

Antibacterial activity of iron oxide nanoparticles synthesized by co--precipitation technology against *Bacillus cereus* and *Klebsiella pneumoniae*

Shakeel Ahmed Ansari^{1*}, Mohammad Oves², Rukhsana Satar³, Anish Khan⁴, Syed Ismail Ahmad⁵, Mohammad Alam Jafri¹, Syed Kashif Zaidi¹, Mohammad Husein Alqahtani¹

¹King Abdulaziz University, Center of Excellence in Genomic Medicine Research, Jeddah-21589, Kingdom of Saudi Arabia ²King Abdulaziz University, Center of Excellence in Environmental Studies, Jeddah-21418, Kingdom of Saudi Arabia ³Ibn Sina National College for Medical Sciences, Department of Biochemistry, Jeddah-21418, Kingdom of Saudi Arabia ⁴King Abdulaziz University, Center of Excellence for Advanced Materials Research, Jeddah-21589, Kingdom of Saudi Arabia ⁵Ibn Sina National College for Medical Sciences, Physics Division, Basic Science Department, Jeddah-21418, Kingdom of Saudi Arabia

*Corresponding author: e-mail: shakeel.cegmr@gmail.com

The present study investigates the synthesis and characterization of iron oxide nanoparticles (Fe₃O₄-NPs) for their antibacterial potential against *Bacillus cereus* and *Klebsiella pneumonia* by modified disc diffusion and broth agar dilution methods. DLS and XRD results revealed the average size of synthesized Fe₃O₄-NPs as 24 nm while XPS measurement exhibited the spin-orbit peak of Fe $2p_{3/2}$ binding energy at 511 eV. Fe₃O₄-NPs inhibited the growth of *K. pneumoniae* and *B. cereus* in both liquid and soild agar media, and displayed 26 mm and 22 mm zone of inhibitions, respectively. MIC of Fe₃O₄-NPs was found to be 5 μ g/mL against these strains. However, MBC for these strains was observed at 40 μ g/mL concentration of Fe₃O₄-NPs for exhibiting 40–50% loss in viable bacterial cells and 80 μ g/mL concentration of Fe₃O₄-NPs acted as bactericidal for causing 90–99% loss in viability. Hence, these nanoparticles can be explored for their additional antimicrobial and biomedical applications.

Keywords: antibacterial activity, co-precipitation, iron oxide nanoparticles, Klebsiella pneumoniae, physicochemical characterization.

Abbreviations: Fe₃O₄-NPs, iron oxide nanoparticles; MBC, minimum bactericidal concentration; MIC, minimum inhibitory concentration; XPS; X-ray photoelectron spectroscopy.

INTRODUCTION

Antibiotics have been a mainstay of human health. They have been extensively used in clinics to treat respiratory tract infections, urinary tract infections, gastrointestinal infections and infections of nervous system¹. However, the emergence of multidrug resistant bacterial strains such as Methicillin resistant staphylococcus aureus (MRSA), Vancomycin resistant staphylococcus aureus (VRSA) and New Delhi metallo-beta-lactamase (NDM) harboring Gram-negative bacteria, has severely compromised their effectiveness in the clinics. Therefore, novel antibiotics with newer mechanism of action are urgently needed to treat infections caused by MDR-bacterial strains. Numerous nanoparticle have been reported to possess considerable activity against MDR-bacterial strains²⁻⁵ including resistant Enterobacteriaceae (E. coli, K. pneumoniae), non-fermenters (Acinetobacter baumanii, Pseudomonas aeruginosa) and acid-fast bacteria (Mycobacterium tuberculosis) as well as resistant Gram-positives such as vancomycin resistant Enterococci and methicillin-resistant Staphylococcus aureus^{5, 6}. Furthermore, new strategies like surface modification of nanoparticles are being explored to enhance their potency against pathogenic MDR bacterial strains⁷.

Klebsiella pneumoniae is an encapsulated Gram-negative rod-shaped facultative anaerobe belonging to *Enterobacteriaceae* family which causes pneumonia, bloodstream infections, surgical site infections and meningitis⁸. The NDM carrying *Klebsiella pneumoniae* stains are frequently involved in nosocomial (hospital-acquired) and common public infections. These strains are resistant to almost every antibiotic and infections caused by them are extremely difficult to treat. Moreover, *Klebsiella pneumoniae* strains have the potential to produce slime, which facilitates the adhesion and formation of biofilms on implantable devices via non-specific interactions like electrostatic, dipole-dipole, H-bond, hydrophobic and vander Waals forces^{9, 10}. Formation of such biofilms enhances the bacterial resistance by preventing antibiotic action. *Bacillus cereus* is Gram positive, rod shaped bacterium, widely distributed in diverse environments¹¹. It is commonly involved in food poisoning causing severe diarrhea. However, in immunocompromised patients it is associated with wound infections, bacteremia, septicaemia, endocarditis, meningitis and pneumonia as well as respiratory infections which may be life threatening¹².

