REVIEW ARTICLE

Molecular advances on agricultural crop improvement to meet current cultivating demands

T. Margaritopoulou^{1,*} and D. Milioni²

Abstract Sunflower, maize and potato are among the world's principal crops. In order to improve various traits, these crops have been genetically engineered to a great extent. Even though molecular markers for simple traits such as, fertility, herbicide tolerance or specific pathogen resistance have been successfully used in marker-assisted breeding programs for years, agronomical important complex quantitative traits like yield, biotic and abiotic stress resistance and seed quality content are challenging and require whole genome approaches. Collections of genetic resources for these crops are conserved worldwide and represent valuable resources to study complex traits. Nowadays technological advances and the availability of genome sequence have made novel approaches on the whole genome level possible. Molecular breeding, including both transgenic approach and marker-assisted breeding have facilitated the production of large amounts of markers for high density maps and allowed genome-wide association studies and genomic selection in sunflower, maize and potato. Marker-assisted selection related to hybrid performance has shown that genomic selection is a successful approach to address complex quantitative traits and to facilitate speeding up breeding programs in these crops in the future.

Additional keywords: Crop improvement, agricultural biotechnology, marker assisted selection, improved agronomic traits

Introduction

Agriculture is a human invention since more than 10,000 years and is estimated to have used more than 7,000 species to satisfy basic human needs (Esquinas-Alcázar, 2005). The primitive crop cultivars, known as landraces, were adapted to local growing conditions and practices, and therefore remained genetically diverse for traits such as product qualities, stress tolerance, disease resistance, and yield stability. Today's agricultural commodities and modern varieties derived from the genetic modification of wild plants through thousands of years of gradual selection, domestication and breed-

ing, are more genetically uniform than their wild relatives (Fu, 2015). Given that plant genetic diversity increases options for innovative, plant-based solutions to major environmental challenges such as water scarcity, deforestation, energy and climate change, molecular plant breeding can be a valuable tool to meet these demands by rapid incorporation of important traits from wild relatives into established crops and by shortening new crop domestication time (da Silva Dias, 2015).

Nowadays affordable high throughput DNA sequencing, coupled with improved bio-informatics and statistical analyses, is bringing major advances in the field of molecular plant breeding. Multidisciplinary breeding programs on the world's major crop plants are able to investigate genome-wide variations in DNA sequences and link them to inherited highly complex traits which are controlled by several genes, such as hybrid vigor and flowering. Furthermore, there has been

¹ Benaki Phytopathological Institute, Department of Phytopathology, Laboratory of Mycology, St. Delta 8, GR-145 61 Kifisia, Attica, Greece.

² Agricultural University of Athens, Department of Biotechnology, Iera Odos 75, GR-118 55 Votanikos, Athens, Greece.

 $[\]hbox{* Corresponding author: th.margaritopoulou@bpi.gr}$

a step-change in speed and cost-effectiveness (Robinson *et al.,* 2014). The availability of dense genetic maps can facilitate researchers to perform flexible marker-trait associations, concerning the correlations between pathogen resistance and alternative genes, and develop high performance markers that will promote marker- assisted choice (MAS) selection for resistant populations in segregating breeding programs (Ben-Ari and Lavi, 2012).

Herein, the molecular advances on agricultural crop improvement to meet current cultivating demands are reviewed for three economically important crops worldwide, i.e. sunflower, maize, potato.

Sunflower (*Helianthus annuus* L., Asteraceae)

Sunflower is the foremost seed crop cultivated within the world (Fernández-Luqueño et al., 2014). Sunflower oil contains less than 11% total saturated fat and does not contain any trans fat. Inexpensive production of biofuel from sunflower oil has been achieved (Boumesbah et al., 2015). Furthermore, sunflower is an ideal plant for producing high quality rubber from its leaves and stems and some of the taller perennial species have high latex yield potential (Lu and Hoeft, 2009).

The multiple usages of sunflower products in food, feed, and industry are stimulating the discovery of new sources of biodiversity for sunflower molecular breeding programs in combination with the application of high throughput approaches and genetic manipulation. The primary objective for sunflower breeders it to increase the yield and agronomical performance of high oleic sunflower hybrids. To accomplish these goals, breeders need to address pathogens, pests, and environmental constraints that have the potential to drastically reduce yield where sunflowers are grown (Dimitrijevic and Horn, 2018).

Genomic resources

A rich and various germplasm assortment is the backbone of each crop improve-

ment program. Assessing genetic diversity within a genetic pool of novel breeding germplasm could make crop improvement more efficient by the directed accumulation of desired alleles (Darvishzadeh et al., 2010). Several bacterial artificial chromosome (BAC) libraries have been constructed for sunflower (Feng et al., 2006; Gentzbittel et al., 2002; Özdemir et al., 2004). The libraries are equivalent to approximately 8 haploid genomes of sunflower and provide a greater than 99% probability of obtaining a clone of interest and they have been employed for isolating and physical mapping of loci such as the FAD2-1 locus (Schuppert et al., 2006) or the fertility restorer Rf1 locus (Hamrit et al., 2008). In situ hybridization techniques involving Fluorescent In Situ Hybridization (FISH) and BAC-FISH have being optimized for diversity and biological process studies between species of the genus Helianthus and development of a physical helianthus map allowing a cross reference to the genetic map (Giordani et al., 2014).

Various EST sequencing programs have been carried out in sunflower, including the Compositae Genome Project, and other programs (Tamborindeguy et al., 2004) and (Ben et al., 2005). The Compositae Genome Program (http://compgenomics.ucdavis.edu/index.php) has developed and is utilizing a 2.6 million feature Affymetrix chip based on 87,000 unigenes from seven Helianthus spp. (Lai et al., 2012). Interesting associations have been detected between Expressed Sequence Tags (ESTs) and Quantitative Trait Loci (QTLs) for salt tolerance and for domestication traits (Lai et al., 2005). Until today, 94.33 % of HA412-HO ESTs are correctly mapped and 90,935 protein coding genes are predicted, excluding transposable elements (http://www.sunflowergenome. org). Extensive genotyping has been performed for vegetative and flower sunflower organs together with uncovering gene networks for oil metabolism and flowering time (Badouin et al., 2017; Renaut 2017).

Efficient breeding strategy development

Biotechnology has the potential to help

evoke the full potential of this valuable crop (Fig. 1).

Resistance to pathogens

MAS technology has been used in sunflower breeding for various disease resistance traits (Brahm and Friedt 2000). With the development of an array of molecular markers and a dense genetic map of the sunflower genome, MAS for both single genes and QTLs is now possible (Babu et al., 2004; Bowers et al., 2012). For example, biotechnology offers a variety of methods for managing white rot caused by Stromatinia cepivora (also known as Sclerotium cepivorum) (Schnabl et al., 2002), including defense activation, pathogen inhibition and detoxification (Lu, 2003). According to Hu et al. (2003), the enzyme oxalate oxidase can confer resistance against Sclerotinia sclerotiorum, (Lib.) de Bary which causes sclerotinia wilt (midstalk rot), in transgenic sunflower plants while according to Sawahel and Hagran (2006), overexpression of a human lysozyme gene in sunflower confers resistance to the pathogen. Recently, the quantitative nature of Sclerotinia resistance has been exploited and QTL analysis showed that different genomic regions may contribute to resistance in different tissues of the plant (Würschum et al., 2014).

Alternative transgenic methods have been developed to reinforce sunflower resistance to diseases. A number of homologues resistance (R) gene have been isolated from sunflower, providing a valuable resource for engineering disease resistance in sunflower (Dimitrijevic and Horn 2018; Hewezi et al., 2006; Qi et al., 2016; Talukder et al., 2016).

Quality traits. Sunflower with high oleic acid content is optimal for the biodiesel industry since the produced oil has up to 90% mono-unsaturated fatty acid concentration, which has high oxidative stability and uniformity. Therefore, producing high concentrations of industrially valuable fatty acids in plant seeds through biotechnological improvements along with modifications of the fatty acid composition can make vegetable oil more versatile for its use (Burton et al., 2004).

One of the challenges for oil composition modification in sunflower is increasing the extent of the new fatty acids. Much work has been performed for the identification of genes involved in primary metabolic pathways and signal transduction at various growth and stress conditions (Liang et al., 2017; Pan et al., 2016; Velasco et al., 2014) to gain insight into the mechanism of antioxidant defense. New genes have been identified and the metabolism of ROS and RNS have been analyzed under various biot-

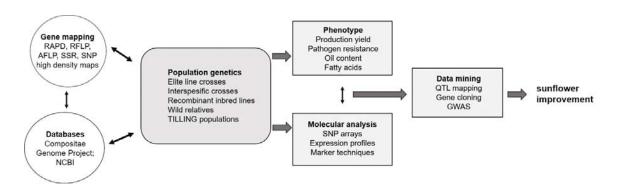


Fig. 1. Schematic depiction of the available resources in sunflower for marker-assisted selection and future genomic selection. Sunflower diverse genetic information is available for breeding and represents a large portion of genetic diversity that can be exploited for improving sunflower traits. Accessing sunflower genome sequences, large resources of SNP or high resolution maps and/or SNP arrays, along with huge amount of expression data can accelerate sunflower breeding by making the selection steps more efficient and precise. Marker-assisted breeding toward genomic selection can produce high quality breeding values.

ic and abiotic conditions (Chaki et al., 2013; Chaki et al., 2008; Chaki et al., 2011).

Overall, transgenic sunflower has the potential to meet the demands for yield improvement, to increase the efficient use of renewable resources, such as land, water and soil nutrients, and to significantly benefit everyday life by providing additional nutritive and healthy foods and valuable industrial products.

Ease of use and robustness of molecular markers

Markers' validation assesses their linkage to and association with QTLs and their effectiveness in selection of the target phenotype in independent populations and different genetic backgrounds (Collard *et al.,* 2005). An overall QTL mapping has been performed using microsatellite and Single Nucleotide Polymorphisms (SNP) markers in sunflower giving the ability to assess the genetic diversity and population structure across different sunflower populations (Filippi *et al.,* 2015).

Validation of genomic Simple Sequence Repeats (SSRs) in four genotypes of sunflower (RHA266, PAC2, HA89 and RHA801) resulted in amplification of 74 sequences from a total of 127 analyzed. Out of them, 13% represented polymorphic loci, 45% monomorphic, 5% null alleles and the remaining 37% showed either no amplification product, nonspecific amplification or complex or difficult to resolve banding patterns (Talia et al., 2010). The percentage of polymorphisms within sunflower that can be genetically mapped using SSR markers is shown to be less than 10% that comes in agreement with reports from other species (Varshney et al., 2005).

Examples of markers/QTLs validation across various genetic backgrounds in sun-flower include:

 A set of markers have been validated in a number of different genetic backgrounds for the Or5 gene conferring resistance to race E of the parasitic weed broomrape (Orobancche cumana), infecting the sunflower roots (Höniges et

- al., 2008; Pérez-Vich et al., 2004; Tang and Knapp, 2003).
- Markers have been validated for the dominant PI genes determining resistance to different downy mildew races (Brahm and Friedt 2000; Hvarleva et al., 2009; Ma et al., 2017) and to the R1, Radv and Pu6 genes conferring resistance to rust (Bulos et al., 2014).
- QTLs controlling three resistant (stem lesion, leaf lesion and speed of fungal control) and two morphological (leaf length and leaf length with petiole) traits have been validated for S. sclerotiorum across generations (Micic et al., 2005) and across environments (Talukder et al., 2016).
- QTLs have been validated for sunflower oil content, across generations, environments and mapping populations (Tang et al., 2006b).
- Markers have been developed in sunflower for simple traits selection, based on gene mutations underlying the trait of interest. There has been identified a mutation in codon 205 in the acetohydroxyacid synthase gene AHAs-1 that confers resistance to imidazolinone (IMI) herbicides and developed a SNP genotyping assay diagnostic for it (Kolkman et al., 2004).

Maize (Zea mays L., Poaceae)

Cultivation of maize is extensively widespread throughout the world and is surpassing any other grains (Council, 2019). With a fraction of total maize production being consumed by humans, its main products are ethanol, animal feed and processed corn starch and corn syrup (Klopfenstein *et al.*, 2013). Maize has high nutritional value but also is a fine source of various major phytochemicals such as carotenoids, phenolic compounds, and phytosterol, depicting its potential health benefits (Rouf Shah *et al.*, 2016).

