Case Report

Internal transcribed spacer sequence based molecular confirmation and drug efficacy assessment against *Toxascaris leonina* (Linstow, 1909) infection in Asiatic lions (*Panthera leo persica*)

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**Summary**

The eggs recovered during faecal screening of Asiatic lions (kept at MC Zoological Park, Punjab, India) were delineated as *Toxascaris leonina* eggs based on morphometric and molecular studies (polymerase chain reaction targeting internal transcribed spacer sequences). Therapeutic management with fenbendazole @10 mg/kg body weight, once daily orally for three consecutive days proved ineffective with maximum faecal egg count reduction (FECR) on day 3 post treatments (69.35%). But, therapeutic intervention with extended period dose schedule (5 consecutive days) with fenbendazole (@10 mg/kg body weight) proved effective and showed a maximum FECR of 95.34% at day 7 post treatments. But, when ivermectin (@100 μg/kg body weight) was given orally on three alternate days, proved effective as FECR of 95.74% was recorded at day 7 post treatments. Thus, present study highlights the molecular confirmation of *T. leonina* and its management using fenbendazole and ivermectin in Asiatic lions.

**Keywords**: Asiatic lion; fenbendazole; ITSs; ivermectin; *Toxascaris leonina*

**Introduction**

*Toxascaris leonina*, an ascarid nematode targets domestic and wild felines and canines as definitive hosts (Okulewicz et al., 2012). Various reports had also been documented for its concurrent infections with other ascarids including *Toxocara cati* (Labarthe et al., 2004; Dalimi et al., 2006; Dubna et al., 2007; Reperant et al., 2007; Itoh et al., 2011) and *T. canis* (Reperant et al., 2007) in domestic and wild carnivores. The parasite possesses significant zoonotic potential and larvae of *T. leonina* can invade the tissues of laboratory animals (Despommier, 2003). There is only one report of Beaver & Bowman (1984) which described a larva of *T. leonina* from the eye of a child in East Africa.

The specific identification of the helminth parasites of wild felids and canids, belonging to family ascarididae is a prerequisite for studying their infection, population biology, systematics and epidemiology (Okulewicz et al., 2012). But, the morphological identification of the eggs from the faecal samples corresponding to a particular species is cumbersome job, which requires skill, experience and expertise (Moudgil et al., 2014a). The only characteristic which helps in differentiation of the eggs of *Toxocara* species from *T. leonina* is that egg shells of *Toxocara* species are pitted with different pit dimensions, but the eggs of *T. leonina* are smooth walled with translucent appearance (Okulewicz et al., 2012).

Recently, molecular techniques have provided alternative and apt approaches for the identification of eggs and adults of ascarid species. Various studies have demonstrated that the first (ITS-1) and second (ITS-2) internal transcribed spacers (ITS) of nuclear ribosomal DNA (rDNA) act as reliable genetic markers for the identification of *T. canis, T. cati* and *T. leonina* in domestic and wild animals (Zhu et al., 2000; Li et al., 2007). Various reports of *T. leonina* infection in captive wild felids includ-
ing lions, Bengal tigers, jaguars, pumas and lynx from various zoo-
logical gardens had been documented from different parts of world
from time to time (Okulewicz et al., 2002; Okulewicz et al., 2012).
The therapeutic interventions for the control of T. leonina infection
in wild fields had proved difficult and reoccurrence of infection was
recorded after some time (Okulewicz et al., 2012). Okulewicz et al.
(2012) also proposed that elimination of T. leonina and Toxocara
species from the zoo environment is very difficult. Thus, the pres-
ent study was envisaged with the objective of morphological and
PCR-based molecular confirmation of T. leonina targeting internal
transcribed spacer regions (ITSs) and therapeutic management
with fenbendazole and ivermectin to control the ascarid infection.

Material and Methods

Collection of the samples
The faecal sample of 3 lions (2 males and 1 female), kept at MC
Zoological Park, Chhatbir, Punjab, India were screened seasonally
(from January 2013 to September 2014) at regular basis for any
parasitic infestation. All the lions were kept individually in sepa-
rate enclosures with wooden floors. The faecal samples of all the
lions were collected carefully from the floor in separate polythene
bags (with individual animal identity on it) and were transferred to
the laboratory at the Department of Veterinary Parasitology, GAD-
VASU, Ludhiana (India) at 4 °C for retrieval of parasitic eggs to
carry out morphological and molecular analysis.

Coprological examination and morphological analysis
Before starting the processing, the faecal sample was divided into
two aliquots, one for morphological and other for molecular anal-
ysis. The morphological examination was performed with light mi-
croscopy after concentrating the eggs by Sheather’s sugar floata-
tion technique (Soulsby, 1982). The egg per gram (EPG) values to
assess the intensity of infection was determined by McMaster
technique (Soulsby, 1982) with faecal egg count sensitivity of 50
eggs. The morphometric analysis targeting the size of the eggs
was performed as per Kazacos & Turek (1983). The length and
breadth of the eggs (n = 10) was depicted in micrometers and was
expressed considering mean ± standard deviation.

