

Comparison of the histological methods in the diagnostic of deer cysticercosis

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Summary

Histochemical methods for the detection and diagnosis of the developmental stages of the canine tapeworm, from the genus *Taenia* found in the heart and lungs of red deer (*Cervus elaphus*) and roe deer (*Capreolus capreolus*) hunted in Eastern Slovakia, is presented here. Detailed morphology of cysticerci (*Cysticercus* spp.), based on microscopic and histochemical analysis is described. For confirmation and demonstration of PAS-positive substances in the body of parasitic tissue (tegument and mesenchyme) the McManus - PAS method was used. The histochemical method according to Van Kossa was very effective for confirmation of calcareous corpuscles, which are one of the most important histological markers of cestode tissues (larva or adult).

Key words: cysticercosis; deer; histochemistry; histology

Introduction

Cysticercosis is a worldwide parasitic infection of domestic and wild animals and humans. Cysticercus, a larval stage in the cestode life cycle which is a fluid-filled cyst containing an attached single invaginated scolex, typically found in intermediate hosts. In the present study we have attempted to give a more emphasis on cysticercosis specifically in cervids. Canidae, mainly red fox and wolf, serve as final hosts. Free-living ruminants, like cervids while grazing pick up eggs and act as intermediate hosts of various tapeworms. In cervids, especially in roe deer, cestode larvae are often detected in the musculature (myocardium, and skeletal muscle), lung, liver and brain. The larval stages are of great importance than their adult stages, due to their pathogenesis. The histopathological changes frequently noticed are: atrophy, necrosis, caseation or calcification of muscle tissue.

Cysticercus cervi, the larval stage of the carnivore ta-

peworm, *Taenia cervi* (Christiansen, 1931) has been repeatedly considered synonymous to larval stages of *Taenia krabbei* Moniez, 1789 or *Taenia ovis* Cobbold, 1869 (Murai & Sugár, 1979; Verster, 1969). It is not possible to differentiate between *Taenia ovis* and *T. krabbei* on morphological grounds with certainty. For differential diagnosis, several biological characteristics were used. Red deer have been reported to be refractory to *Taenia ovis* infection whereas other potential intermediate hosts like cattle, goats, pigs and sheep have been shown to be refractory to *T. krabbei* (Flueck & Jones, 2006).

The incidence of cysticercosis in cervids was recorded in Yugoslavia (Rukavina *et al.*, 1973), Czech Republic (Kolář *et al.*, 1978), Russia (Romanenko, 1988) and Poland (Tropilo *et al.*, 2001). Information of the incidence of *Cysticercus* spp. in deer in Slovak Republic are incomplete and sporadic (Mituch, 1969; Kotrlý & Kotrlá, 1977).

In case of suspected pathoanatomic medical finding it is necessary to confirm degenerating cysticerci by direct histopathological examination. In our study we described the histopathological diagnosis of tissue infestation by cysticerci in red and roe deer. These methods emphasize structures typical for larval stages of cestodes, i.e. cysticercus, designated like the most important histological markers. The aim of present study is the evaluation of various basic, special and histochemical staining methods for diagnosis of deer cysticercosis.

Material and methods

Animals and experimental design

During standard field necropsies, six cysticerci were obtained from the hearts, localized on pericardium and under the epicardium of a two-year-old male red deer (*Cervus elaphus*) and a three-year-old female roe deer (*Capreolus capreolus*). From the lungs of a four-year-old male roe

deer three cysticerci were collected, which were localized under the visceral pleura in the lung parenchyma. All collected cysticerci were viable, well-developed with transparent envelopes filled with transparent fluid and contained an invaginated scolex. All animals included in the experiment were hunted in the Eastern Slovakian region. During the autopsy, special attention was given to the anatomical changes in cardiovascular and respiratory systems.