Genome as the core base

B73 decoding. The 2.3-billion-base genome of an inbred line of maize called B73, an important commercial crop variety has been decoded (Schnable *et al.*, 2009). It has

been reported that the Palomero genome, a corn variety diverged from B73 about 9,000 years ago, is around 400 million nucleotides smaller and contains about 20% less repetitive DNA than B732 (Vielle-Calzada et al., 2009). To map maize haplotypes a part of the gene-rich region of 27 maize varieties was sequenced. 'HapMap' revealed thousands of genes around the centres of the chromosomes, where they were unlikely to be shuffled around during recombination (Gore et al., 2009). Schnable et al. (2011) demonstrated that the maize subgenomes are differentiated by genome dominance and both ancient and ongoing gene loss. Most of the economically important traits considered in maize breeding are inherited quantitatively. Multiple genes or quantitative trait loci (QTLs) affecting flowering traits, root characteristics, cell wall traits, and tolerance to biotic/abiotic stresses panicle morphology and grain development have been cloned, and gene expression research has provided new information about the nature of complex genetic networks involved in the expression of these traits (Buckler et al., 2009; Chung et al., 2011; Fernandez et al., 2009; Messmer et al., 2009; Poland et al., 2011; Trachsel et al., 2009). A meta-analysis of QTL associated with plant digestibility and cell wall composition in maize identified key chromosomal regions involved in silage quality and potentially associated genes for most of these regions (Truntzler et al., 2010).

Association mapping (associating specific DNA polymorphisms with traits of interest based on linkage disequilibrium). McMullen et al. (2009) described the maize NAM population generated by crossing 25 diverse inbred lines to a common line, inbred B73. Sequenome-based SNP-typing assay was used to identify 1,359 SNPs in maize transcriptome and 75% of these SNPs were confirmed and applied in association analysis (Liu et al., 2010). Currently, there are over 2 million maize ESTs in GenBank (Benson et al., 2009). However, the assembly of these ESTs into gene models presents practical problems. Therefore, a full length cDNA library has been recently constructed for *Zea mays*

(http:www.maizecdna.org/) (Soderlund *et al.*, 2009). A normalized cDNA library, covering most of the developmental stages of maize seeds, was also constructed and 57 putative transcription factors were identified (Wang *et al.*, 2010). The cDNA libraries can serve as primary resources for designing microarray probes and as clone resources for genetic engineering to improve crop efficiency.

Maize GDB (http://www.maizegdb.org/). Maize GDB is a database that provides documentation and data for the microarrays produced by the Maize Gene Discovery Project. An extensive expression atlas covering a wide array of tissues and developmental stages of maize using a NimbleGen microarray encompassing 80 301 probe sets was recently constructed (Sekhon et al., 2011). Random-sheared, paired-end Illumina GAII reads have been generated from 103 maize, teosinte and maize landrace inbred lines at a depth ranging from 4-30x (Chia et al., 2012; Hufford et al., 2012). Microarray studies have also been performed to study cell wall metabolism in maize, with the aim of identifying tissue-specific or developmentally regulated gene expression of members of multigene families or to obtain a better understanding of regulatory networks that are exposed when cell wall-related genes are mutated (Guillaumie et al., 2007a; Guillaumie et al., 2007b). The MAIZEWALL sequence database and expression profiling resource has been developed (www.polebio.scsv.upstlse.fr/MAIZEWALL). Rajhi and co-workers performed transcriptome analysis in maize root cortical cells during lysigenous aerenchyma formation and discovered a number of genes whose expression changed in response to ethylene under waterlogged conditions (Rajhi et al., 2011).

Maize small RNAs. Small RNAs in the wild type and in the isogenic Mediator Of Paramutation1 loss-of- function (mop1-1) mutant have been examined by deep sequencing to analyze the size distribution of maize small RNAs (Nobuta et al., 2008). Small RNAs are playing roles as major components of epigenetic processes and gene networks

involved in development and homeostasis. It has been recently demonstrated that a change in expression of a key component of the RNA silencing pathway is associated with both vegetative phase change and shifts in epigenetic regulation of a maize transposon (Li *et al.*, 2010).

RNA interference (RNAi) [RNA-mediated gene silencing by sequence-specific degradation of homologous mRNA triggered by double-stranded RNA (dsRNA)]. The RNAi system was used to improve resistance to maize dwarf mosaic virus on transgenic maize (Zhang et al., 2011). Maize lines expressing RNAi to chromatin remodeling factors were shown to be similarly hypersensitive to UV-B radiation but exhibit distinct transcriptome responses (Casati and Walbot 2008). By using near infrared reflectance spectroscopy (NIRS), a set of 39 maize mutants with altered spectral phenotypes ('spectrotypes') have been identified (Vermerris et al., 2007). A number of these mutants were shown to have altered lignin-to-carbohydrate ratios (Penning et al., 2009). Sequence- specific DNA binding Transcription Factors (TFs) are key molecular switches that control or influence many biological processes, such as development or response to environmental changes. The Maize Transcription Factor Database provides a comprehensive collection of 764 predicted transcription factors from maize with available links to information on mutants, map positions or putative functions for these transcription factors (MaizeTFDB) (http://grassius.org/browsefamily.html?species=Maize). Information resources related to metabolomics can play major role not only in metabolomics research but also in synergistic integration with other omics data. MaizeCYc is a biochemical pathway database that provides manually curated or reviewed information about metabolic pathways in maize.

Molecular breeding for current needs

Molecular breeding, including both transgenic approach and marker-assisted breeding, is primary associated with the challenges for developing cultivars with combinations of adaptive traits (Brown et al., 2011; Varshney et al., 2011). For making molecular marker-assisted breeding successful, marker-trait associations are now known for almost all important economic traits, including thousands of mapped microsatellite or SSR markers, and additional recently, SNPs, and insertion-deletion (InDel) markers. For maize, there is an updated compilation of mapped QTL for abiotic stress resistance (http://www.plantstress.com; http:// www.maizegdb.org; http://www.gramene. org). Additionally, a large number of genes controlling various aspects of plant development, biotic and abiotic stress resistance, quality characters, etc. have been cloned and characterized in maize, which are excellent assets for molecular marker- assisted breeding (Aslam and Ali 2018; Prasanna et al., 2010).

Tolerance against drought. Since drought is considered to be the most important constraint across all areas where maize is cultivated, and global warming is predicted to further exacerbate drought's impact, a total management plan is necessary for increasing maize yield in stress-prone environments (Fig. 2). The high variability to drought stress and also the uncontrollable fact that drought response has great fluctuations across environments, have made it difficult to spot specific metabolic pathways which limits breeding efforts towards drought tolerance (Collins et al., 2008). A Marker-Assisted BackCross (MABC) selection approach meant for improving grain yield under water limited conditions in tropical maize, was successfully conducted at CIMMYT (Ribaut and Ragot 2006) and more recently at sub-Saharan Africa (Beyene et al., 2016). However, this approach delivers a restricted level of improvement in drought tolerance since it provides an improved version of an existing genotype (Ribaut et al., 2009). Nevertheless, a molecular breeding approach-marker-assisted recurrent selection (MARS) can be used to overcome this problem. MARS studies exploit association mapping and can effectively double the rate of yield gain compared to conventional

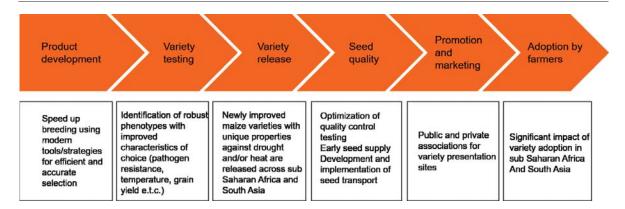


Fig. 2. Schematic representation that highlights the required key steps to facilitate enhanced adoption and impacts of improved climate-resilient maize varieties in the developing world. Increasing maize yields in stress-prone environments and reducing year-to-year variability is an important step in improving food safety, livelihoods and adaptation to the changing climate in the developing world (Cairns and Prasana, 2018).

breeding in elite germplasms when favored and stress environments are been examined (Crosbie et al., 2006; Eathington et al., 2007; Edgerton 2009). Most recently, the role of Abscisic Acid (ABA) pathway in drought resistance has been investigated and natural variants of ABA-(PYR1/PYL/RCAR) protein (PYL) receptors have been identified that can serve as potential molecular markers for breeding drought-resistant maize cultivars (He et al., 2018).

Resistance against pathogens. Efforts to scale down maize losses from pathogen attacks through resistant crop varieties could provide tremendous opportunities for increasing and stabilizing maize productivity. QTL related to resistance to several diseases, such as downy mildew and rust, and insect-pests are known and mapped in maize, creating marker assisted choice as a potentially viable strategy to improve resistance to these biotic stresses (Ali and Yan 2012; García-Lara et al., 2009; Krakowsky et al., 2004; Wisser et al., 2006).

Resistance against insect pests. The industry has made substantial progress with insect resistant maize through transformation with insecticidal proteins from *Bacillus thuringiensis* (Bt) which have been particularly successful in providing protection against several corn borers (Glaser and Matten 2003; Jiang *et al.*, 2018).

Quality traits. Quality traits, like oil con-

tent or high nutritional value molecules, have induced a shift in maize production far from strictly an identity-preserved cultivation to more a value-added product. The capability of changing cell membrane polysaccharides into possible sugars for grain ethanol production depends on cell membrane structure. Molecular markers can be a valuable tool when breeding for feed maize but with improved quality on grain ethanol.

QTLs with comparatively efficient results are found for feed maize including cell membrane composition and glucose release (GL-CRel) (Lorenzana et al., 2010), and some important constitutive and adaptive QTLs are identified by using meta-analysis (Hao et al., 2010). (Torres et al., 2015) presented the molecular progress that has been made in altering maize's cellulosic content in order to exploit useful biomass characteristics and design new breeding strategies.

Quality traits and tolerance to abiotic stress. There has been increasing interest in addressing advanced traits like grain quality and abiotic/biotic stress tolerances through recombinant DNA technology. Elite inbred South African transgenic corn plants were modified in 3 separate metabolic pathways to produce increased quantities of vitamin β-carotene, ascorbate and folate (Naqvi et al., 2009). It has been demonstrated that engineering of the alkaloid synthesis pathway could have great impact on im-

proving cold tolerance in maize (Quan et al., 2004). Furthermore, genome-wide association analyses (GWAS) in temperate maize inbred lines is serving as a tool to find strategies for identifying genes for cold tolerance (Revilla et al., 2016) and has been reported that the introduction of an antisense gene for pyruvate orthophosphate dikinase (PPDK) into maize with Agrobacterium-mediated transformation resulted in shifting the break point 3°C less than that of the wild type (Ohta et al., 2004).

Drought is another stress factor that has been addressed in maize improvement. Nuclear Factor-Y (NF-Y) is a 3- subunit complex that has been shown to play major role in growth, development, and response to environmental stress. Except studies that have been performed for characterizing NF-Y gene families in maize (Zhang et al., 2016), when ZmNF-YB2 or ZmNF-YB16 were constitutively expressed in elite maize inbred lines, the transgenic lines displayed improved drought tolerance compared to wild-type plants under water-stressed conditions in the field (Nelson et al., 2007; Wang et al., 2018). (Castiglioni et al., 2008) demonstrated that transgenic maize lines recombinant with bacterial RNA chaperones resulted in not only abiotic stress tolerance but also improved grain yield under waterlimited conditions. The application of this technology has the potential to considerably impact maize production systems that have drought. However, commercialization of transgenic maize for abiotic stresses like drought tolerance has been terribly restricted (Xu et al., 2009).

Moreover, the past ten years we have witnessed extensive efforts toward the development of an efficient *Agrobacterium*-mediated transformation system for an array of maize developing organs with particular emphasis on increasing the efficiency and extending the range of amenable genotypes (Cao *et al.*, 2014; Lee and Zhang 2014; Shrawat and Lörz, 2006).

Validation of quantitative traits

In maize, a trait that has been exten-

sively investigated as an indirect measure of drought tolerance is the capacity of ABA accumulation. The presence of a major QTL for root features (root-ABA1) was mapped on bin 2.04 in Os420 \times IABO78. This major QTL affecting abscisic acid (ABA) concentration in the leaf, root traits and relative water content was further evaluated in maize using NILs (Landi et al., 2005). Interestingly, the QTL allele for larger root mass and higher ABA concentration negatively affected grain yield (Landi et al., 2006). Laurie et al. (2004) were able to detect 50 QTL accounting for genetic variance in maize oil content with a resolution of the order of a few centimorgans across generations.