Iron oxides are abundantly distributed in the earth crust and have been frequently used in a wide variety of biomedical and biotechnological applications. Nanoparticles synthesized from these oxides have also been employed for diagnostic imaging (magnetic resonance imaging), cancer treatment (magnetic hyperthermia, thermal ablation), scale-up bioseparation processes and biosensing-based applications^{13, 14}. Fe₃O₄-NPs-based biomedical applications have received considerable attention due to their ease of laboratory synthesis, cost effectiveness, physical and chemical stability, as well as biocompatibility and environmental safety^{15–17}. Therefore, Fe₃O₄-NPs may be reasonable candidates for their potential use as antibacterial therapy. The current study was undertaken to synthesize Fe₃O₄-NPs by co-precipitation method for evaluating their antimicrobial activity against Klebsiella pneumoniae and Bacillus cereus.

EXPERIMENTAL METHODS

Materials

Ferrous chloride tetrahydrate (FeCl₂ · 4H₂O), ferric chloride hexahydrate (FeCl₃ · 6H₂O) and sodium hydroxide (NaOH) were obtained from Sisco Research Laboratories Pvt. Ltd. (Mumbai, India) and used as such without further purification. The bacterial culture media, nutrients broth and nutrient agar were purchased from Hi Media, Mumbai, India. The deionized water was used for the preparation of reagents and culture media. The bacterial strains *Klebsiella pneumoniae* and *Bacillus cereus* used in this study were obtained from local hospital.

Synthesis of Fe₃O₄-NPs

The co-precipitation method was used for the synthesis of Fe_3O_4 -NPs. Briefly, ferric chloride (200 mg) and ferrous chloride (100 mg) were dissolved in deionized water (25 mL) under continuous stirring and heated at 60°C. The solution was bubbled with Argon for preventing unwanted oxidation. Subsequently, 2.5 M NaOH (15 mL) was injected at 60°C and the reaction was continued under similar incubation conditions for 20 minutes before the flask was removed from heating and stirring. Magnetic field was applied to separate Fe_3O_4 -NPs formed as black precipitate and obtained Fe_3O_4 -NP were thoroughly washed with distilled water and finally transferred to an oven at 100°C for overnight drying. Dried power of Fe_3O_4 -NPs was used for subsequent analysis and physical characterization.

Physico-chemical characterization of Fe₃O₄-NPs

The synthesized Fe_3O_4 -NPs were subjected for recording FTIR spectrum at 500-4000 cm⁻¹ by using Interspec 2020 model FTIR instrument (USA). The dynamic light scattering characterization was carried out in a Malvern Zetasizer Nano ZS while their structural characterization was done by analyzing XRD pattern at room temperature with the help of Rigaku-Miniflex X-ray diffractometer using CuK_a ($\lambda = 1.54056$ Å) radiations as X-ray source in 2q range from 20° to 80° with scan rate of 2°/min. For XPS analysis of Fe₃O₄-NPs, we used a Thermo Scientific K-Alpha KA1066 spectrometer with monochromatic AlKa X-ray radiation as excitation source and beam-spot size fixed at 300 µm. The spectral recording was performed at 200 eV (pass energy) in the fixed analyzer transmission mode whereas scanning was carried out at pressures of less 10^{-8} Torr.

Antibacterial assay

Antibacterial test of Fe_3O_4 -NPs was performed against *Klebsiella pneumonia* and *Bacillus cereus* bacterial stains. The inoculum of test organisms was prepared by inoculating flasks containing sterilized 25 mL of nutrient broth with the isolated colonies of test organisms and incubating them at 37°C overnight on an orbital shaker incubator. The overnight cultures were diluted with fresh nutrient broth and spread onto solid nutrients agar plates with the help of sterilized glass spreader or cotton swab. The plates were left to dry for 15 min and then 8 nm wells were bored in each plate with the help of a sterilized borer. The different concentrations of Fe_3O_4 -NPs aqueous suspension were loaded in the wells of each plate and one

well served as a control for standard antibiotics. Then, each well was sealed by molten agar (0.8%) to prevent the leakage of loaded compounds in base of well. The plates were incubated at 37°C for 24 hours. Following incubation, the developed zone of inhibition of bacterial growth around the well was recorded.

