

Histamine content in rennet ripening cheeses during storage at different temperatures and times

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Abstract

Introduction: In recent years, there has been a great interest in biogenic amines such histamine, as they are associated with the quality and safety of some kinds of fermented foods. The aim of this study was to evaluate the effect of temperature and storage time on the content of histamine in cheeses. **Material and Methods:** Samples of mould and hard cheeses were examined with RP-HPLC with an organic-aqueous mobile phase containing acidic buffer and chaotropic salt. The samples were stored either at $22 \pm 2^{\circ}$ C for 42 days (mould and hard cheeses) or at $4 \pm 2^{\circ}$ C for 112 days (mould cheeses) and 133 days (hard cheeses). **Results:** The mean total histamine content in cheeses stored at 22°C was higher than the content in those stored at 4°C, with the highest concentrations found in Gorgonzola Piccante cheese (730.47 mg/kg). Histamine concentration in some types of cheeses exceeded the toxic threshold dose, indicating that after long or inadequately cool storage they may not be safe for consumers. **Conclusion:** To protect cheeses from contamination with histamine-producing bacteria and to safeguard consumers from poisoning, factors conducive to this amine's formation should be minimised during cheese processing. Suitable temperature and time during storage of cheeses are recommended to avoid the intoxication. Monitoring of this toxin in food is necessary to ensure safety of consumers.

Keywords: histamine, cheeses, HPLC-DAD, storage conditions, health risk.

Introduction

Histamine is a heterocyclic biogenic amine which can be formed in a large variety of types of food of animal origin, particularly in fish and cheeses. Histamine is a product of decarboxylation of histidine in a reaction, catalysed by bacterial decarboxylase (5, 32). Cheeses are a favourable environment for the formation of this amine due to the presence of free amino acids and microorganisms releasing decarboxylases (4, 19). Histamine production in cheeses can be influenced by many factors, e.g. the type of bacterial starter culture, cheese pH, salt during concentration, contamination production process, storage temperature, and ripening time (16, 24, 27, 30). The most common bacterial species responsible for production of histamine Morganella morganii, Photobacterium are: psychrotolerans, Photobacterium phosphoreum, *Lactobacillus casei, Klebsiella pneumoniae, Enterobacter aerogenes,* and *Hafnia alvei* (14, 17, 25). The highest amount of histamine is produced during the ripening of cheeses, when proteins are degraded to free amino acids including histidine (6, 9, 24, 31).

Cheeses are one of the most common products responsible for histamine poisoning in humans (11, 23). This intoxication manifests itself in headaches, hypotension, vomiting, diarrhoea, cramping, skin rashes and swellings (3). Histamine is degraded in the body by oxidative deamination catalysed by monoamine and diamine oxidases (21, 26). However, after ingestion of food with high concentrations of histamine, in the absence or scarcity of these enzymes (*e.g.* due to genetic factors, allergy, or ingested alcohol) the detoxification system is unable to eliminate histamine sufficiently (3, 15).

In 1967 the first report was made of histamine poisoning after consumption of Gouda cheese with

histamine in the range of 850–1,870 mg/kg (8). Other literature data reported histamine levels in cheeses up to 2,500 mg/kg (3, 10).

The provisions of European Union regulations specify a permissible level of histamine only for fish and fish products (7). Currently, there are no established criteria relating to the level of this amine in cheeses. The aim of this study was to determine the histamine content in hard and mould cheeses stored at different temperatures and time periods.

Material and Methods

Sample collection. Four types of mould cheese (Lazur Blue, Président Brie Natural, Camembert Erival, and Gorgonzola Piccante) and two types of hard cheese (Salami, Mlekdamer) from various local shops in the eastern region of Poland were investigated. Each kind of cheese sample was divided into two equal portions and one part was stored at $4 \pm 2^{\circ}$ C for up to 112 (mould cheeses) and 133 days (hard cheeses) and the other was kept at room temperature of $22 \pm 2^{\circ}$ C for up to 42 days. After a suitable storage period, a piece of cheese (200 g) was homogenised and analysed. All cheeses were tested for histamine content immediately after delivery to the laboratory (day 0). Mould cheeses were then examined on days 14, 28, and 42 of storage at room temperature and on days 28, 56, 84, and 112 if stored at $4 \pm 2^{\circ}$ C. Hard cheeses were additionally tested at day 35 of storage at room temperature and on days 91 and 133 if stored at $4 \pm 2^{\circ}$ C. For each type of cheese the experiments were performed in triplicates.