Table 1. Survey of various histological staining methods with microscopic results

Tissue components	Hematoxylin & Eosin	Goldner's Green Trichrome	Masson's Blue Trichrome	Van Gieson Picrofuchsin	PAS	Van Kossa
nucleus	violet	blue	blue/black	brown/black	—	—
cytoplasm	pink	pink	red	yellow	—	—
erythrocyte	red	orange	pink/red	—	—	—
muscle tissue	red/pink	purple/red	red	yellow/brown	—	—
collagen fibres	pink	green	blue	red	—	—
elastic fibres	—	—	red	yellow	—	—
polysaccharide (e.g. glycogen)	—	—	—	—	pink/red	—
calcium	—	—	—	—	—	brown/black

Histological investigations

Tissue samples with cysticerci obtained from the heart and lungs were fixed in 10 % neutral buffered formalin and subjected to routine paraffin processing. 180 – 200 histological sections (5 – 6 µm thick) per cysticercus were prepared. Some of these sections were stained with hematoxylin-eosin (H&E) as described Čunderlíková and Balážová (1990).

One quarter of total number of sections was stained with special staining methods: Goldner's modification of green trichrome (GGT), Masson's blue trichrome (MBT), and Van Gieson's picrofuchsin (VGP) (Vacek, 1995). The GGT and MBT determined the distribution of muscle, collagen fibres, fibrin and erythrocytes. VGP staining method also demonstrated collagen.

The histochemical methods for evidence of calcium accor-

ding to Van Kossa (VK) and Periodic Acid Schiff (PAS) method according to McManus for evidence of carbohydrates were used in about 10 – 15 of histological sections (Čunderlíková & Balážová 1990) (Tab.1). The classic staining method for the demonstration of calcium and certain other salts in tissues is that of VK method. The PAS staining is used to demonstrate carbohydrates (glycogen, glycoprotein, proteoglycans). It is used to distinguish diffe-

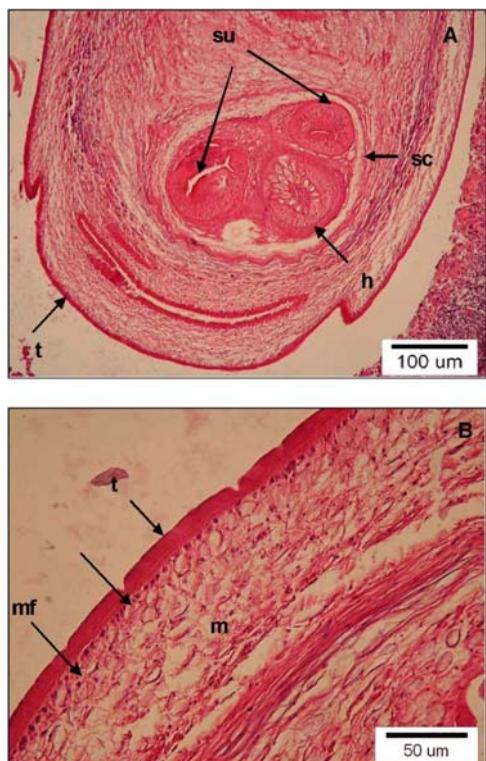


Fig.1. The neck region of cysticercus. Tegument (t), invaginated scolex (sc), suckers (su), hooklets (h), basal layer (bl), muscle fibres (mf), mesenchyme (m) (H&E, heart, A - 200x, B - detail, 400x)

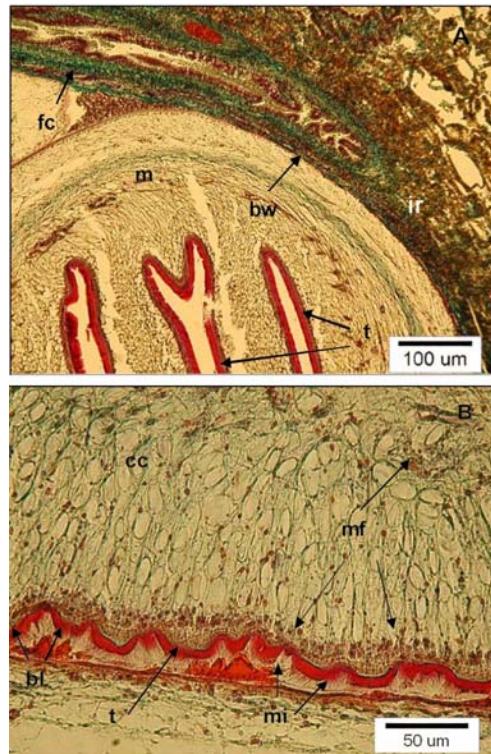


Fig.2. Goldner's green trichrome staining method. Bladder wall (bw), fibrous capsule (fc), inflammatory reaction (ir), tegument (t), mesenchyme (m), microvilli (mi), basal layer (bl), muscle fibres (mf), calcareous corpuscles (cc) (lungs, A - 100x, B - detail, 400x)