QTL conditioning resistance to plant pathogens (rQTL) have been discovered and reviewed by several authors (Balint-Kurti and Johal, 2009; Redinbaugh and Pratt, 2009). To date only a few QTL conferring resistance to maize streak mastrevirus, Cercospora zeae-maydis, Exserohilum turcicum (Pass.) and Peronosclerospora sorghiin have been validated (Abalo et al., 2009; Asea et al., 2009; Nair et al., 2005). For Cercospora resistance in maize, QTLs have been validated across genetic backgrounds (Pozar et al., 2009) and environments (Juliatti et al., 2009). Furthermore, a major QTL controlling maize streak virus resistance explains 50-70% of total phenotypic variation (Pernet et al., 1999). Several microsatellite markers associated with this QTL were validated across populations and have been successfully used for the selection of resistant lines (William et al., 2007).

Analyses for evaluating the significance of QTL x genetic background interactions in several diverse mapping populations, have been performed in maize for grain moisture, silking date and grain yield (Blanc et al., 2006; Huo et al., 2016). QTL meta-analysis is another approach to identify consensus QTL across studies, to validate QTL effects across environments/genetic backgrounds, and also to refine QTL positions on the consensus map (Goffinet and Gerber 2000). The concept of meta-analysis has been applied to the analysis of QTL/genes for flowering

time (Chardon *et al.*, 2004) and drought tolerance in maize (Hao *et al.*, 2010). A meta-analysis of QTL associated with plant digestibility and cell wall composition in maize has been carried out and fifteen meta QTL with confidence interval (CI) smaller than 10cM were identified (Truntzler *et al.*, 2010).

Potato (*Solanum tuberosum*, L., Solanaceae)

Cultivated potato is the world's third most important human food crop (www. cipotato.org). It is also used as raw material for starch and alcohol production (Cantos-Lopes *et al.*, 2018). The basic chromosome number for potato species is 12. Even though one of the most widespread food crop around the world, the genetics of many potato traits is poorly understood.

Insights in genomic properties

An ultrahigh-density (UHD) genetic map composed of approximately 10,000 Amplified Fragment Length Polymorphism (AFLP) markers has been developed, which is most likely the densest map for a plant species ever constructed (Van Os et al., 2006). Recently, the relationship between the genetic and chromosome map in potato was displayed and two linkage maps were integrated with potato genome sequence developing 8303 Single Nucleotide Polymorphism (SNP) for genome-guided breeding (Felcher et al., 2012). Moreover, (Sharma et al., 2013) elaborated 2469 marker loci in a linkage map which was integrated with potato reference genome (DM) and other physical and genetic maps of potato providing detailed information about chromosomal gene distribution. Using RFLP and AFLP markers, a QTL and linkage map of two segregating diploid populations previously evaluated for sugar content after cold storage, was generated. Ten potato genes with unknown function in carbon metabolism or transport were mapped and tested for their effects on sugar content. Results displayed linkage between glucose, fructose and sucrose QTLs and all of eight candidate gene loci (AGPaseS, AGPaseB, Sbel,

GapC, Invap, Ppa1, Sut1, Sut2) (Menéndez et al., 2002). Several QTLs affecting the ability to form tubers under long photoperiods (earliness) have been identified (šimko et al., 1999). A functional map for pathogen resistance, enriched with RGA (resistance gene analog) and DRL (defence related locus) sequences, SNPs and insertion-deletion polymorphisms (InDels) tightly linked or located within Nucleotide Binding Site - Leucine Rich Repeat (NBS-LRR) -like genes, has been developed on the basis of two potato populations (BC9162 and F1840) (Rickert et al., 2003; Trognitz et al., 2002). Recently, twenty-one QTL and eight reference published potato maps were merged together and the first consensus map was built. Individual QTLs for resistance to the late blight pathogen, Phytophthora infestans (Mont.) de Bary, and maturity traits were projected onto the consensus map and the first meta-analysis performed deals with both development trait and resistance to a biotic stress in potato (Danan et al., 2011).

As a major follow-up, the genome of potato (850 Mb) was sequenced by the international Potato Genome Sequencing Consortium (PGSC), which was comprised by 13 countries [http://www.potatogenome.net/]. The new genome sequence data provides information about extensive copy number variation (CNV) which has great impact on 219.8 Mb (30.2%) of the potato genome. Almost 30% of genes are subjected to at least partial duplication or deletion which reveals the highly heterogeneous nature of the potato genome (Hardigan et al., 2016). Comparative sequence analysis of Solanum and Arabidopsis in a hot spot for pathogen resistance on potato chromosome V has also been performed and revealed a patchwork of conserved and rapidly evolving genome segments (Ballvora et al., 2007).

Several efforts to generate EST resources for potato have been performed (Flinn *et al.*, 2005). Potato cDNA microarray analysis was performed to assess the potential of transcriptomics to detect differences in gene expression due to genetic differences or environmental conditions (van Dijk *et al.*, 2009). A

cDNA- AFLP approach and bulked segregate analysis (BSA) was used to identify genes cosegregating with earliness of tuberization in a diploid potato population. 81 candidate polymorphic transcript-derived fragments (TDFs) showing polymorphism between the early and late bulks were selected for further analysis (Fernández-del-Carmen et al., 2007). Genetic engineering could enhance desirable characteristics of crops by modifying key regulatory steps for entire metabolic or developmental pathways. The optimal conditions for genetic transformation of Solanum spp mediated by Agrobacterium tumefaciens have been established (Chakravarty et al., 2007). It has been demonstrated that transgenic katahdin plants containing the RB gene showed resistance to all tested Pythophtora isolates, including a super race that can overcome all eleven known R genes in potato. An RNA interference (RNAi)based potato gene silencing approach using agroinfiltration, has been recently established (Bhaskar et al., 2009).

How to design efficient breeding strategies

Tolerance to salt stress. Potato crop production is highly inversely connected to salt stress with substantial economic impacts (Katerji et al., 2000). When potato is subjected to salt stress, increased activation of antioxidant enzymes, accumulation of proline, decrease in micro tubers and negative effects on physiological characteristics occur (Rahnama and Ebrahimzadeh 2004; Tang et al., 2006a; Zhang et al., 2005). Gene expression studies on potato cultivars under different stress conditions, such as cold, heat or salt, revealed that transcription factors, signal transduction factors and heat shock protein (HSP) are associated with abiotic stress responses (Rensink et al., 2005; Tang et al., 2016). In addition, when Δ -pyrroline-5carboxylase synthetase, which is involved in proline production, is overexpressed, it confers salt tolerance to potato (Hmida-Sayari et al., 2005).

Aghaei et al. (2008) examined closely in a protein level the differences between a salt

tolerant and a salt sensitive potato culture. They pointed out that among the proteins that were differentially expressed photosynthesis- and protein synthesis-related proteins were drastically down-regulated, whereas osmotine-like proteins, type VI secretion immunity protein (TSI-1), heat-shock proteins, protein inhibitors, calreticulin, and five novel proteins were remarkedly up-regulated. Under salt conditions, major changes occur within the photosytem protein machinery and the Calvin cycle as demonstrated by an in-depth cDNA microarray map constructed from potato leaves (Legay et al., 2009).

More recently, advances have been made in identifying several genes that play key roles to biotic and abiotic stress responses. A pathogen-related protein, named PR-10a, has been identified which is not only induced under biotic stress conditions in potato, but also exhibits significantly increased tolerance under salt and osmosis conditions (El-Banna et al., 2010). Two different studies showed that the metal zinc finger protein St ZFP1 could participate to salt associated potato responses through the ABA- dependent pathway (Tian et al., 2010) and also the cinnamyl alcohol dehydrogenase ibCAD1 may play a very important role in each abiotic and biotic stress resistance mechanisms (Kim et al., 2010).

Tolerance to drought. Another major abiotic stress issue that ends up in crop losses in potato cultivars, is drought. The development of drought tolerant cultivars is of primary importance for maintaining yields beneath temperature change conditions and for the extension of cultivation to sub-optimal cropping areas. Extensive cDNA microarray analysis showed that a tolerant accession to drought, named 397077.16, presented differentially expressed genes when compared to a sensitive variety (Legay et al., 2011). The genes belonged to groups of carbohydrate metabolism, cell protection and detoxification, meaning that the tolerant accession can respond more efficiently to stress and be more adaptive when compared to the sensitive one. Additionally, the work of other groups identified a transcription factor which is involved in the activation of drought related genes (Shin *et al.,* 2011) and showed the importance of the overexpression of the L-gulono-c-lactone oxidase (GLOase gene) gene to the resistance to various abiotic stress factors (Upadhyaya *et al.,* 2009).

Resistance to pathogens. The use of resistant varieties is taken into account to be the foremost appropriate approach for the management of Phytophthora infestans. Extensive examination of potato genotype SD20 revealed WRKY domain transcription factor (WRKY), single AP2/ERF domain transcription factor (ERF), MAP kinase (MAPK), and NBS-LRR gene families that play essential role in late blight (Yang et al., 2018). Moreover, it has been suggested that the R8 gene, found in field trials, is responsible for late blight resistance and that its mapping on the long arm of chromosome IX along with the generation of markers would be a helpful tool for marker assisted breeding (Jo et al., 2011). Nowadays, R8 gene is a worldwide tool for late blight resistance (Vossen et al., 2016). The introduction of simultaneously three resistance genes from three potato accessions to a sensitive cultivar (Zhu et al., 2012), the silencing of six S-genes in the susceptible potato cultivar Desiree (Sun et al., 2016) or the contribution of R-gene dosage and biochemical pathways to resistance (Gao and Bradeen 2016), are good examples in the literature, considering transformation techniques for late blight resistance. On the other hand, since potato late blight resistance has been thoroughly studied, an extensive map of QTLs and Rpi-genes (resistance genes to Phytophthora infestans) has been generated (Danan et al., 2011; Jiang et al., 2018; Stefańczyk et al., 2017).

Other efforts to increase potato resistance to pathogens include exploitation of inhibitor genes. (Khadeeva et al., 2009) showed that transformation of potato plants with an inhibitor gene of buckwheat provides protection to the plants against pathogens. Furthermore, a gene family that function against nematode infections have been sequenced and char-

acterized from *Solamun tuderosum* cv. Desiree (Turra *et al.*, 2009). Also, advances have been made in the identification of genes that are involved in the mechanisms controlling the arbuscular mychorrhizal establishment by the regulation of plant defense genes (Gallou *et al.*, 2012).

Molecular markers as a key tool for crop improvement

Tuber susceptibility to bruising. Diagnostic markers for tuber bruising and enzymatic discoloration, which are very crucial characteristics to crop quality of the cultivated potato, have been validated (Urbany et al., 2011). The markers diagnostic for increased or decreased bruising susceptibility is expected to facilitate the combination of superior alleles in breeding programs.

Potato germplasm (use of sources of resistance to pests and diseases in order to breed varieties cheaper to grow). Although the actual copy number of the genes is not known, DNA markers located close to genes that encode resistance or hypersensitive response to the Potato virus Y (PVY), which can reduce yield up to 80 percent while being relatively symptomless, have been identified and validated (Fulladolsa et al., 2015; Szajko et al., 2014; Tomczyńska et al., 2014). Furthermore, Cleaved Amplified Polymorphic Sequences (CAPs) and Sequence Characterized Amplified Regions (SCARs) have allowed the breeding of genotypes resistant to PVY (Kasai et al., 2000).

The successful employment of four PCR-based diagnostic assays to combine the Ry adg gene for extreme resistance to PVY with Gro1 for nematode resistance and with Rx1 for extreme resistance to potato virus X (PVX, genus Potexvirus), or with Sen1 for wart resistance (*Synchytrium endobioticum*) has been reported (Gebhardt *et al.*, 2006).

The availability of DNA-based markers, which are easy to score, cost-effective and diagnostic for resistance to Pathotypes 2/3 (Pa2/3) of the most significant soilborne pests of potato, the potato cyst nematode (*Globodera pallida*), would greatly speed up the process of new variety development. A

set of markers have been validated for QTL on linkage group IV (renamed GpalV adg s) across a wide range of germplasm (Moloney et al., 2010).

Field resistance to *Phytophthora infestans* has been characterized in a potato segregating family of 230 full-sub progenies derived from a cross between two hybrid *S. phure-ja x S. stenotomum* clones. QTLs have been identified and validated for the new genetic loci in this diploid potato family contributing to general resistance against late blight (Costanzo *et al.,* 2005).

Potato breeding widely exploits molecular techniques for generation and conservation of advanced clones, increasing the potato cultivar number every year (Fig. 3). Reliable maintenance of large culture collections is becoming more problematic and a rapid and robust method for variety differentiation is becoming highly desirable. The validation of a set of six SSRs markers that can be used

to differentiate over 400 potato cultivars has been reported (Reid and Kerr, 2007).