Genomic DNA extraction from eggs
The aliquots of the faecal material collected before and after de-
worming were subjected to Sheather’s sugar floatation technique
as described above. Approximately 50 μl of the supernatant con-
taining concentrated material was aspirated using micropipettes
and was transferred to 1.5 ml eppendorf vials (Traversa et al.,
2004). Individual sample was then introduced to repetitive freeze-
thaw process to induce complete disruption of the parasitic eggs.
Genomic DNA from the eggs was extracted by using QIAamp DNA
Stool Mini Kit (Qiagen, Hilden, Germany) by following manufactur-
er’s protocol. Finally, DNA was eluted by using 100 μl of elution
(AE) buffer.

Primers used and PCR Procedure
For molecular confirmation, the PCR protocol of Li et al. (2007)
was followed, targeting ITS-1 and ITS-2 ribosomal DNA (rDNA)
genetic markers by using the forward (5'-ATATCGGAAAGGAC
GCA CA-3') and reverse (5'-TTATGTTCTTCTCGCTCGCT-3')
primers. The PCR protocol for amplification of the templates was
performed in a 25 μl total reaction volume (containing 12.5 μl of
premix, 1.5 μl of each forward and reverse primers, 1 μl of MgCl2,
3.5 μl of nuclease free water and 5 μl of DNA) with following condi-
tions in sequential order: (i) Initial denaturation (94 °C for 5 min.),
(ii) Denaturation (35 cycles of 94 °C for 30 sec.), (iii) Annealing (35
cycles of 57 °C for 30 sec.), (iv) Extension (35 cycles of 72 °C for
1 min.), and (v) Final extension (72°C for 5 min.). For complete
confirmation of the eggs belonging to the same parasite, the eggs
retrieved from the faecal sample at day 1 post deworming were
also used for DNA extraction and PCR amplification process. The
DNA from the eggs of T. canis was also run simultaneously as
dNA to control the presence of any other related species.

Therapeutic management
The animals targeted in the present study were not dewormed reg-
ularly. The anthelmintics used earlier for the treatment of ascarid
infections in Asiatic lions of MC Zoological Park were ivermectin
and pyrantel pamoate (Singla et al., 2003; Singh et al., 2006). But,
there was reappearance of eggs in the treated hosts after a month
of treatment. Thereafter the animals were treated for ascarid infec-
tions only with fenbendazole (due to its wide safety margin), but
the efficacy of the drug was not assessed in the wild captive ani-
mals. In the present study, the therapeutic management was car-
rried out thrice (orally in the meat) during the study period i.e. from
January, 2013 to September, 2014. For the very first time (March,
2013) the treatment was given with fenbendazole @10 mg/kg
body weight, once daily for three consecutive days. Therapeutic
intervention with extended period dose schedule (5 consecutive
days) with fenbendazole (@10 mg/kg body weight) was carried out
during October, 2013. But, ivermectin @100 μg/kg body weight,
once daily for three alternate days was used for therapeutic man-
agement during the third treatment schedule i.e. in March, 2014.

Assessment of drug efficacy
The faecal egg count (FEC) is the most widely performed test to
detect drug efficacy (Cabaret & Berrag, 2004) and had been used
in captive-wild animals (Goossens et al., 2005; Young et al., 2000).
Thus, FEC was employed on the faecal samples collected from
Asiatic lions at day 1, 3, 7 and 21 post-treatment by using the
formula of Young et al. (2000).

Percent efficacy = \frac{(\text{Pre-treatment mean EPG} - \text{Post-treatment mean EPG}) \times 100}{\text{Pre-treatment mean EPG}}
Molecular confirmation

With the optimized cycling conditions, the DNA extracted from the faecal samples of the Asiatic lions before and after deworming, amplified a product of approximately 380 base pairs (Fig. 2) confirming the infection of T. leonina only, which was almost in concordance with the findings of Li et al. (2007). No product was amplified from DNA extracted from T. canis eggs, determining the specificity of the primers.