rent types of glycogen storage pathological processes in tissues. Sections utilizing all these staining methods were dehydrated in alcohols, cleared in xylene, and mounted in Entellan (Merck).

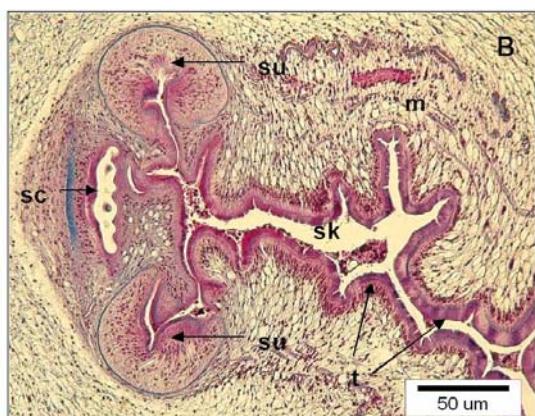
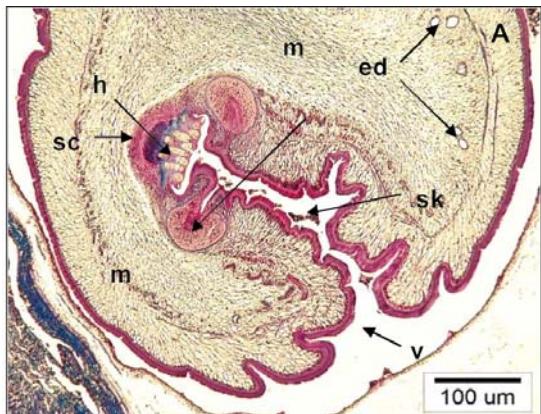


Fig.3. Masson's blue trichrome staining method. Vestibule (v), spiral canal (sk), scolex (sc), suckers (su), hooklets (h), mesenchyme (m), excretory ducts (ed), fibrous capsule (fc) (lungs, A - 100x, B - detail, 400x)

Histological sections were viewed with a light microscope under x100, x200 and x400 magnifications in conjunction with Olympus SP350 and Olympus BX50 camera. Images were then analyzed using DP-Soft version 3.1 software.

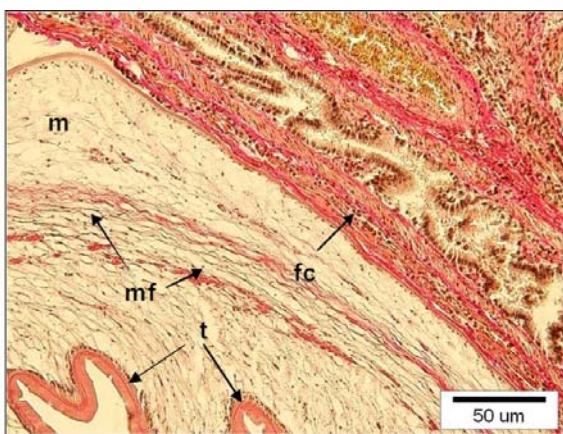


Fig.4. Van Gieson's Pikrofuchsin staining method. Fibrous capsule (fc), tegument (t), mesenchyme (m), muscle fibres (mf) (lungs, 200x)

Results

In the series of histological sections stained with H&E, mature viable vesicles of *Cysticercus* spp. with completely developed invaginated scolex were found. The cysticerci were spherical or oval in shape, approximately pea size (5 x 8 mm), whitish, and were filled with a watery transparent fluid.