Prospects

Genomic research allows high-throughput analysis for crop improvement. Genetic markers designed to cover a genome extensively allow not only identification of individual genes associated with complex traits by quantitative trait loci analysis but also the exploration of genetic diversity with regard to natural variations.

Wild relatives are valuable knowledge that can upscale with valuable traits the crop species. Nowadays, only a little fraction is exploited for crop improvement. One of the basic issues of crop improvement is to access the genetic variation from such wild species. This is particularly important to the transfer of valuable, novel genes from wild

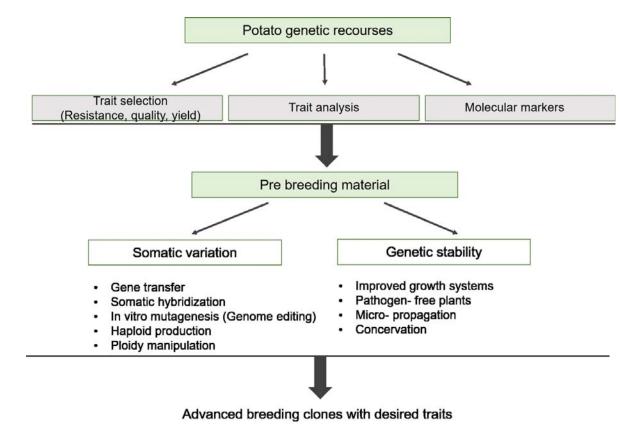


Fig. 3. Gene variants are a valuable tool for improving potato cultivars. Schematic overview of the individual sections that constitute the integrated management of potato genomic resources for the generation of elite breeding clones with improved agronomical traits of interest.

relatives to crops for non-food uses. Biotechnology offers the greatest potential in contributing solutions to problems that agriculture is facing now and the years to come.

This work was part of the Crops2Industry project that was funded by the Seventh (7th) Research Framework Program of the European Community.

The authors declare there is no conflict of interest.

Literature cited

- Abalo, G., Tongoona, P., Derera, J. and Edema, R. 2009. A comparative analysis of conventional and marker-assisted selection methods in breeding maize streak virus resistance in maize. *Crop Science*, 49: 509-520.
- Aghaei, K., Ehsanpour, A.A. and Komatsu, S. 2008. Proteome analysis of potato under salt stress. *Journal of Proteome Research*, 7: 4858-4868.
- Ali, F. and Yan, J. 2012. Disease resistance in maize and the role of molecular breeding in defending against global threat. *Journal of Integrative Plant Biology*, 54: 134-151.
- Asea, G., Vivek, B.S., Bigirwa, G., Lipps, P.E. and Pratt, R.C. 2009. Validation of consensus quantitative trait loci associated with resistance to multiple foliar pathogens of maize. *Phytopathology*, 99: 540-547.
- Aslam, F. and Ali, B. 2018. Halotolerant Bacterial Diversity Associated with *Suaeda fruticosa* (L.) Forssk. Improved Growth of Maize under Salinity. *Stress Agronomy*, 8: 131.
- Babu, R., Nair, S.K., Prasanna, B. and Gupta, H. 2004. Integrating marker-assisted selection in crop breeding-prospects and challenges. *Current Science*, 607-619.
- Badouin, H., Gouzy, J., Grassa, C.J., Murat, F., Staton, E.S., Cottret, L., Lelandais-Brière, C., Owens, G.L., Carrère, S., Mayjonade, B., Legrand, L., Gill, N., Kane, N.C., Bowers, J.E., Hubner, S., Bellec, A., Bérard, A., Bergès, H., Blanchet, N., Boniface, M.-C., Brunel, D., Catrice, O., Chaidir, N., Claudel, C., Donnadieu, C., Faraut, T., Fievet, G., Helmstetter, N., King, M., Knapp, S.J., Lai, Z., Le Paslier, M.-C., Lippi, Y., Lorenzon, L., Mandel, J. R., Marage, G., Marchand, G., Marquand, E., Bret-Mestries, E., Morien, E., Nambeesan, S., Nguyen, T., Pegot-Espagnet, P., Pouilly, N., Raftis, F., Sallet, E., Schiex, T., Thomas, J., Vandecasteele, C., Varès, D., Vear, F., Vautrin, S., Crespi, M., Mangin, B., Burke, J.M.,

- Salse, J., Muños, S., Vincourt, P., Rieseberg, L.H. and Langlade, N.B. 2017. The sunflower genome provides insights into oil metabolism, flowering and Asterid evolution. *Nature*, 546: 148.
- Balint-Kurti, P.J. and Johal, G.S. 2009. Maize disease resistance. In: *Handbook of maize: its biology*. Springer, pp 229-250.
- Ballvora, A., Jöcker, A., Viehöver, P., Ishihara, H., Paal, J., Meksem, K., Bruggmann, R., Schoof, H., Weisshaar, B. and Gebhardt, C. 2007. Comparative sequence analysis of *Solanum* and *Arabidopsis* in a hot spot for pathogen resistance on potato chromosome V reveals a patchwork of conserved and rapidly evolving genome segments. *BMC Genomics*, 8: 112.
- Ben-Ari, G. and Lavi, U. 2012. Marker-assisted selection in plant breeding. In: *Plant Biotechnology and Agriculture*. Elsevier, pp 163-184.
- Ben, C., Hewezi, T., Jardinaud, M.F., Bena, F., Ladouce N., Moretti, S., Tamborindeguy, C., Liboz, T., Petitprez, M. and Gentzbittel, L. 2005. Comparative analysis of early embryonic sunflower cDNA libraries. *Plant molecular biology*, 57: 255-270.
- Benson, D., Karsch-Mizrachi, I., Lipman, D., Ostell, J. and Wheeler, D. 2009. Database issue GenBank, *Nucleic Acids Research*, 37: 26-31.
- Beyene, Y., Semagn, K., Mugo, S., Prasanna, B.M., Tarekegne, A., Gakunga, J., Sehabiague, P., Meisel, B., Oikeh, S.O., Olsen, M. and Crossa, J. 2016. Performance and grain yield stability of maize populations developed using marker-assisted recurrent selection and pedigree selection procedures. *Euphytica*, 208: 285-297.
- Bhaskar, P.B., Venkateshwaran, M., Wu, L., Ané, J.-M. and Jiang, J. 2009. Agrobacterium-mediated transient gene expression and silencing: a rapid tool for functional gene assay in potato. *PLoS ONE*, 4: e5812.
- Blanc, G., Charcosset, A., Mangin, B., Gallais, A. and Moreau, L. 2006. Connected populations for detecting quantitative trait loci and testing for epistasis: an application in maize. *Theoretical and Applied Genetics*, 113: 206-224.
- Boumesbah, I., Hachaïchi-Sadouk, Z., and Ahmia, A.C. 2015. Biofuel Production from Sunflower Oil and Determination of Fuel Properties. *In: Progress in Clean Energy,* Volume 2. Springer, pp 105-111.
- Bowers, J.E., Nambeesan, S., Corbi, J., Barker, M.S., Rieseberg, L.H., Knapp, S.J. and Burke, J.M. 2012. Development of an ultra-dense genetic map of the sunflower genome based on single-feature polymorphisms. *PLoS ONE*, 7: e51360.
- Brahm, L. and Friedt, W. 2000. PCR-based markers facilitating marker assisted selection in sunflower for resistance to downy mildew. *Crop Science*, 40: 676-682.
- Brown, P.J., Upadyayula, N., Mahone, G.S., Tian, F., Bradbury, P.J., Myles, S., Holland, J.B., Flint-Gar-

- cia, S., McMullen, M.D., Buckler, E.S. and Rocheford, T.R. 2011. Distinct genetic architectures for male and female inflorescence traits of maize. *PLoS Genetics*, 7: e1002383.
- Buckler, E.S., Holland, J.B., Bradbury, P.J., Acharya, C.B., Brown, P.J., Browne, C., Ersoz, E., Flint-Garcia, S., Garcia, A., Glaubitz, J.C., Goodman, M.M., Harjes, C., Guill, K., Kroon, D.E., Larsson, S., Lepak, N.K., Li, H., Mitchell, S.E., Pressoir, G., Peiffer, J.A., Rosas, M.O., Rocheford, T.R., Romay, M.C., Romero, S., Salvo, S., Sanchez Villeda, H., da Silva, H.S., Sun, Q., Tian, F., Upadyayula, N., Ware, D., Yates, H., Yu, J., Zhang, Z., Kresovich, S. and McMullen, M.D. 2009. The genetic architecture of maize flowering time. *Science*, 325: 714-718.
- Bulos, M., Vergani, P.N. and Altieri, E. 2014. Genetic mapping, marker assisted selection and allelic relationships for the Pu6 gene conferring rust resistance in sunflower. *Breeding Science*, 64: 206-212.
- Burton, J.W., Miller, J.F., Vick, B., Scarth, R. and Holbrook, C.C. 2004. Altering fatty acid composition in oil seed crops. *Advances in Agronomy*, 84: 273-306.
- Cairns, J.E. and Prasanna, B.M. 2018. Developing and deploying climate-resilient maize varieties in the developing world. *Current Opinion in Plant Biology*, 45: 226-230.
- Cantos-Lopes, A., Vilela-de Resende, J.T., Machado, J., Perez-Guerra, E. and Vilela-Resende, N. 2018. Alcohol production from sweet potato (*Ipomoea batatas* (L.) Lam.) genotypes in fermentative medium. *Acta Agronómica*, 67: 231-237.
- Cao, S.-I., Masilamany, P., Li, W.-B. and Pauls, K.P. 2014. Agrobacterium tumefaciens -mediated transformation of corn (Zea mays L.) multiple shoots. Biotechnological Equipment, 28: 208-216.
- Casati, P. and Walbot, V. 2008. Maize lines expressing RNAi to chromatin remodeling factors are similarly hypersensitive to UV-B radiation but exhibit distinct transcriptome responses. *Epigenetics*, 3: 216-229.
- Castiglioni, P., Warner, D., Bensen, R.J., Anstrom, D.C., Harrison, J., Stoecker, M., Abad, M., Kumar, G., Salvador, S., D'Ordine, R., Navarro, S., Back, S., Fernandes, M., Targolli, J., Dasgupta, S., Bonin, C., Luethy, M.H. and Heard, J.E. 2008. Bacterial RNA chaperones confer abiotic stress tolerance in plants and improved grain yield in maize under water-limited conditions. *Plant Physiology*, 147: 446-455.
- Chaki, M., Carreras, A., López-Jaramillo, J., Begara-Morales, J.C., Sánchez-Calvo, B., Valderrama, R., Corpas, F.J. and Barroso, J.B. 2013. Tyrosine nitration provokes inhibition of sunflower carbonic anhydrase (β-CA) activity under high temperature stress. *Nitric Oxide*, 29: 30-33.
- Chaki, M., Fernández-Ocaña, A.M., Valderrama, R., Carreras, A., Esteban, F.J., Luque, F., Gómez-Rodríguez, M.V., Begara-Morales, J.C., Corpas, F.J. and Barroso, J.B. 2008. Involvement of reactive nitrogen and ox-