Determination of minimum inhibition concentration (MIC) and minimum bactericidal concentration (MBC)

Klebsiella pneumonia and Bacillus cereus overnight cultures were diluted with nutrient broth in order to obtain a final inoculum of 1×10⁶ CFU/mL. Stock solution of Fe₃O₄-NPs for MBC and MIC were separately prepared. Different concentrations of Fe₃O₄-NPs were added to the tubes containing test organism. Positive and negative controls were also prepared separately with and without Fe_3O_4 -NPs and organisms, respectively. The tubes were incubated at 37°C in an incubator overnight. Turbidity was checked for analyzing the growth of organism in presence of compound. Lowest concentration which inhibited the growth of organism indicated MIC whereas MBC was considered as a concentration that caused 2-3 log (90-99%) reduction in CFU compared to starting inoculum. All MIC and MBC experiment were performed in duplicate.

Statistical analysis

For antibacterial activity, the size of growth inhibition zones was expressed in terms of mean value \pm standard error and a P value lower than 0.05 was taken as a significant value.

RESULTS AND DISCUSSION

Selection of Fe₃O₄-NPs

We selected iron oxide nanoparticles because of their relatively better safety and superior biological compatibility. These nanoparticles have been reported to have lower toxicity due to the presence of 4 unpaired electrons in 3d shell of Fe^{2+} ions and 5 unpaired electrons of 3d shell of Fe³⁺ which facilitate strong magnetic moment. Therefore, the crystals formed by these ions are ferromagnetic or ferrimagnetic in nature which provides them better biocompatibility and lower toxicity^{13, 15, 18}. The co-precipitation method was adopted due to the simplicity of their synthesis in aqueous solution. This method involves precipitation of Fe²⁺ and Fe³⁺ ions (mixed in molar ratio of 1:2) into Fe₃O₄ in a basic solution (ammonium hydroxide, potassium hydroxide, sodium hydroxide, pH 8-14). However, it has been reported that size, morphology and magnetic properties of nanoparticles synthesized by co-precipitation method vary significantly. In the current study, Fe₃O₄-NPs synthesis was carried out in presence of NaOH which yielded nanoparticle of 24 nm average size.

Furthermore, for wider bioapplications of iron oxide NPs, they should be highly dispersed in aqueous phase and biocompatible with negligible toxicity. These properties can be achieved by adopting co-precipitation method using multifunctional water-soluble polymers as capping ligands¹⁹. Hence, the method of synthesis utilized here for obtaining Fe_3O_4 -NPs imparts uniformity in size and

morphology to them. Moreover, due to their higher saturation magnetization, they could be manipulated in dispersed state with the use of an external magnet for extending their scope in various biomedical applications.

Physico-chemical characterization of Fe₃O₄-NPs

The infrared spectrum of Fe_3O_4 -NPs in the range of 500–4000 cm⁻¹ was carried out to identify the chemical bonds and functional groups present in it. Fig 1 demonstrated the O-H stretching vibration in O-H groups as suggested by the large broad band at 3625 cm⁻¹. The



Figure 1. FTIR spectra of Fe₃O₄-NPs

absorption peaks around 1597 cm⁻¹ correspond to the asymmetric and symmetric bending vibration of C=O while the absorption peak around 1024 cm⁻¹ is ascribed to polyvinylpyrrolidone. The strong band observed below 700 cm⁻¹ was produced by Fe-O stretching mode while the Fe-O stretching mode band of Fe₃O₄ was observed at 598 cm^{-1 20}. The average mean size of synthesized Fe₃O₄. -NPs was in the range of 20–25 nm (Fig. 2). Figure 3



Figure 2. Differential light scattering of Fe₃O₄-NPs

suggested that the size of the synthesized Fe_3O_4 -NPs were of 24 nm. The results showed the spinel phase structure of magnetite which is in agreement with the standard XRD for magnetite nanoparticles synthesized by other methods^{13, 18}. X-ray photoelectron spectroscopy (XPS) is frequently used for elucidating numerous chemical parameters including the presence of various elements, empirical-formula, and chemical and electronic states of the elements constituting the test material. XPS spectra are obtained by irradiating test materials with a beam of X-rays and determining the kinetic energy as well as the number of electrons released from the top of the material surface^{21, 22}. We used XPS measure-



Figure 3. XRD analysis of Fe₃O₄-NPs

ments in order to find out the chemical states of iron and oxygen in Fe₃O₄-NPs. XPS spectrum of Fe₃O₄-NPs is presented in Figure 4. The spin-orbit peak of the Fe $2p_{3/2}$ binding energy observed at 511 eV (Fig. 4b) is fully concordant with the standard reference data for iron²³. A distinct peak observed at 532.6 eV in O1s spectrum (Fig. 4c) indicated the presence of oxygen (i.e. O₂⁻) in the Fe₃O₄-NPs²⁴. Therefore, it can be concluded that the synthesized Fe₃O₄-NPs contains only two elements and is free from other impurities.