In our histological sections, cysticerci with typical morphological signs of viable cysticerci, scolex and neck were recorded. The scolex was solid, bearing four suckers and rostellum with two rows of hooks, filled with mesenchyme, which is typical for *Cysticercus* spp. The scolex itself, and the neck area, were covered by tegument, forming numerous primary and secondary folds. Thick layer of tegument without microvilli, the characteristic feature of the neck area, was also observed. Moreover, all above histological findings were achieved in each staining method in all collected viable cysticerci from red or roe deer.

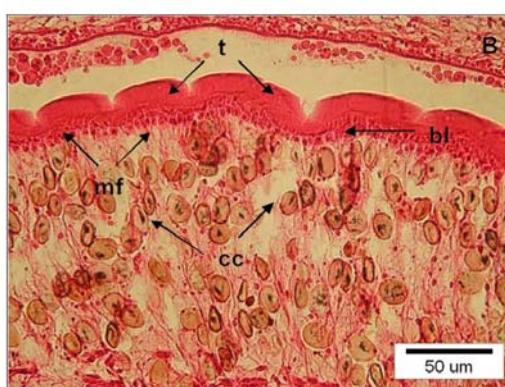
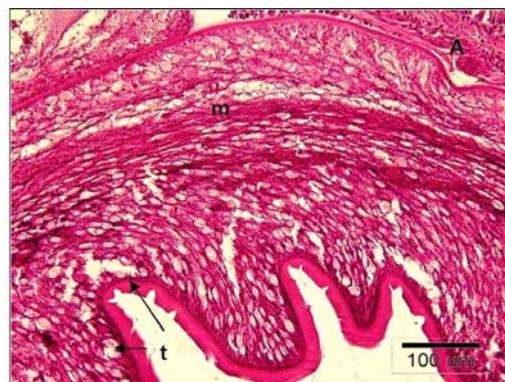


Fig.5. Various histochemical staining methods. Tegument (t), mesenchyme (m), calcareous corpuscles (cc), basal layer (bl), muscle fibres (mf) (A – PAS-method, lungs, 200x, B – Van Kossa method & nuclear red, heart, 400x)

Cysticerci were distinctly marked by a fibrous capsule from the surrounding tissues. Sometimes the section comprised a fibrous wall, a fully distended bladder membrane, or more often collapsed against the scolex. Leukocytes consisting mostly of lymphocytes, plasma cells, eosinophils and sporadic isolated macrophages infiltrated the tissues. Histologically, a slight inflammatory reaction was

observed in the surrounding tissues. The intermediate host tissue formed by the fibrous capsule was composed exclusively of parallelly arranged collagen fibres with sporadic infiltration of leukocytes. The superficial part of the bladder did not enter into the spiral canal, but it extended and formed the spiral vestibule. The vestibule was separated by the parenchymal part and the spiral canal ended with a tightly invaginated scolex leading into the vestibule. (Fig 1, 3).

The special histological staining methods serve especially for distinguishing the components of connective tissue (GGT, MBT & VGP). They are important for differentiating dense regular connective tissues (collagen fibres - the host fibrous capsule) from elastic fibres (total absence in the parasite body) and muscular tissue (muscle fibres and suckers). These staining techniques are very effective for photo documentation as well. (Fig. 2, 3, 4, 5).

The bladder wall of cysticercus consists of several different layers. The outer surface was composed of a thin homogenous eosinophilic and PAS-positive layer tegument (neodermis). Numerous microvilli were projected from this dense layer (Fig. 2B). Under the tegument, a thin fibrous basal layer as well as the muscles and thin connective tissue - mesenchyme with the excretory ducts were observed. Under the basal layer, and the layer of dense parallelly arranged spindle shaped muscle fibres, that passes continuously into the thin connective tissue, a mesenchyme was localized.

The mesenchyme is extremely rich in various carbohydrates (e.g. glycogen) and has PAS-positive reaction (Fig. 5A). In the mesenchyme loosely arranged tubular structures, the excretory ducts and numerous dark staining nuclei, without apparent cytoplasm of mesenchymal cell were observed. The rest of the body was filled by the mesenchyme that contained numerous specific calcareous corpuscles (calcospherules). The calcareous corpuscles are round or oval, usually composed of concentric layers of calcium, and thus stain positively with VK histochemistry method (Fig 5B).