- ygen species (RNS and ROS) in sunflower–mildew interaction. *Plant and Cell Physiology*, 50: 265-279.
- Chaki, M., Valderrama, R., Fernández-Ocaña, A.M., Carreras, A., Gómez-Rodríguez, M.V., López-Jaramillo, J., Begara-Morales, J.C., Sánchez-Calvo, B., Luque, F., Leterrier, M., Corpas, F.J. and Barroso, J.B. 2011. High temperature triggers the metabolism of S-nitrosothiols in sunflower mediating a process of nitrosative stress which provokes the inhibition of ferredoxin–NADP reductase by tyrosine nitration. *Plant Cell and Environment*, 34: 1803-1818.
- Chakravarty, B., Wang-Pruski, G., Flinn, B., Gustafson, V. and Regan, S. 2007. Genetic transformation in potato: approaches and strategies. *American Journal of Potato Research*, 84: 301-311.
- Chardon, F., Virlon, B., Moreau, L., Falque, M., Joets, J., Decousset, L., Murigneux, A. and Charcosset, A. 2004. Genetic architecture of flowering time in maize as inferred from quantitative trait loci meta-analysis and synteny conservation with the rice genome. *Genetics*, 168: 2169-2185.
- Chia, J.-M., Song, C., Bradbury, P.J., Costich, D., de Leon, N., Doebley, J., Elshire, R.J., Gaut, B., Geller, L., Glaubitz, J.C., Gore, M., Guill, K.E., Holland, J., Hufford, M.B., Lai, J., Li, M., Liu, X., Lu, Y., McCombie, R., Nelson, R., Poland, J., Prasanna, B.M., Pyhäjärvi, T., Rong, T., Sekhon, R.S., Sun, Q., Tenaillon, M.I., Tian, F., Wang, J., Xu, X., Zhang, Z., Kaeppler, S.M., Ross-Ibarra, J., McMullen, M.D., Buckler, E.S., Zhang, G., Xu, Y. and Ware, D. 2012. Maize HapMap2 identifies extant variation from a genome in flux. *Nature Genetics*, 44: 803-807.
- Chung, C.-L., Poland, J., Kump, K., Benson, J., Longfellow, J., Walsh, E., Balint-Kurti, P. and Nelson, R. 2011. Targeted discovery of quantitative trait loci for resistance to northern leaf blight and other diseases of maize. *Theoretical and Applied Genetics*, 123: 307-326.
- Collard, B., Jahufer, M., Brouwer, J. and Pang, E. 2005. An introduction to markers, quantitative trait loci (QTL) mapping and marker-assisted selection for crop improvement: the basic concepts. *Euphytica*, 142: 169-196.
- Collins, N.C., Tardieu, F. and Tuberosa, R. 2008. Quantitative trait loci and crop performance under abiotic stress: where do we stand? *Plant physiology*, 147: 469-486.
- Costanzo, S., Simko, I., Christ, B. and Haynes, K. 2005. QTL analysis of late blight resistance in a diploid potato family of Solanum phureja × S. stenotomum. Theoretical and Applied Genetics, 111: 609-617.
- Council, I.G. 2019. Grain Market report GMR498.
- Crosbie, T.M., Eathington, S.R., Johnson Sr, G.R, Edwards, M., Reiter, R., Stark, S., Mohanty, R.G., Oyervides, M., Buehler, R.E., Walker, A.K., Dobert, R.C., Delannay, X., Pershing, J.C., Hall, M.A. and Kendall, L. 2008. Plant breeding: past, present, and future. In: *Plant breeding: the Arnel R. Hallauer international symposium*, Wiley Online Library, pp 3-50.

- da Silva Dias, J.C. 2015. Biodiversity and Plant Breeding as Tools for Harmony Between Modern Agriculture Production and the Environment. *In: Molecular Approaches to Genetic Diversity.* IntechOpen.
- Danan, S., Veyrieras, J.-B. and Lefebvre, V. 2011. Construction of a potato consensus map and QTL meta-analysis offer new insights into the genetic architecture of late blight resistance and plant maturity traits. *BMC Plant Biology*, 11: 16.
- Darvishzadeh, R., Azizi, M., Hatami-Maleki, H., Bernousi, I., Mandoulakani, B.A., Jafari, M. and Sarrafi, A. 2010. Molecular characterization and similarity relationships among sunflower (*Helianthus annuus* L.) inbred lines using some mapped simple sequence repeats. *African Journal of Biotechnology*, 9: 7280-7288.
- Dimitrijevic, A, and Horn, R. 2018. Sunflower hybrid breeding: From markers to genomic selection. *Frontiers in Plant Science*, 8: 2238.
- Eathington, S.R., Crosbie, T.M., Edwards, M.D., Reiter, R.S. and Bull, J.K. 2007. Molecular markers in a commercial breeding program. *Crop Science*, 47: S-154-S-163.
- Edgerton, M.D. 2009. Increasing crop productivity to meet global needs for feed, food, and fuel. *Plant Physiology*, 149: 7-13.
- El-Banna, A., Hajirezaei, M.R., Wissing, J., Ali, Z., Vaas, L., Heine-Dobbernack, E., Jacobsen, H.J., Schumacher, H.M. and Kiesecker, H 2010. Over-expression of PR-10a leads to increased salt and osmotic tolerance in potato cell cultures. *Journal of Biotechnology*, 150: 277-287.
- Esquinas-Alcázar, J. 2005. Protecting crop genetic diversity for food security: political, ethical and technical challenges. *Nature Reviews Genetics*, 6: 946-953.
- Felcher, K.J., Coombs, J.J., Massa, A.N., Hansey, C.N., Hamilton, J.P., Veilleux, R.E., Buell, C.R. and Douches, D.S. 2012. Integration of two diploid potato linkage maps with the potato genome sequence. *PloS ONE*, 7: e36347.
- Feng, J., Vick, B.A., Lee, M.-K., Zhang, H.-B. and Jan, C. 2006. Construction of BAC and BIBAC libraries from sunflower and identification of linkage group-specific clones by overgo hybridization. *Theoretical and Applied Genetics*, 113: 23-32.
- Fernández-del-Carmen, A., Celis-Gamboa, C., Visser, R.G. and Bachem, C.W. 2007. Targeted transcript mapping for agronomic traits in potato. *Journal of Experimental Botany*, 58: 2761-2774.
- Fernández-Luqueño, F., López-Valdez, F., Miranda-Arámbula, M. and Rosas-Morales, M. 2014. An Introduction to the Sunflower Crop. *In book: Sunflowers: Growth and Development, Environmental Influences and Pests/Diseases,* Edition: First, Chapter: 1, Publisher: Nova Science Publishers, Editors: Juan Ignacio Arribas, pp.1-18
- Fernandez, M.G.S., Becraft, P.W., Yin, Y. and Lübber-

- stedt, T. 2009. From dwarves to giants? Plant height manipulation for biomass yield. *Trends in Plant Science*, 14: 454-461.
- Filippi, C.V., Aguirre, N., Rivas, J.G., Zubrzycki, J., Puebla, A., Cordes, D., Moreno, M.V., Fusari, C.M., Alvarez, D., Heinz, R.A., Hopp, H.E., Paniego, N.B. and Lia, V.V. 2015. Population structure and genetic diversity characterization of a sunflower association mapping population using SSR and SNP markers. *BMC Plant Biology*, 15: 52.
- Flinn, B., Rothwell, C., Griffiths, R., Lägue, M., DeKoeyer, D., Sardana, R., Audy, P., Goyer, C., Li, X.Q., Wang-Pruski, G. and Regan, S. 2005. Potato expressed sequence tag generation and analysis using standard and unique cDNA libraries. *Plant Molecular Biology*, 59: 407-433.
- Fu, Y.-B. 2015. Understanding crop genetic diversity under modern plant breeding. *Theoretical and Applied Genetics*, 128: 2131-2142.
- Fulladolsa, A.C., Navarro, F.M., Kota, R., Severson, K., Palta, J.P. and Charkowski, A.O. 2015. Application of marker assisted selection for *Potato virus Y* resistance in the University of Wisconsin Potato Breeding Program. *American Journal of Potato Research*, 92: 444-450.
- Gallou, A., Declerck, S. and Cranenbrouck, S. 2012. Transcriptional regulation of defence genes and involvement of the WRKY transcription factor in arbuscular mycorrhizal potato root colonization. Functional & integrative genomics, 12: 183-198.
- Gao, L. and Bradeen, J.M. 2016. Contrasting potato foliage and tuber defense mechanisms against the late blight pathogen *Phytophthora infestans*. *PloS ONE*, 11: e0159969.
- García-Lara, S., Khairallah, M.M., Vargas, M. and Bergvinson, D.J. 2009. Mapping of QTL associated with maize weevil resistance in tropical maize. *Crop Science*, 49: 139-149.
- Gebhardt, C., Bellin, D., Henselewski, H., Lehmann, W., Schwarzfischer, J. and Valkonen, J. 2006. Marker-assisted combination of major genes for pathogen resistance in potato. *Theoretical and Applied Genetics*, 112: 1458-1464.
- Gentzbittel, L., Abbott, A., Galaud, J., Georgi, L., Fabre, F., Liboz, T. and Alibert, G. 2002. A bacterial artificial chromosome (BAC) library for sunflower, and identification of clones containing genes for putative transmembrane receptors. *Molecular Genetics and Genomics*, 266: 979-987.
- Giordani, T., Cavallini, A. and Natali, L. 2014. The repetitive component of the sunflower genome. *Current Plant Biology,* 1: 45-54.
- Glaser, J.A. and Matten, S.R. 2003, Sustainability of insect resistance management strategies for transgenic Bt corn. *Biotechnology Advances*, 22: 45-69.
- Goffinet, B. and Gerber, S. 2000. Quantitative trait loci: a meta-analysis. *Genetics*, 155:463-473
- Gore, M.A., Chia, J.M., Elshire, R.J., Sun, Q., Ersoz, E.S., Hurwitz, B.L., Peiffer, J.A., McMullen, M.D.,

- Grills, G.S., Ross-Ibarra, J., Ware, D.H. and Buckler, E.S. 2009. A first-generation haplotype map of maize. *Science*, 326: 1115-1117.
- Guillaumie, S., Pichon, M., Martinant, J.-P., Bosio, M., Goffner, D. and Barrière, Y. 2007a. Differential expression of phenylpropanoid and related genes in brown-midrib bm1, bm2, bm3, and bm4 young near-isogenic maize plants. *Planta*, 226: 235-250.
- Guillaumie, S., San-Clemente, H., Deswarte, C., Martinez, Y., Lapierre, C., Murigneux, A., Barrière, Y., Pichon, M. and Goffner, D. 2007b. MAIZEWALL. Database and developmental gene expression profiling of cell wall biosynthesis and assembly in maize. *Plant Physiology*, 143: 339-363.
- Hamrit, S., Kusterer, B., Friedt, W. and Horn, R. 2008. Verification of positive BAC clones near the Rf1 gene restoring pollen fertility in the presence of the PET1 cytoplasm in sunflower (*Helianthus annuus* L.) and direct isolation of BAC ends. In: *Proceedings of the 17th International Sunflower Conference*; Córdoba, Spain. 8–12 June 2008; pp. 623–628.
- Hao, Z., Li, X., Liu, X., Xie, C., Li, M., Zhang, D., Zhang S (2010) Meta-analysis of constitutive and adaptive QTL for drought tolerance in maize. *Euphytica*, 174: 165-177.
- Hardigan, M.A., Crisovan, E., Hamilton, J.P., Kim, J., Laimbeer, P., Leisner, C.P., Manrique-Carpintero, N.C., Newton, L., Pham, G.M., Vaillancourt, B., Yang, X., Zeng, Z., Douches, D.S., Jiang, J., Veilleux, R.E. and Buell, C.R. 2016. Genome reduction uncovers a large dispensable genome and adaptive role for copy number variation in asexually propagated *Solanum tuberosum*. *The Plant Cell*, 28: 388-405.
- He, Z., Zhong, J., Sun, X., Wang, B., Terzaghi, W. and Dai, M. 2018. The Maize ABA Receptors ZmPYL8, 9, and 12 Facilitate Plant Drought Resistance. *Frontiers in Plant Science*, 9: 422.
- Hewezi, T., Mouzeyar, S., Thion, L., Rickauer, M., Alibert, G., Nicolas, P. and Kallerhoff, J. 2006. Antisense expression of a NBS-LRR sequence in sunflower (*Helianthus annuus* L.) and tobacco (*Nicotiana tabacum* L.): evidence for a dual role in plant development and fungal resistance. *Transgenic research*, 15: 165-180.
- Hmida-Sayari, A., Gargouri-Bouzid, R., Bidani, A., Jaoua, L., Savouré, A. and Jaoua, S. 2005. Overexpression of Δ 1-pyrroline-5-carboxylate synthetase increases proline production and confers salt tolerance in transgenic potato plants. *Plant Science*, 169: 746-752.
- Höniges, A., Wegmann, K. and Ardelean, A. 2008. Orobanche resistance in sunflower/resistencia a Orobanche en girasol/résistance à l'orobanche chez le tournesol. *Helia*, 31: 1-12.
- Hu, X., Bidney, D.L., Yalpani, N., Duvick, J.P., Crasta, O., Folkerts, O. and Lu, G. 2003. Overexpression of a gene encoding hydrogen peroxide-generating oxalate oxidase evokes defense responses in sunflower. *Plant Physiology*, 133: 170-181.