Reagents. All reagents used were HPLC or analytical grade. Histamine dihydrochloride and 1-octanesulphonic acid were obtained from Acros Organics (Belgium). Trichloroacetic acid (TCA), potassium dihydrogen phosphate, orthophosphoric acid, and methanol were purchased from POCh (Poland). Polimeric weak anion solid phase extraction (SPE) cartridges (200 mg, 3 mL) were sourced from Phenomenex (USA) and nylon syringe filters (0.45 μ m) were procured from Agilent Technology (USA). Water was purified using the Milli-Q system (Millipore Sigma, USA).

Histamine stock standard solution (10 g/L) was prepared by dissolving of 828 ± 0.1 mg of histamine dihydrochloride in 50 mL of deionised water and storing the solution at 2–8°C for up to one month. Working standard solutions for the calibration curve were prepared on the day of analysis by appropriate dilutions of the stock in phosphate buffer (0.01 M, pH 3.0).

Extraction of histamine from cheese samples and clean-up. Cheese samples of weight 10 ± 0.01 g were weighed in a 50 mL polypropylene centrifuge tube. A total of 25 mL of TCA (0.2 M) was added and the sample was mixed by vortexing for 2 min, centrifuged for 10 min at 4,000 × g at 15°C, and kept at \leq -18°C for 30 min. The upper lipid layer was discarded and 5 mL of the supernatant was transferred into a 10 mL flask which was filled up with deionised water. The sample was filtered through a 0.45 µm nylon syringe filter into a glass tube and then cleaned up by SPE apparatus (J.T. Baker, USA) using polimeric weak anion SPE cartridges. The cartridges had previously been conditioned with 5 mL of methanol and then with 10 mL of phosphate buffer (0.01 M, pH 3.0). Seven millilitres of the extract was transferred to the SPE column and passed through the cartridge into a 10 mL flask. Afterwards, the column was washed with 2 mL of phosphate buffer. The eluate was supplemented to 10 mL with phosphate buffer.

HPLC-DAD conditions and validation parameters. The content of histamine was analysed by high performance liquid chromatography using the Pro Star system (Varian, Netherlands) equipped with a diode array detector, quaternary pump, autosampler, and column thermostat, and controlled by the Galaxie Workstation software (Varian). The chromatographic separation was performed on a Unisol C18 column of 150×4.6 mm, particle size 3 μ m, connected to a C18 precolumn of 10×3 mm (Agela Technologies, USA). The mobile phase, consisting of 15% methanol in 0.1 M potassium dihydrogen phosphate (150/850, v/v) with 1.6 mM 1-octanesulphonic acid, was applied under isocratic conditions. The column oven temperature was maintained at 25°C, the flow rate at 0.5 mL/min, and the injection volume was 20 µL. The UV detection was monitored at 215 nm.

The range of the method for hard cheeses was 4.25-420 mg/kg and the limits of detection and quantification were 3.52 mg/kg and 4.25 mg/kg respectively, whereas the range of the method for mould cheeses was 7.13-420 mg/kg and the limits of detection and quantification were 6.27 mg/kg and 7.13 mg/kg respectively. The repeatability (n = 6) and the reproducibility (n = 18) for hard cheeses were 3.6% and 4.6% respectively, whereas for mould cheeses these parameters were 1.9% and 3.0%. Recovery of the method ranged from 80.1% for hard cheeses to 82% for mould cheeses.

Results

Histamine content in mould cheeses. The histamine content in mould cheeses is presented in Figs 1 and 2 as the mean \pm SD of three replicates. The results show changes in the amine formation during storage at different temperatures and time. The highest amount of histamine (730.47 \pm 20.01 mg/kg) was identified in Gorgonzola Piccante cheese stored for 42 days at room temperature (Fig. 1). In Camembert Erival cheese kept at 4°C, histamine was detected at a high level (405.21 \pm 30.06 mg/kg) at the end of the storage period (112 days). In Gorgonzola Piccante cheese stored at 22°C and in Camembert Erival kept at 4°C, an

increase in the amount of histamine was observed with the extended period of storage. Besides the Italian Gorgonzola Piccante and French Camembert Erival cheeses, other ambient-stored cheeses showed a slight increase in histamine content up to 40.49 ± 18.29 mg/kg

(Président Brie Natural cheese) and 10.85 ± 3.97 mg/kg (Lazur Blue cheese) after 42 days of storage. With storage over 42 days at 22°C, the average concentration of histamine was more than twofold higher than that found in cheeses stored at 4°C for 112 days.



