Lako M, Stojkovic M. Ethical and Safety Issues of Stem Cell-Based Therapy. *Int J Med Sci* 2018;15:36-45.
48. Breitbach M, Bostani T, Roell W, Xia Y, Dewald O, Nygren JM, Fries JW, Tiemann K, Bohlen H, Hescheler J, Welz A, Bloch W, Jacobsen SE, Fleischmann BK. Potential risks of bone marrow cell transplantation into infarcted hearts. *Blood* 2007;110:1362-1369.
49. Yoon YS, Park JS, Tkebuchava T, Luedeman C, Losordo DW. Unexpected severe calcification after transplantation of bone marrow cells in acute myocardial infarction. *Circulation* 2004;109:3154-157.
50. Ortiz LA, Dutreil M, Fattman C, Pandey AC, Torres G, Go K, Phinney DG. Interleukin 1 receptor antagonist mediates the antiinflammatory and antifibrotic effect of mesenchymal stem cells during lung injury. *Proc Natl Acad Sci U S A* 2007; 104:11002-11007.
51. Day C, Patel R, Guillen C, Wardlaw AJ. The chemokine CXCL16 is highly and constitutively expressed by human bronchial epithelial cells. *Exp Lung Res* 2009; 35:272-283.
52. Morgan AJ, Guillen C, Symon FA, Huynh TT, Berry MA, Entwisle JJ, Briskin M, Pavord ID, Wardlaw AJ. Expression of CXCR6 and its ligand CXCL16 in the lung in health and disease. *Clin Exp Allergy* 2005; 35:1572-1580.
53. Huang L, Zhang L, Ju H, Li Q, Pan JS, Al-Lawati Z, Sheikh-Hamad D. Stanniocalcin-1 inhibits thrombin-induced signaling and protects from bleomycin-induced lung injury. *Sci Rep* 2015;5:18117.
54. Nanki T, Shimaoka T, Hayashida K, Taniguchi K, Yonehara S, Miyasaka N. Pathogenic role of the CXCL16-CXCR6 pathway in rheumatoid arthritis. *Arthritis Rheum* 2005;52:3004-3014.
55. Rump L, Matthey DL, Kehoe O, Middleton J. An initial investigation into endothelial CC chemokine expression in the human rheumatoid synovium. *Cytokine* 2017;97:133-140.
56. Jin Y, Shen L, Chong EM, Hamrah P, Zhang Q, Chen L, Dana MR. The chemokine receptor CCR7 mediates corneal antigen-presenting cell trafficking. *Mol Vis* 2007;13:626-634.



57. Shaykhiev R, Sackrowitz R, Fukui T, Zuo WL, Chao IW, Strulovici-Barel Y, Downey RJ, Crystal RG. Smoking-induced CXCL14 expression in the human airway epithelium links chronic obstructive pulmonary disease to lung cancer. *Am J Respir Cell Mol Biol* 2013;49:418-425.
58. Zaidi N, Nixon AJ. Stem cell therapy in bone repair and regeneration. *Ann N Y Acad Sci* 2007;1117:62-72.
59. Hu K, Olsen BR. The roles of vascular endothelial growth factor in bone repair and regeneration. *Bone* 2016;91:30-38.
60. Clarkin CE, Gerstenfeld LC. VEGF and bone cell signalling: an essential vessel for communication? *Cell Biochem Funct* 2013;31:1-11.
61. Huang H, Shen J, Viores SA. Blockade of VEGFR1 and 2 suppresses pathological angiogenesis and vascular leakage in the eye. *PLoS One* 2011; 6:e21411.
62. Hasegawa E, Oshima Y, Takeda A, Saeki K, Yoshida H, Sonoda KH, Ishibashi T. IL-27 inhibits pathophysiological intraocular neovascularization due to laser burn. *J Leukoc Biol* 2012;91:267-273.
63. Amadi-Obi A, Yu CR, Liu X, Mahdi RM, Clarke GL, Nussenblatt RB, Gery I, Lee YS, Egwuagu CE. TH17 cells contribute to uveitis and scleritis and are expanded by IL-2 and inhibited by IL-27/STAT1. *Nat Med* 2007;13:711-718.
64. Shi G, Field DJ, Ko KA, Ture S, Srivastava K, Levy S, Kowalska MA, Poncz M, Fowell DJ, Morrell CN. Platelet factor 4 limits Th17 differentiation and cardiac allograft rejection. *J Clin Invest* 2014;124:543-552.
65. Yu B, Zhang X, Li X. Exosomes derived from mesenchymal stem cells. *Int J Mol Sci* 2014;15:4142-4157.
66. Cruz FF, Rocco PRM. Stem-cell extracellular vesicles and lung repair. *Stem Cell Investig* 2017;4:78.
67. Yamada J, Dana MR, Sotozono C, Kinoshita S. Local suppression of IL-1 by receptor antagonist in the rat model of corneal alkali injury. *Exp Eye Res* 2003;76:161-167.