- Hufford, M.B., Xu, X., van Heerwaarden, J., Pyhäjärvi, T., Chia, J.M., Cartwright, R.A., Elshire, R.J., Glaubitz, J.C., Guill, K.E., Kaeppler, S.M., Lai, J., Morrell, P.L., Shannon, L.M., Song, C., Springer, N.M., Swanson-Wagner, R.A., Tiffin, P., Wang, J., Zhang, G., Doebley, J., McMullen, M.D., Ware, D., Buckler, E.S., Yang, S. and Ross-Ibarra, J. 2012. Comparative population genomics of maize domestication and improvement. *Nature genetics*, 44: 808-811.
- Huo, D., Ning, Q., Shen, X., Liu, L. and Zhang, Z. 2016. QTL mapping of kernel number-related traits and validation of one major QTL for ear length in maize. *PloS ONE*, 11: e0155506.
- Hvarleva, T., Tarpomanova, I., Hristova-Cherbadji, M., Hristov, M., Bakalova, A., Atanassov, A. and Atanasov, I. 2009. Toward marker assisted selection for fungal disease resistance in sunflower. Utilization of H. Bolanderi as a source of resistance to Downy mildew. *Biotechnology & Biotechnological Equipment*, 23: 1427-1430.
- Jiang, R., Li, J., Tian, Z., Du, J., Armstrong, M., Baker, K., Tze-Yin Lim, J., Vossen, J.H., He, H., Portal, L., Zhou, J., Bonierbale, M., Hein, I., Lindqvist-Kreuze, H. and Xie, C. 2018. Potato late blight field resistance from QTL dPl09c is conferred by the NB-LRR gene R8. *Journal of experimental botany*, 69: 1545-1555.
- Jo, K.-R., Arens, M., Kim, T.-Y., Jongsma, M.A., Visser, R.G., Jacobsen, E. and Vossen, J.H. 2011. Mapping of the S. demissum late blight resistance gene R8 to a new locus on chromosome IX. *Theoretical and Applied Genetics*, 123: 1331-1340.
- Juliatti, F.C., Pedrosa, M.G., Silva, H.D. and da Silva, J.V.C. 2009. Genetic mapping for resistance to gray leaf spot in maize. *Euphytica*, 169: 227-238.
- Katerji, N., Van Hoorn, J., Hamdy, A. and Mastrorilli, M. 2000. Salt tolerance classification of crops according to soil salinity and to water stress day index. Agricultural Water Management, 43: 99-109.
- Khadeeva, N., Kochieva, E.Z., Tcherednitchenko, M.Y., Yakovleva, E.Yu., Sydoruk, K.V., Bogush, V.G., Dunaevsky, Y.E. and Belozersky, M.A. 2009. Use of buckwheat seed protease inhibitor gene for improvement of tobacco and potato plant resistance to biotic stress. *Biochemistry (Moscow)*, 74: 260-267.
- Kim, Y.-H., Bae, J.M. and Huh, G.-H. 2010. Transcriptional regulation of the cinnamyl alcohol dehydrogenase gene from sweetpotato in response to plant developmental stage and environmental stress. *Plant cell reports*, 29: 779-791.
- Klopfenstein, T., Erickson, G. and Berger, L. 2013. Maize is a critically important source of food, feed, energy and forage in the USA. *Field Crops Research*, 153: 5-11.
- Kolkman, J.M., Slabaugh, M.B., Bruniard, J.M., Berry, S., Bushman, B.S., Olungu, C., Maes, N., Abratti, G., Zambelli, A., Miller, J.F., Leon, A. and Knapp, S.J. 2004. Acetohydroxyacid synthase mutations conferring resistance to imidazolinone or sulfonylurea herbicides in sunflower. *Theoreti*-

- cal and Applied Genetics, 109: 1147-1159.
- Krakowsky, M., Lee, M., Woodman-Clikeman, W., Long, M. and Sharopova, N. 2004. QTL Mapping of Resistance to Stalk Tunneling by the European Corn Borer in RILs of Maize Population B73× De8. *Crop Science*, 44: 274-282.
- Lai, Z., Kane, N.C., Kozik, A., Hodgins, K.A., Dlugosch, K.M., Barker, M.S., Matvienko, M., Yu, Q., Turner, K.G., Pearl, S.A., Bell, G.D., Zou, Y., Grassa, C., Guggisberg, A., Adams, K.L., Anderson, J.V., Horvath, D.P., Kesseli, R.V., Burke, J.M., Michelmore, R.W. and Rieseberg, L.H. 2012. Genomics of Compositae weeds: EST libraries, microarrays, and evidence of introgression. *American Journal of Botany*, 99: 209-218.
- Lai, Z., Livingstone, K., Zou, Y., Church, S., Knapp, S., Andrews, J. and Rieseberg, L. 2005. Identification and mapping of SNPs from ESTs in sunflower. *Theoretical and Applied Genetics*, 111: 1532-1544.
- Landi, P., Sanguineti, M.C., Liu, C., Li, Y., Wang, T.Y., Giuliani, S., Bellotti, M., Salvi, S. and Tuberosa, R. 2006. Root-ABA1 QTL affects root lodging, grain yield, and other agronomic traits in maize grown under well-watered and water-stressed conditions. *Journal of experimental botany*, 58: 319-326.
- Landi, P., Sanguineti, M.C., Salvi, S., Giuliani, S., Bellotti, M., Maccaferri, M., Sergio, S. and Tuberosa, R. 2005. Validation and characterization of a major QTL affecting leaf ABA concentration in maize. *Molecular Breeding*, 15: 291-303.
- Laurie, C.C., Chasalow, S.D., LeDeaux, J.R., McCarroll, R., Bush, D., Hauge, B., Lai, C., Clark, D., Rocheford, T.R. and Dudley, J.W. 2004. The genetic architecture of response to long-term artificial selection for oil concentration in the maize kernel. *Genetics*, 168: 2141-2155.
- Lee, H. and Zhang, Z.J. 2014. Agrobacterium-mediated transformation of maize (*Zea mays*) immature embryos. In: *Cereal Genomics*. Springer, pp 273-280
- Legay, S., Lamoureux, D., Hausman, J.-F., Hoffmann, L. and Evers, D. 2009. Monitoring gene expression of potato under salinity using cDNA microarrays. *Plant Cell Reports*, 28: 1799-1816.
- Legay, S., Lefèvre, I., Lamoureux, D., Barreda, C., Luz, R.T., Gutierrez, R., Quiroz, R., Hoffmann, L., Hausman, J.F., Bonierbale, M., Evers, D. and Schafleitner, R. 2011. Carbohydrate metabolism and cell protection mechanisms differentiate drought tolerance and sensitivity in advanced potato clones (Solanum tuberosum L.). Functional & Integrative Genomics, 11: 275-291.
- Li, H., Freeling, M. and Lisch, D. 2010. Epigenetic reprogramming during vegetative phase change in maize. *Proceedings of the National Academy of Sciences*, 107: 22184-22189.
- Liu, S., Chen, H.D., Makarevitch, I., Shirmer, R., Emrich, S.J., Dietrich, C.R., Barbazuk, W.B., Springer, N.M. and Schnable, P.S. 2010. High-throughput genetic mapping of mutants via quantitative

- single nucleotide polymorphism typing. *Genetics*, 184: 19-26.
- Lorenzana, R.E., Lewis, M.F., Jung, H.-J.G. and Bernardo, R. 2010. Quantitative trait loci and trait correlations for maize stover cell wall composition and glucose release for cellulosic ethanol. *Crop Science*, 50: 541-555.
- Lu, G. 2003. Engineering *Sclerotinia sclerotiorum* resistance in oilseed crops. *African Journal of Biotechnology*, 2: 509-516.
- Lu, G. and Hoeft, E. 2009. Sunflower Compendium of transgenic. *Crop Plants*, 125-168.
- Ma, G., Markell, S., Song, Q. and Qi, L. 2017. Genotyping-by-sequencing targeting of a novel downy mildew resistance gene Pl20 from wild *Helianthus argophyllus* for sunflower (*Helianthus annuus* L.). *Theoretical and Applied Genetics*, 130: 1519-1529.
- McMullen, M.D., Kresovich, S., Villeda, H.S., Bradbury, P., Li, H., Sun, Q., Flint-Garcia, S., Thornsberry, J., Acharya, C., Bottoms, C., Brown, P., Browne, C., Eller, M., Guill, K., Harjes, C., Kroon, D., Lepak, N., Mitchell, S.E., Peterson, B., Pressoir, G., Romero, S., Oropeza Rosas, M., Salvo, S., Yates, H., Hanson, M., Jones, E., Smith, S., Glaubitz, J.C., Goodman, M., Ware, D., Holland, J.B. and Buckler, E.S. 2009. Genetic properties of the maize nested association mapping population. *Science*, 325: 737-740.
- Menéndez, C.M., Ritter, E., Schäfer-Pregl, R., Walkemeier, B., Kalde, A., Salamini, F., Gebhardt, C. 2002. Cold sweetening in diploid potato: mapping quantitative trait loci and candidate genes. *Genetics*, 162: 1423-1434.
- Messmer, R., Fracheboud, Y., Bänziger, M., Vargas, M., Stamp, P. and Ribaut, J.-M. 2009. Drought stress and tropical maize: QTL-by-environment interactions and stability of QTLs across environments for yield components and secondary traits. *Theoretical and Applied Genetics*, 119: 913-930.
- Micic, Z., Hahn, V., Bauer, E., Melchinger, A., Knapp, S., Tang, S. and Schön, C. 2005. Identification and validation of QTL for Sclerotinia midstalk rot resistance in sunflower by selective genotyping. *Theoretical and Applied Genetics*, 111: 233-242.
- Moloney, C., Griffin, D., Jones, P.W., Bryan, G.J., McLean, K., Bradshaw, J.E. and Milbourne, D. 2010. Development of diagnostic markers for use in breeding potatoes resistant to *Globodera pallida* pathotype Pa2/3 using germplasm derived from *Solanum tuberosum* ssp. *andigena* CPC 2802. *Theoretical and Applied Genetics*, 120: 679-689.
- Nair, S.K., Prasanna, B.M., Garg, A., Rathore, R., Setty, T. and Singh, N. 2005. Identification and validation of QTLs conferring resistance to sorghum downy mildew (*Peronosclerospora sorghi*) and Rajasthan downy mildew (*P. heteropogoni*) in maize. *Theoretical and Applied Genetics*, 110: 1384-1392.
- Naqvi, S., Zhu, C., Farre, G., Ramessar, K., Bassie, L.,

- Breitenbach, J., Perez Conesa, D., Ros, G., Sandmann, G., Capell, T. and Christou, P. 2009. Transgenic multivitamin corn through biofortification of endosperm with three vitamins representing three distinct metabolic pathways. *Proceedings of the National Academy of Sciences*, 106: 7762-7767.
- Nelson, D.E., Repetti, P.P., Adams, T.R., Creelman, R.A., Wu, J., Warner, D.C., Anstrom, D.C., Bensen, R.J., Castiglioni, P.P., Donnarummo, M.G., Hinchey, B.S., Kumimoto, R.W., Maszle, D.R., Canales, R.D., Krolikowski, K.A., Dotson, S.B., Gutterson, N., Ratcliffe, O.J. and Heard, J.E. 2007. Plant nuclear factor Y (NF-Y) B subunits confer drought tolerance and lead to improved corn yields on waterlimited acres. *Proceedings of the National Academy of Sciences*, 104: 16450-16455.
- Nobuta, K., Lu, C., Shrivastava, R., Pillay, M., De Paoli, E., Accerbi, M., Arteaga-Vazquez, M., Sidorenko, L., Jeong, D.H., Yen, Y., Green, P.J., Chandler, V.L. and Meyers, B.C. 2008. Distinct size distribution of endogenous siRNAs in maize: Evidence from deep sequencing in the mop1-1 mutant. *Proceedings of the National Academy of Sciences*, 105: 14958-14963.
- Ohta, S., Ishida, Y. and Usami, S. 2004. Expression of cold-tolerant pyruvate, orthophosphate dikinase cDNA, and heterotetramer formation in transgenic maize plants. *Transgenic research*, 13: 475-485.
- Özdemir, N., Horn, R. and Friedt, W. 2004. Construction and characterization of a BAC library for sunflower (*Helianthus annuus* L.) *Euphytica*, 138: 177-183.
- Penning, B.W., Hunter, C.T. 3rd, Tayengwa, R., Eveland, A.L., Dugard, C.K., Olek, A.T., Vermerris, W., Koch, K.E., McCarty, D.R., Davis, M.F., Thomas, S.R., McCann, M.C. and Carpita, N.C. 2009. Genetic resources for maize cell wall biology. *Plant Physiology*, 151: 1703-1728.
- Pérez-Vich, B., Akhtouch, B., Knapp, S., Leon, A., Velasco, L., Fernández-Martínez, J. and Berry, S. 2004. Quantitative trait loci for broomrape (*Orobanche cumana* Wallr.) resistance in sunflower. *Theoretical and Applied Genetics*, 109: 92-102.
- Pernet, A., Hoisington, D., Dintinger, J., Jewell, D., Jiang, C., Khairallah, M., Letourmy, P., Marchand, J.L., Glaszmann, J.C. and González de León, D. 1999. Genetic mapping of maize streak virus resistance from the Mascarene source. I. Resistance in line D211 and stability against different virus clones. *Theoretical and Applied Genetics*, 99: 524-539.
- Poland, J.A., Bradbury, P.J., Buckler, E.S. and Nelson, R.J. 2011. Genome-wide nested association mapping of quantitative resistance to northern leaf blight in maize. *Proceedings of the National Academy of Sciences*, 108: 6893-6898.
- Pozar, G., Butruille, D., Silva, H.D., McCuddin, Z.P. and Penna, J.C.V. 2009. Mapping and validation of quantitative trait loci for resistance to Cer-