Fig. 1. Concentration of histamine in mould cheeses during storage at $22 \pm 2^{\circ}$ C. Each bar represents results as mean \pm SD



Fig. 2. Concentration of histamine in mould cheeses during storage at 4 \pm 2°C. Each bar represents results as mean \pm SD



Fig. 3. Concentration of histamine in hard cheeses during storage at $22 \pm 2^{\circ}$ C. Each bar represents results as mean \pm SD of three replicates



Fig. 4. Concentration of histamine in hard cheeses during storage at $4 \pm 2^{\circ}$ C. Each bar represents results as mean \pm SD of three replicates

Histamine content in hard cheeses. The analysis of histamine presence in hard cheeses revealed that it was highest in Mlekdamer cheese after 35 days of storage at room temperature ($35.84 \pm 4.12 \text{ mg/kg}$) (Fig. 3). Histamine concentration in this cheese ranged from 10 to 15 mg/kg during the 91 days of storage at 4 ± 2 °C and decreased after 112 days (Fig. 4). In Salami cheese the amine was found only in small amounts in samples stored at room temperature, and in samples stored at 4 ± 2 °C it was below the limit of detection of 3.52 mg/kg. Storage over 42 days at 22°C resulted in the average concentration of histamine being more than twofold higher than the level found after storage at 4°C for 133 days.

Discussion

Histamine content in Gorgonzola cheese at the beginning of the study (day 0) was 162.63 ± 23.57 mg/kg, which may suggest improper storage conditions during ripening or manufacture of the cheese from raw material with a significant amount of histamine. The ripening and storage periods are considered one of the key factors influencing histamine production (4, 8, 11, 32). During that time, a marked change in texture and flavour occur which is mainly associated with enzymatic degradation of casein resulting in steady increase in free amino acids. Some of the amino acids are subjected to a further breakdown reaction as decarboxylation catalysed by bacterial decarboxylases, which gives rise to the formation of carbon dioxide and histamine (11). Thus, cheese could be considered an ideal environment for this amine production through bacterial decarboxylation of histidine.

Given that the toxic threshold dose for histamine in cheeses has been temporarily established at 400 mg/kg (29), it was found that the amine content exceeded this value in Gorgonzola cheese after 28 and 42 days of storage at 22°C, where its levels reached 400.29 \pm 10.13 and 730.47 \pm 20.01 mg/kg, respectively, and reached this toxicity level in Camembert cheese on day 112 stored at 4°C (405.21 \pm 30.06 mg/kg). The wide variability of

histamine concentration in different cheeses may depend on type of cheese, ripening time, conditions of manufacturing process, and bacterial starter culture used (16, 20, 21). In the present study less histamine was detected in hard cheeses than soft ones and its amount was low (in a range from below the limit of detection to $35.84 \pm 4.12 \text{ mg/kg}$). This may be caused by soft cheeses' additional active microflora content (for example moulds) which is very active in transforming histidine to histamine. In none of the tested samples among hard cheeses was histamine concentration found to exceed the limit proposed by Rauscher-Gaberning *et al.* (29), which may indicate that these cheeses were safe for consumers.

The total histamine content in cheeses kept at 22°C was twofold higher than that kept at 4°C. This result may support the opinion that storage temperature has a great influence on histamine concentrations, and the production of this amine is promoted at room temperature. This finding may be linked to the effect of temperature of 22 ± 2 °C on some factors which influenced the biogenic amines formation such as microbial growth and enzymatic activity (4, 11).

The results of many studies have shown that the level of biogenic amines, including histamine, increased with storage time but also significantly depended on the temperature (1, 2, 12, 22). The histamine content was studied in traditional Czech curd cheese (Olomoucké tvarůžky) stored several weeks at different temperatures (5°C and 20°C). This amine was detected in amounts of 74-411 mg/kg and its level increased throughout the storage period and at 20°C as compared to 5°C (32). El-Kosi et al. (11) studied the effect of time (7 to 120 days) and temperature (7°C, 22°C, and 32°C) on the content of histamine in traditional Egyptian cheese varieties. Their results confirmed the theory of the stimulating effects of long storage time and higher temperatures on histamine level. The effects of storage conditions on histamine content were also tested in other Egyptian cheeses and in them likewise the accumulation of histamine increased remarkably in the ripening and storage periods (13). The growth of biogenic amines during production and storage may also occur in

unripened cheeses. In 12-day, 25° C-stored Mexican queso chihuahua white cheese an increased content of biogenic amines was observed (histamine up to 452 mg/kg) (28). Budak *et al.* (4) noted that histamine concentration reached the level of 265.5 mg/kg after 90 days' ripening at 10°C and it was detected in an amount higher than those of other biogenic amines in the Turkish white cheese samples investigated.

The results of the current investigation showed that histamine level did not always increase linearly during storage of cheeses. This finding may indicate that the formation of histamine in cheese depends on complex factors accelerating or retarding the production of this amine such as different numbers of decarboxylating microorganisms in certain parts of the cheese and internal microenvironmental conditions (access to O_2 or effects caused by water) (18, 22).

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