Antibacterial activity of Fe₃O₄-NPs

Resistance to currently used antibiotics is a serious public health problem nowadays. The prevalence of bacterial isolates with resistance to multiple antibiotics has increased significantly in both hospitals as well as in community. Novel antibiotics are needed to effectively treat infections caused by these resistant strains. The nanoparticles have been extensively evaluated for their potential as suitable antibacterial candidates which may be developed as antibacterial therapy for clinical application. Numerous organic and inorganic nano-materials as well as a large number of nanoparticles have been synthesized for this purpose of providing safe and effective antimicrobial agents against MDR bacterial strains. Fe₃O₄-NPs exhibit considerable antibacterial effects because of their specific features including their size and larger surface area which facilitates their interactions with critically important biomolecules inside the bacterial cell. In the current study, Fe₃O₄-NPs demonstrated significant antibacterial activity against both Bacillus cereus and Klebsiella pneumoniae strains. The exact molecular mechanisms of antibacterial action of iron oxide nanoparticles are less understood. However, it is possible that they exert their antibacterial action through the interactions of ferric ions with the proteins and DNA thereby causing the conformational changes in protein structure and DNA molecule, which ultimately leads to the mortality of bacterial cell. However, generation of reactive oxygen species (ROS) has also been implicated as one of the possible mechanism of antibacterial action of iron nanoparticles. These nanoparticles also displayed a bactericidal action against Klebsiella pneumoniae and Bacillus cereus which



Figure 4. X-ray photoelectron spectroscopy of Fe₃O₄-NPs

is an important attribute from pharmacodynamic point of view as bactericidal antibiotics completely eradicate bacterial cells which reduces chances of emergence of resistant bacterial strains.

In our investigation, Fe₃O₄-NPs were employed to evaluate their bactericidal potential for *Klebsiella pneumonia* and Bacillus cereus (Fig. 5). They showed antibacterial activity in a concentration dependent manner with higher concentrations exhibiting larger zones of inhibition. The zone of inhibition was 15 and 13 mm at 40 μ g/well concentration of Fe₃O₄-NPs against *Klebsiella pneumoniae* and Bacillus cereus respectively. However, at higher concentration of 80 μ g/well, the zone size was increased to 26 and 22 mm (Fig. 6). We also determined MIC values of Fe₃O₄-NPs against these strains. MIC value of Fe₃O₄-NPs was found to be 5 μ g/mL for *Klebsiella pneumoniae* and *Bacillus cereus*. Further, we also tried to find out whether antibacterial action of Fe₃O₄-NPs was bacteriostatic or bactericidal. MBC experiments were performed in liquid broth media and results were obtained in terms of % viability of bacterial cells following 24 h exposure to different concentrations of Fe₃O₄-NPs (Fig. 7). The significant killing of *Klebsiella pneumoniae* and *Bacillus cereus* cells was observed at 40



Figure 5. Antimicrobial potential of synthesized Fe_3O_4 -NPs doses (i) 80 and (ii) 60 μ g/well on solid agar media (A) K. pneumonia and (B) B. cereus



Figure 6. Zone of inhibition pattern of different dose against Gram negative *K. pneumonia* and Gram positive *B. cereus*



Figure 7. Viability of bacterial cell in presence of different dose of nanomaterials in liquid medium

 μ g/mL exhibiting around 40–50% loss in viable bacterial cells whereas 80 μ g/mL concentration of Fe₃O₄-NPs was found to be bactericidal causing 90–99% loss in viability. Mechanistically, these nanoparticles may cause damage to bacterial cell membrane either directly binding to lipid molecules or through the release of ROS which may cause intracellular damage leading to a powerful bactericidal action. Similar mechanism has been proposed for positively charged nanoparticles against various bacterial strains^{25–27}. However, exact molecular nature of antibacterial action of Fe₃O₄-NPs remains to be clearly elucidated.