- cospora zeae-maydis infection in tropical maize (*Zea mays* L.). *Theoretical and Applied Genetics*, 118: 553-564.
- Prasanna, B., Pixley, K., Warburton, M.L., Xie, C.-X. 2010. Molecular marker-assisted breeding options for maize improvement in Asia. *Molecular Breeding*, 26: 339-356.
- Qi, L., Long, Y., Talukder, Z.I., Seiler, G.J., Block, C.C. and Gulya, T.J. 2016. Genotyping-by-sequencing uncovers the introgression alien segments associated with Sclerotinia basal stalk rot resistance from wild species—I. *Helianthus argophyllus* and *H. petiolaris. Frontiers in Genetics*, 7: 219.
- Quan, R., Shang, M., Zhang, H., Zhao, Y. and Zhang, J. 2004. Improved chilling tolerance by transformation with betA gene for the enhancement of glycinebetaine synthesis in maize. *Plant Science*, 166: 141-149.
- Rahnama, H. and Ebrahimzadeh, H. 2004. The effect of NaCl on proline accumulation in potato seedlings and calli. *Acta Physiologiae Plantarum*, 26: 263-270.
- Rajhi, I., Yamauchi, T., Takahashi, H., Nishiuchi, S., Shiono, K., Watanabe, R., Mliki, A., Nagamura, Y., Tsutsumi, N., Nishizawa, N.K. and Nakazono, M. 2011. Identification of genes expressed in maize root cortical cells during lysigenous aerenchyma formation using laser microdissection and microarray analyses. *New Phytologist*, 190: 351-368.
- Redinbaugh, M.G. and Pratt, R.C. 2009. Virus resistance. In: *Handbook of maize: Its biology*. Springer, pp 251-270
- Reid, A. and Kerr, E. 2007. A rapid simple sequence repeat (SSR)-based identification method for potato cultivars. *Plant Genetic Resources*, 5: 7-13.
- Renaut, S. 2017. Genome sequencing: Illuminating the sunflower genome. *Nature Plants*, 3: 17099.
- Rensink, W., Hart, A., Liu, J., Ouyang, S., Zismann, V. and Buell, C.R. 2005. Analyzing the potato abiotic stress transcriptome using expressed sequence tags. *Genome*, 48: 598-605.
- Revilla, P., Rodríguez, V.M., Ordás, A., Rincent, R., Charcosset, A., Giauffret, C., Melchinger, A.E., Schön, C.-C., Bauer, E., Altmann, T., Brunel, D., Moreno-González, J., Campo, L., Ouzunova, M., Álvarez, A., Ruíz de Galarreta, J.I., Laborde, J. and Malvar, R.A. 2016. Association mapping for cold tolerance in two large maize inbred panels. *BMC Plant Biology*, 16: 127.
- Ribaut, J.-M., Betran, J., Monneveux, P. and Setter, T. 2009. Drought tolerance in maize. In: *Handbook of Maize: Its Biology*. Springer, pp 311-344.
- Ribaut, J.-M. and Ragot, M. 2006. Marker-assisted selection to improve drought adaptation in maize: the backcross approach, perspectives, limitations, and alternatives. *Journal of Experimental Botany*, 58: 351-360.
- Rickert, A.M., Kim, J.H., Meyer, S., Nagel, A., Ballvora, A., Oefner, P.J. and Gebhardt, C. 2003. First-

- generation SNP/InDel markers tagging loci for pathogen resistance in the potato genome. *Plant Biotechnology Journal*, 1: 399-410.
- Robinson, M.R., Wray, N.R. and Visscher, P.M. 2014. Explaining additional genetic variation in complex traits. *Trends in Genetics*, 30: 124-132.
- Rouf Shah, T., Prasad, K. and Kumar, P. 2016. Maize—A potential source of human nutrition and health: *A review Cogent Food & Agriculture*, 2: 1166995.
- Sawahel, W. and Hagran, A. 2006. Generation of white mold disease-resistant sunflower plants expressing human lysozyme gene. *Biologia Plantarum*, 50: 683-687.
- Schnabl, H., Binsfeld, P., Cerboncini, C., Dresen, B., Peisker, H., Wingender, R. and Henn, A. 2002. Biotechnological methods applied to produce *Sclerotinia sclerotiorum* resistant sunflower/métodos biotecnológicos empleados para producir girasol resistente contra *Sclerotinia sclerotiorum*/méthodes biotechnologiques ont appliqué pour produire le *Sclerotinia sclerotiorum* résistant tournesol. *Helia*, 25: 191-198.
- Schnable, J.C., Springer, N.M. and Freeling, M. 2011. Differentiation of the maize subgenomes by genome dominance and both ancient and ongoing gene loss. *Proceedings of the National Academy of Sciences*, 108: 4069-4074.
- Schnable, P.S., Ware, D., Fulton, R.S., Stein, J.C., Wei, F., Pasternak, S., Liang, C., Zhang, J., Fulton, L., Graves, T.A., Minx, P., Reily, A.D., Courtney, L., Kruchowski, S.S., Tomlinson, C., Strong, C., Delehaunty, K., Fronick, C., Courtney, B., Rock, S.M., Belter, E., Du, F., Kim, K., Abbott, R.M., Cotton, M., Levy, A., Marchetto, P., Ochoa, K., Jackson, S.M., Gillam, B., Chen, W., Yan, L., Higginbotham, J., Cardenas, M., Waligorski, J., Applebaum, E., Phelps, L., Falcone, J., Kanchi, K., Thane, T., Scimone, A., Thane, N., Henke, J., Wang, T., Ruppert, J., Shah, N., Rotter, K., Hodges, J., Ingenthron, E., Cordes, M., Kohlberg, S., Sgro, J., Delgado, B., Mead, K., Chinwalla, A., Leonard, S., Crouse, K., Collura, K., Kudrna, D., Currie, J., He, R., Angelova, A., Rajasekar, S., Mueller, T., Lomeli, R., Scara, G., Ko, A., Delaney, K., Wissotski, M., Lopez, G., Campos, D., Braidotti, M., Ashley, E., Golser, W., Kim, H., Lee, S., Lin, J., Dujmic, Z., Kim, W., Talag, J., Zuccolo, A., Fan, C., Sebastian, A., Kramer, M., Spiegel, L., Nascimento, L., Zutavern, T., Miller, B., Ambroise, C., Muller, S., Spooner, W., Narechania, A., Ren, L., Wei, S., Kumari, S., Faga, B., Levy, M.J., McMahan, L., Van Buren, P., Vaughn, M.W., Ying, K., Yeh, C.T., Emrich, S.J., Jia, Y., Ka-Iyanaraman, A., Hsia, A.P., Barbazuk, W.B., Baucom, R.S., Brutnell, T.P., Carpita, N.C., Chaparro, C., Chia, J.M., Deragon, J.M., Estill, J.C., Fu, Y., Jeddeloh, J.A., Han, Y., Lee, H., Li, P., Lisch, D.R, Liu, S., Liu, Z., Nagel, D.H., McCann, M.C., SanMiguel, P., Myers, A.M., Nettleton, D., Nguyen, J., Penning, B.W., Ponnala, L., Schneider, K.L., Schwartz, D.C., Sharma, A., Soderlund, C., Springer, N.M., Sun, Q., Wang, H., Waterman, M., Westerman, R.,

- Wolfgruber, T.K., Yang, L., Yu, Y., Zhang, L., Zhou, S., Zhu, Q., Bennetzen, J.L., Dawe, R.K., Jiang, J., Jiang, N., Presting, G.G., Wessler, S.R., Aluru, S., Martienssen, R.A., Clifton, S.W., McCombie, W.R., Wing, R.A. and Wilson, R.K. 2009. The B73 maize genome: complexity, diversity, and dynamics. *Science*, 326: 1112-1115.
- Schuppert, G.F., Tang, S., Slabaugh, M.B. and Knapp, S.J. 2006. The sunflower high-oleic mutant Ol carries variable tandem repeats of FAD2-1, a seed-specific oleoyl-phosphatidyl choline desaturase. *Molecular Breeding*, 17: 241-256.
- Sekhon, R.S., Lin, H., Childs, K.L., Hansey, C.N., Buell, C.R., de Leon, N. and Kaeppler, S.M. 2011. Genome-wide atlas of transcription during maize development. *The Plant Journal*, 66: 553-563.
- Sharma, S.K., Bolser, D., de Boer, J., Sønderkær, M., Amoros, W., Carboni, M.F., D'Ambrosio, J.M., de la Cruz, G., Di Genova, A., Douches, D.S., Eguiluz, M., Guo, X., Guzman, F., Hackett, C.A., Hamilton, J.P., Li, G., Li, Y., Lozano, R., Maass, A., Marshall, D., Martinez, D., McLean, K., Mejía, N., Milne, L., Munive, S., Nagy, I., Ponce, O., Ramirez, M., Simon, R., Thomson, S.J., Torres, Y., Waugh, R., Zhang, Z., Huang, S., Visser, R.G., Bachem, C.W., Sagredo, B., Feingold, S.E., Orjeda, G., Veilleux, R.E., Bonierbale, M., Jacobs, J.M., Milbourne, D., Martin, D.M. and Bryan, G.J. 2013. Construction of reference chromosome-scale pseudomolecules for potato: integrating the potato genome with genetic and physical maps G3: Genes, Genomes. Genetics, 3: 2031-2047.
- Shin, D., Moon, S.J., Han, S., Kim, B.G., Park, S.R., Lee, S.K., Yoon, H.J., Lee, H.E., Kwon, H.B., Baek, D., Yi, B.Y. and Byun, M.O. 2011. Expression of StMY-B1R-1, a novel potato single MYB-like domain transcription factor, increases drought tolerance. *Plant Physiology*, 155: 421-432.
- Shrawat, A.K. and Lörz, H. 2006. Agrobacterium-mediated transformation of cereals: a promising approach crossing barriers. *Plant Biotechnology Journal*, 4: 575-603.
- Šimko, I., Vreugdenhil, D., Jung, C. and May, G. 1999. Similarity of QTLs detected for in vitro and greenhouse development of potato plants. *Mo-lecular Breeding*, 5: 417-428.
- Soderlund, C., Descour, A., Kudrna, D., Bomhoff, M., Boyd, L., Currie, J., Angelova, A., Collura, K., Wissotski, M., Ashley, E., Morrow, D., Fernandes, J., Walbot, V. and Yu, Y. 2009. Sequencing, mapping, and analysis of 27,455 maize full-length cDNAs. *PLoS Genetics*, 5: e1000740.
- Stefańczyk, E., Sobkowiak, S., Brylińska, M. and Śliwka, J. 2017. Expression of the potato late blight resistance gene Rpi-phu1 and *Phytophthora infestans* effectors in the compatible and incompatible interactions in potato. *Phytopathology*, 107: 740-748.
- Sun, K., Wolters, A.-M.A., Vossen, J.H., Rouwet, M.E., Loonen, A.E.H.M., Jacobsen, E., Visser, R.G.F., Ba-