Anthelmintic efficacy assessment

Anthelmintic efficacy against T. leonina in three lions was evaluated using a benzimidazole group drug, fenbendazole and a macrocyclic lactone, ivermectin. Fenbendazole used for three consecutive days schedule was proved ineffective to eliminate the infection in lions and showed a maximum egg reduction to 69.35 % at day 3 post treatments (Table 1). The reason could be inappropriate drug dosage given to the animals due to unclear picture of the actual body weights of the animals under study. Fenbendazole used for extended period of time during second treatment period and ivermectin used during third treatment period proved effective with 95.34 and 95.74 % of egg reduction in lions, respectively (Table 1). Ivermectin had earlier been also used by Singla et al. (2003) to check piperazine resistant toxocarosis in lions at MC Zoological Park, Chhatbir, Punjab. But, in the present study the egg per gram values again increased significantly to 516.67 ± 102.07 and 100 ± 32.28 (P<0.05), respectively after treatment II and III on day 21 post treatments (Table 1). This could be attributed to the housing conditions of the zoo animals, especially the wooden floors where the faecal material remained clogged in the wooden planks. The eggs survived for a longer time even under harsh conditions, contributing to generation of a perpetual source for the transmission of infection (Bowman, 1999; Singh et al., 2006). Due to the direct life cycle of T. leonina and development of infective stages inside the eggs within a shorter period (around 9 days) favour the earlier completion of the life cycle (Moudgil et al., 2014b). This could be considered as a reason for persistence of T. leonina infection in well sanitized cages (Bowman, 1999; Singh et al., 2006). The similar findings were observed by Singh et al. (2006) and Dehuri et al. (2013) while carrying out the assessment of efficacy of pyrethral pamoate and ivermectin, respectively, against T. leonina infection in lions. On the other hand, the pour-on administration of ivermec-

Table 1. Drug efficacy assessment against T. leonina infection in Asiatic lions (n = 3).

<table>
<thead>
<tr>
<th>Animals</th>
<th>EPG Pre treatment</th>
<th>Post treatments (FECR %)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day 1</td>
<td>Day 3</td>
</tr>
<tr>
<td>Treatment I</td>
<td>1033.33 ± 251.66</td>
<td>533.33 ± 152.75(48)</td>
</tr>
<tr>
<td>Treatment II</td>
<td>1433.33 ± 251.66</td>
<td>783.33 ± 86.60(46)</td>
</tr>
<tr>
<td>Treatment III</td>
<td>783.33 ± 175.59</td>
<td>366.67 ± 104.08(53)</td>
</tr>
</tbody>
</table>

Values expressed as Mean ± Standard deviation

‘n’ indicates number of animals
Different ‘superscripts’ indicate values varying significantly at P < 0.05.

Statistical analysis

One-way analysis of variance (ANOVA) using Statistical Package for the Social Sciences (SPSS) 20.0 software was applied on eggs per gram values on different days (1, 3, 7 and 21) post treatment for the assessment of drug efficacies.

Results and Discussion

Morphological studies and parasitic intensity assessment

On detailed microscopic morphological examination, translucent ascarid eggs recovered were round to oval in shape with smooth walls (Fig. 1). Detailed morphometric studies (n=10) revealed the size of the eggs to be 81.78 ± 5.90 μm (74.8 – 86.2 μm) × 65.8 ± 4.90 μm (57.0 – 71.2 μm) (length × breadth). The results were in concordance with the findings of Gonzalez et al. (2007) and Okulewicz et al. (2012), who reported T. leonina eggs with smooth walls and without any albuminous coat. The lions showed high egg per gram values (1033.33 ± 251.66, 1433.33 ± 251.66 and 783.33 ± 175.59) for T. leonina eggs before the therapeutic interventions during the three treatment schedules, which was further reduced with appropriate anthelmintics used (fenbendazole and ivermectin).

Fig. 1. Photomicrograph of unembryonated Toxascaris leonina egg.
tin in dogs against T. leonina infection proved 98.54 % effective even after 28 days of treatment (Khayatnouri & Garedaghi, 2012). The reason could be attributed to the fact that actual body weight of dogs could easily be assessed for preparing the total dosage of anthelmintic, whereas it could not be assessed accurately in case of wild animals. The vehicle of drug to captive wild animals (especially wild felines) plays an important role in administering exact dosage, as in order to cover the wastage of drug a relatively higher dosage is given in certain cases and overconsumption of such drugs over and again may lead to toxicity or development of anthelmintic resistance. In order to prevent toxicity of drugs in case of precious wild animals kept at zoological gardens, relatively lower dosage regimen is generally followed, which is a limiting factor in exact assessment of drug efficacy and may further lead to development of anthelmintic resistance due to under and repetitive dosing of drug.

The management conditions should be given utmost importance in Zoological Parks as they play an important role in controlling the ascarid infections, because parasitic eggs possess the potential to withstand the harsh environmental conditions and also it is difficult to remove T. leonina and Toxocara species from the zoo environment due to passage of large number of eggs everyday by the infected individuals. The present findings associated with the effective ascarid treatment with fenbendazole over an extended dosage schedule, involving repeated dosing highlighted the first case of fenbendazole resistance in Asiatic lions. Coles et al. (2001) mentioned that resistance against benzimidazoles develop in stages and in the initial phase the individual parasites respond to repeated doses of the drug, as was observed in present study. Although, ivermectin exhibited more than 95 % faecal egg count reduction on day 7 post treatments, but also at day 21 post treatments there was recorded slight resistance against the drug. The observation could be attributed to the management conditions such as wooden floors of the enclosure. Further, a combination of drugs (avermectins and benzimidazoles) or alternative anthelmintics such as moxidectin could be adopted for successful treatment of the ascarid infections in precious wild animals. Further studies on this aspect are warranted.

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References