CONCLUSION

The nanoparticles have great potential as nanomedicine for the treatment of various bacterial diseases. The ultra-small size of nanoparticles may facilitate their diffusion through porin channels of bacterial cell wall to exert their antibacterial action via destruction of the membrane integrity and intracellular damage. However, robust preclinical studies are required to establish their potential as antibacterial agents as well as to demonstrate sound proof of concept in the animal models of infection.

ACKNOWLEDGEMENT

This article was funded by the Deanship of Scientific Research (DSR), King Abdulaziz University, Jeddah. The authors, therefore, acknowledge with thanks DSR technical and financial support.

CONFLICT OF INTEREST

We have no conflict of interest to declare.

LITERATURE CITED

1. Singh, R., Smitha, M.S. & Singh, S.P. (2014). The role of nanotechnology in combating multi-drug resistant bacteria. J. Nanosci. Nanotechnol. 14(7), 4745–4756. DOI: 10.1166/jnn.2014.9527.

2. Stubbings, W. & Labischinski, H. (2009). New antibiotics for antibiotic-resistant bacteria. Biol. Rep. 17(1) 40–46. DOI: 10.3410/B1-40.

3. Patrascu, J.M., Nedelcu, I.A., Sonmez, M., Ficai, D., Ficai, A. & Vasile, B.S. (2015). Composite scaffolds based on silver nanoparticles for biomedical applications. J. Nanomat. 8 pages. DOI: http://dx.doi.org/10.1155/2015/587989.

4. Caamano, M.A., Carrillo-Morales, M. & Olivares-Trejo, J.J. (2016). Iron oxide nanoparticle improve the antibacterial activity of erythromycin. J. Bacteriol. Parasitol. 7(4), 267–270. DOI: 10.4172/2155-9597.1000267.

5. Kalishwaralal, K., Barathmanikanth, S., Pandian, S.R.K., Deepak, V. & Gurunathan, S. (2010). Silver nanoparticles impede the biofilm formation by *Pseudomonas aeruginosa* and *Staphylococcus epidermidis*. Coll. Surf. B: Biointerf. 79(2), 340–344. DOI: 10.1016/j.colsurfb.2010.04.014.

6. Mihu, M.R., Sandkovsky, U., Han, G., Friedman, J.M., Nosanchuk, J.D. & Martinez, L.R. (2010). The use of nitric oxide releasing nanoparticles as a treatment against *Acinetobacter baumannii* in wound infections. Virulence 1(2), 62–67. DOI: 10.4161/viru.1.2.10038.

7. Satar, R., Syed, I.A., Rasool, M., Pushparaj, P.N. & Ansari, S.A. (2016). Investigating the antibacterial potential of agarose nanoparticles synthesized by nanoprecipitation technology. Pol. J. Chem. Technol. 18(2), 9–12. DOI: https://doi.org/10.1515/pjct-2016-0022.

8. Fang, C.T., Lai, S.Y., Yi, W.C., Hsueh, P.R., Liu, K.L. & Chang, S.C. (2007). *Klebsiella pneumoniae* genotype K1: an emerging pathogen that causes septic ocular or central nervous system complications from pyogenic liver abscess. Clin. Infect. Dis. 45(3), 284–290. DOI: 10.1086/519262.

9. Donlan, R.M. (2001). Biofilms and device-associated infections. Emer. Inf. Dis. 7(2), 277–281. DOI: 10.3201/ eid0702.700277.

10. Jagnow, J. & Clegg, S. (2003). *Klebsiella pneumoniae* MrkD-mediated biofilm formation on extracellular matrix and collagen-coated surfaces. Microbiology 149(9), 2397–2405. DOI: 10.1099/mic.0.26434-0.

11. Ash, C., Farrow, J.A., Dorsch, M., Stackenbrandt, E. & Collins, M.D. (1991). Comparative analysis of *Bacillus anthracis, Bacillus cereus*, and related species on the basis of reverse transcriptase of 16S rRNA. Int. J. Syst. Bacteriol. 41(3), 343–346. DOI: 10.1099/00207713-41-3-343.

12. Bottone, E.J. (2010). *Bacillus Cereus*, a volatile human pathogen. Clin. Microbiol. Rev. 23(2), 382–398. DOI: 10.1128/CMR.00073-09.

13. Wu, W., He, Q. & Jiang, C. (2008). Magnetic iron oxide nanoparticles: Synthesis and surface functionalization strategies. Nan. Res. Lett. 3(11), 397–415. DOI: 10.1007/s11671-008-9174-9.