- icorresponding, Y. 2016. Silencing of six susceptibility genes results in potato late blight resistance. *Transgenic Research*, 25: 731-742.
- Szajko, K., Strzelczyk-Żyta, D. and Marczewski, W. 2014. Ny-1 and Ny-2 genes conferring hypersensitive response to *Potato Virus Y* (PVY) in cultivated potatoes: mapping and marker-assisted selection validation for PVY resistance in potato breeding. *Molecular Breeding*, 34: 267-271.
- Talia, P., Nishinakamasu, V., Hopp, H.E., Heinz, R.A. and Paniego, N. 2010. Genetic mapping of EST-SSR, SSR and InDel to improve saturation of genomic regions in a previously developed sunflower map. *Electronic Journal of Biotechnology*, 13: 7-8.
- Talukder, Z.I., Seiler, G.J., Song, Q., Ma, G. and Qi, L. 2016. SNP discovery and QTL mapping of Sclerotinia basal stalk rot resistance in sunflower using genotyping-by-sequencing. *The Plant Genome*. 9.
- Tamborindeguy, C., Ben, C., Liboz, T. and Gentzbittel, L. 2004. Sequence evaluation of four specific cDNA libraries for developmental genomics of sunflower. *Molecular Genetics and Genomics*. 271: 367-375.
- Tang, L., Kwon, S.Y., Kim, S.H., Kim, J.S., Choi, J.S., Cho, K.Y., Sung, C.K., Kwak, S.S. and Lee, H.S. 2006a. Enhanced tolerance of transgenic potato plants expressing both superoxide dismutase and ascorbate peroxidase in chloroplasts against oxidative stress and high temperature. *Plant Cell Reports*, 25: 1380-1386.
- Tang, R., Zhu, W., Song, X., Lin, X., Cai, J., Wang, M. and Yang, Q. 2016. Genome-wide identification and function analyses of heat shock transcription factors in potato. *Frontiers in Plant Science*, 7: 490.
- Tang, S. and Knapp, S.J. 2003. Microsatellites uncover extraordinary diversity in native American land races and wild populations of cultivated sunflower TAG. *Theoretical and Applied Genetics*, 106: 990-1003.
- Tang, S., Leon, A., Bridges, W.C. and Knapp, S.J. 2006b. Quantitative trait loci for genetically correlated seed traits are tightly linked to branching and pericarp pigment loci in sunflower. Crop Science, 46: 721-734.
- Tian, Z.D., Zhang, Y., Liu, J. and Xie, C.H. 2010. Novel potato C2H2-type zinc finger protein gene, StZFP1, which responds to biotic and abiotic stress, plays a role in salt tolerance. *Plant Biology*, 12: 689-697.
- Tomczyńska, I., Jupe, F., Hein, I., Marczewski, W. and Śliwka, J. 2014. Hypersensitive response to *Potato Virus Y* in potato cultivar Sárpo Mira is conferred by the Ny-Smira gene located on the long arm of chromosome IX. *Molecular Breeding*. 34: 471-480.
- Torres, A.F., Visser, R.G. and Trindade, L.M. 2015. Bioethanol from maize cell walls: genes, molecular tools, and breeding prospects. *Gcb Bioenergy*, 7: 591-607.

- Trachsel, S., Messmer, R., Stamp, P. and Hund, A. 2009. Mapping of QTLs for lateral and axile root growth of tropical maize. *Theoretical and Applied Genetics*, 119: 1413-1424.
- Trognitz, F., Manosalva, P., Gysin, R., Niñio-Liu, D., Simon, R., del Herrera, M.R., Trognitz, B., Ghislain, M. and Nelson, R. 2002. Plant defense genes associated with quantitative resistance to potato late blight in *Solanum phureja*× dihaploid *S. tuberosum* hybrids. *Molecular Plant-Microbe Interactions*, 15: 587-597.
- Truntzler, M., Barrière, Y., Sawkins, M., Lespinasse, D., Betran, J., Charcosset, A. and Moreau, L. 2010. Meta-analysis of QTL involved in silage quality of maize and comparison with the position of candidate genes. *Theoretical and Applied Genetics*, 121: 1465-1482.
- Turra, D., Bellin, D., Lorito, M. and Gebhardt, C. 2009. Genotype-dependent expression of specific members of potato protease inhibitor gene families in different tissues and in response to wounding and nematode infection. *Journal of Plant Physiology*, 166: 762-774.
- Upadhyaya, C.P., Young, K.E., Akula, N., Kim, H.S., Heung, J.J., Oh, O.M., Aswath, C.R., Chun, C.H., Kim, D.H. and Park, S.W. 2009. Over-expression of strawberry D-galacturonic acid reductase in potato leads to accumulation of vitamin C with enhanced abiotic stress tolerance. *Plant Science*, 177: 659-667.
- Urbany, C., Stich, B., Schmidt, L., Simon, L., Berding, H., Junghans, H., Niehoff, K.H., Braun, A., Tacke, E., Hofferbert, H.R., Lübeck, J., Strahwald, J. and Gebhardt, C. 2011. Association genetics in Solanum tuberosum provides new insights into potato tuber bruising and enzymatic tissue discoloration. *BMC genomics* 12: 7.
- Van Dijk, J.P., Cankar, K., Stanley, J. Scheffer, S.J., Beenen, H.G., Shepherd, L.V.T., Derek Stewart, D., Davies, H.V., Wilkockson, S.J., Leifert, C., Gruden, K. and Kok, E.J. 2009. Transcriptome Analysis of Potato Tubers Effects of Different Agricultural Practices. Journal of Agricultural and Food Chemistry, 57: 1612-1623.
- Van Os, H., Andrzejewski, S., Bakker, E., Barrena, I., Bryan, G.J., Caromel, B., Ghareeb, B., Isidore, E., de Jong, W., Van Koert, P., Lefebvre, V., Milbourne, D., Ritter, E., Van der Voort, J.N., Rousselle-Bourgeois, F., Van Vliet, J., Waugh, R., Visser, R.G., Bakker, J. and Van Eck, H.J. 2006. Construction of a 10,000-marker ultradense genetic recombination map of potato: providing a framework for accelerated gene isolation and a genomewide physical map. *Genetics*, 173: 1075-1087.
- Varshney, R.K., Bansal, K.C., Aggarwal, P.K., Datta, S.K. and Craufurd, P.Q. 2011. Agricultural biotechnology for crop improvement in a variable climate: hope or hype? *Trends in Plant Science*, 16: 363-371.

- Varshney, R.K., Graner, A. and Sorrells, M.E. 2005. Genic microsatellite markers in plants: features and applications. *Trends in Biotechnology*, 23: 48-55.
- Vermerris, W., Saballos, A., Ejeta, G., Mosier, N.S., Ladisch, M.R. and Carpita, N.C. 2007. Molecular breeding to enhance ethanol production from corn and sorghum stover. *Crop Science*, 47: S-142 -S-153.
- Vielle-Calzada, J.-P., Martínez de la Vega, O., Hernández-Guzmán, G., Ibarra-Laclette, E., Alvarez-Mejía, C., Vega-Arreguín, J.C., Jiménez-Moraila, B., Fernández-Cortés, A., Corona-Armenta, G., Herrera-Estrella, L. and Herrera-Estrella, A. 2009. The Palomero genome suggests metal effects on domestication. *Science*, 326: 1078-1078.
- Vossen, J.H., Van Arkel, G., Bergervoet, M., Jo, K.-R., Jacobsen, E. and Visser, R.G. 2016. The Solanum demissum R8 late blight resistance gene is an Sw-5 homologue that has been deployed worldwide in late blight resistant varieties. *Theoretical and Applied Genetics*, 129: 1785-1796.
- Würschum, T., Anyanga, W.O. and Hahn, V. 2014. Inheritance of Sclerotinia Midstalk Rot Resistance in Elite Sunflower Breeding Germplasm. *Helia*, 37: 193-203.
- Wang, B., Li, Z., Ran, Q., Li, P., Peng, Z. and Zhang, J. 2018. ZmNF-YB16 overexpression improves drought resistance and yield by enhancing photosynthesis and the antioxidant capacity of maize plants. *Frontiers in Plant Science*, 9.
- Wang, G., Hui, W., Jia, Z., Jing, Z., Xiaowei, Z., Fei, W., Yuanping, T., Bing, M., Zhengkai, X. and Rentao, S. 2010. An expression analysis of 57 transcription factors derived from ESTs of developing seeds in maize (*Zea mays*). *Plant Cell Reports*, 29: 545-559.
- William, H.M., Morris, M., Warburton, M. and Hoisington, D.A. 2007. Technical, economic and policy considerations on marker-assisted selection in crops: lessons from the experience at an international agricultural research centre Marker-Assisted Selection: 381.

- Wisser, R.J., Balint-Kurti, P.J. and Nelson, R.J. 2006. The genetic architecture of disease resistance in maize: a synthesis of published studies. *Phytopathology*, 96: 120-129.
- Xu, Y., Skinner, D.J., Wu, H., Palacios-Rojas, N., Araus, J.L., Yan, J., Gao, S., Warburton, M.L. and Crouch, J.H. 2009. Advances in maize genomics and their value for enhancing genetic gains from breeding. *International Journal of Plant Genom*ics 2009.
- Yang, X., Guo, X., Yang, Y., Ye, P., Xiong, X., Liu, J., Dong, D. and Li, G. 2018. Gene Profiling in Late Blight Resistance in Potato Genotype SD20. International Journal of Molecular Sciences, 19: 1728
- Zhang, Z.-Y., Yang, L., Zhou, S.-F., Wang, H.-G., Li, W.-C. and Fu, F.-L. 2011. Improvement of resistance to maize dwarf mosaic virus mediated by transgenic RNA interference. *Journal of Biotechnology*, 153: 181-187.
- Zhang, Z., Li, X., Zhang, C., Zou, H. and Wu, Z. 2016. Isolation, structural analysis, and expression characteristics of the maize nuclear factor Y gene families. *Biochemical and Biophysical Research Communications*, 478: 752-758.
- Zhang, Z., Mao, B., Li, H., Zhou, W., Takeuchi, Y. and Yoneyama, K. 2005. Effect of salinity on physiological characteristics, yield and quality of microtubers in vitro in potato. *Acta Physiologiae Plantarum*, 27: 481-489.
- Zhu, S., Li, Y., Vossen, J.H., Visser, R.G. and Jacobsen, E. 2012. Functional stacking of three resistance genes against *Phytophthora infestans* in potato. *Transgenic Research*, 21: 89-99.

Received: 15 February 2019; Accepted: 27 June 2019

ΑΡΘΡΟ ΑΝΑΣΚΟΠΗΣΗΣ

Μοριακές πρόοδοι στη βελτίωση των γεωργικών καλλιεργειών για την κάλυψη των σύγχρονων απαιτήσεων στη γεωργία

Θ. Μαργαριτοπούλου και Δ. Μηλιώνη

Περίληψη Ο ηλίανθος, ο αραβόσιτος και η πατάτα, είναι μεταξύ των σημαντικότερων καλλιεργειών στον κόσμο. Προκειμένου να βελτιωθούν διάφορα χαρακτηριστικά τους, οι καλλιέργειες έχουν υποστεί γενετική τροποποίηση σε μεγάλο βαθμό. Αν και οι μοριακοί δείκτες έχουν χρησιμοποιηθεί με επιτυχία για την ταυτοποίηση απλών χαρακτηριστικών, όπως η γονιμότητα, η ανοχή σε ζιζανιοκτόνα ή η αντίσταση στα παθογόνα, σημαντικά αγρονομικά χαρακτηριστικά, τα οποία είναι πολύπλοκα και πο-

σοτικά, όπως η απόδοση, η αντοχή σε συνθήκες στρες από βιοτικούς και αβιοτικούς παράγοντες και η ποιότητα του σπόρου, παραμένουν μία πρόκληση και απαιτούν προσεγγίσεις που περιλαμβάνουν τη μελέτη ολόκληρου του γονιδιώματος. Γενετικό υλικό για αυτές τις καλλιέργειες διατηρείται σε τράπε-ζες σε παγκόσμια κλίμακα και αντιπροσωπεύει πολύτιμους πόρους για τη μελέτη σύνθετων χαρακτηριστικών. Σήμερα, οι τεχνολογικές εξελίξεις και η δυνατότητα αλληλούχησης ολόκληρων γονιδιωμάτων έχουν καταστήσει εφικτές νέες προσεγγίσεις στο επίπεδο του γενώματος. Η μοριακή βελτίωση, συμπεριλαμβανομένων τόσο των διαγονιδιακών μεθόδων όσο και της βελτίωσης με τη βοήθεια γενετικών δεικτών, διευκόλυνε την ταυτοποίηση δεικτών για γενετικούς χάρτες υψηλής πυκνότητας και επέτρεψε μελέτες συσχέτισης ολόκληρου του γονιδιώματος και τη γονιδιακή επιλογή στον ηλίανθο, τον αραβόσιτο και την πατάτα. Η επιλογή μέσω γενετικών δεικτών σχετιζόμενων με τις αποδόσεις υβριδίων έχει δείξει ότι η γονιδιωματική επιλογή είναι μια επιτυχημένη προσέγγιση για την αντιμετώπιση σύνθετων ποσοτικών χαρακτηριστικών και μπορεί να διευκολύνει την επιτάχυνση των προγραμμάτων αναπαραγωγής σε αυτές τις καλλιέργειες στο μέλλον.

Hellenic Plant Protection Journal 12: 39-60, 2019	