Although it is not possible to differentiate cysticerci on morphological grounds obtained with histological methods, according several biological characteristics we suggest that the cysticerci may belong to *Taenia krabbei* (heart muscles) and *T. hydatigena* (lungs).

Discussion

Here we present a histopathological study of viable cysticerci from deer with a detailed description of their structure. Precise knowledge of specific structures of the viable cysticerci allowed diagnosis of dead cysticerci in histopathological detection (Slais, 1973). Identification of dead cysticerci is based on comparing similar histological characteristics, with knowledge that all stages of degeneration, disintegration, and calcification may occur. A marked inflammatory reaction around the parasite and the host tissue could be found. Polymorphonuclear cells predominate in recently prepared specimens, while granulocytes,

lymphocytes, and macrophages predominate in those dead for longer periods. Granulation tissue and fibrosis are observed in tissue, and Charcot-Leyden crystals may be present in biopsy and necropsy (Gutierrez, 1990).

In the histopathological study, Chacko *et al.* (2000) found that these calcified intracorporeal vacuoles are more resistant against degenerative and reparative processes in the tissue. The function of the calcareous corpuscles is unknown; however they dissolve easily in acid fixatives or acid hematoxylin. They are one of the most important histological markers of cestode tissue (larva or adult). The calcareous corpuscles are round or oval, usually composed of concentric layers of calcium, and thus stain positively with the VK histochemistry method. The persistence of calcareous corpuscles and/or hooklets may be the only recognizable structures in dead cysticerci. These ovoid bodies may occasionally be the sole parasitic remains seen in a biopsy; therefore their finding is pathognomonic for larval stages of tapeworms. In our study, we detected calcareous corpuscles with a selective histochemical technique, a Van Kossa (VK), which confirmed presence of calcium.

Other reports show that calcareous corpuscles are also formed in the excretory canals of cysticerci, indicating their possible contribution in the regulation of osmolarity (Mackiewicz & Ehrenpris, 1980) and the focal condensation of excessive calcium that protects the parasite from abnormal calcification (Vargas-Parada & Laclette, 1999). The number, size, shape, and the chemical composition of calcareous corpuscles vary between different cestode species, as well as within the same species (Brand *et al.*, 1969) and appear to be related to the composition of the micro-environment (Baldwin *et al.*, 1978).

Our histopathological findings were similar to descriptions of early stages of cysticercosis (Chi & Chi, 1978). Massive proliferation of epithelioid cells was absent in our findings. This was probably caused by the host tissue reactions (e.g. topographic localisation, immunological status and stages of parasitic development). A slight infiltration of leukocytes in the host tissue was also recorded (Kolář *et al.*, 1978).

Immunohistochemical and molecular methods for the diagnosis of larval stages of cestodes are more useful in human medicine for diagnosis of neurocysticercosis (Del Brutto *et al.*, 2001) and in veterinary medicine to diagnose bovine cysticercosis especially for *Cysticercus bovis* (Ogunremi *et al.*, 2004). Till to date for the diagnosis of cysticercosis in cervids no antigen specific antibodies are prepared because of high antigen variability in cestodes. Amongst histological staining methods used in the study, the H&E method is the most suitable histological method for differentiation of the basic basophilic and eosinophilic structures and for assessment of the presence of cellular infiltration. Histochemical methods are effective for confirmation and demonstration of PAS-positive substances in the body of parasitic tissue (tegument and mesenchyme – basic amorphous extracellular matrix) using the McManus - PAS method. The histochemical method according to

Van Kossa is very effective for distribution of calcium (calcareous corpuscles) and detection of calcification in host tissue in the late period of cysticercosis. The histopathological detection of cysticerci from biopsy is considered the most appropriate direct method for confirmation and diagnostics of the human and animal cysticercosis. This fact remains despite the presence of modern medical immunological (Enzyme-Linked Immunosorbent Assay - ELISA and EITB - Electro-Immuno Transfer Blot assay) and biophysical methods (Computer Tomography - CT and Magnetic Resonance Imaging - MRI). Histopathological methods are still the primary diagnostic methods used.

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