14. Mohapatra, M. & Anand, S. (2010). Synthesis and applications of nanostructured iron oxides/hydroxides-a review. Int. J. Eng. Sci. Technol. 2(8), 127–146. DOI: http://dx.doi. org/10.4314/ijest.v2i8.63846. 15. Hui, C., Shen, C., Yang, T., Bao, L., Tian, J. & Ding, H. (2008). Large-scale Fe_3O_4 nanoparticles soluble in water synthesized by a facile method. J. Phys. Chem. C 112(30), 11336–11339. DOI: 10.1021/jp801632p.

16. Ahmed, T., Phul, R., Khatoon, N. & Sardar, M. (2017). Antibacterial efficacy of *Ocimum sanctum* leaf extract-treated iron oxide nanoparticles. New J. Chem. 41(5), 2055–2061. DOI: 10.1039/C7NJ00103G.

17. Irshad, R., Tahir, K., Li, B., Ahmad, A., Siddiqui, A. & Nazir, S. (2017). Antibacterial activity of biochemically capped iron oxide nanoparticles: A view towards green chemistry. J. Photochem. Photobiol. B 170(4), 241–246. DOI: 10.1016/j. jphotobiol.2017.04.020.

18. Mahdavi, M., Ahmad, M.B., Haron, M.J., Gharayebi, Y., Shameli, K. & Nadi, B. (2013). Fabrication and characterization of SiO₂/(3-aminopropyl) triethoxysilane-coated magnetite nanoparticles for lead (II) removal from aqueous solution. J. Inorg. Organomet. Polym. Mater. 23(3), 599–607. DOI: 10.1007/s10904-013-9820-2.

19. Majeed, M.I., Guo, J., Yan, W. & Tan, B. (2016). Preparation of magnetic iron oxide nanoparticles (MIONS) with improved saturation magnetization using multifunctional polymer ligand. Polymers 8(11), 392–408. DOI: 10.3390/polym8110392.

20. Gotic, M. & Music, S. (2007). Mossbauer FT-IR and FE SEM investigation of iron oxides precipitated from $FeSO_4$ solutions. J. Nanostruct. 834–836(7), 445–453. DOI: https://doi. org/10.1016/j.molstruc.2006.10.059.

21. Zhang, F., Wang, P., Koberstein, J., Khalid, S. & Chan, S.W. (2004). Cerium oxidation state in ceria nanoparticles studied with X-ray photoelectron spectroscopy and absorption near edge spectroscopy. Surf. Sci. 563(1–3), 74–82. DOI: https://doi.org/10.1016/j.susc.2004.05.138.

22. Bavand, R., Yelon, A. & Sacher, E. (2015). X-ray photoelectron spectroscopic and morphologic studies of Ru nanoparticles deposited onto highly oriented pyrolytic graphite. Appl. Surf. Sc. 355(5), 279–289. DOI: https://doi.org/10.1016/j. apsusc.2015.06.202.

23. Yamashita, T. & Hayes, P. (2008). Analysis of XPS spectra of Fe²⁺ and Fe³⁺ ions in oxide materials. Appl. Surf. Sci. 254(8), 2441–2449. DOI: https://doi.org/10.1016/j.apsusc.2007.09.063.

24. Rahman, M.M., Khan, S.B., Faisal, M., Rub, M.A., Al-Youbi, M.A. & Asiri, A.M. (2012). Electrochemical determination of olmesartan medoxomil using hydrothermally prepared nanoparticles composed $\text{SnO}_2\text{-}\text{Co}_3\text{O}_4$ nanocubes in tablet dosage forms. Talanta 99(2), 924–931. DOI: https://doi. org/10.1016/j.talanta.2012.07.060.

25. Kon, K. & Rai, M. (2013). Metallic nanoparticles: mechanism of antibacterial action and influencing factors. J. Comp. Clin. Path. Res. 2(3), 160–2174. DOI: 10.4178/jccph/e2015020.

26. Franci, G., Falanga, A., Galdiero, S., Palomba, L., Rai, M. & Morelli, G. (2015). Silver nanoparticles as potential antibacterial agents. Molecules 20(5), 8856–8874. DOI: 10.3390/ molecules20058856.

27. Li, H., Chen, Q., Zhao, J. & Urmila, K. (2015). Enhancing the antimicrobial activity of natural extraction using the synthetic ultrasmall metal nanoparticles. Sci. Rep. 5(5), 11033–11040. DOI: 10.1038/srep11033.