Effect of consecutive aerobic and resistance exercise on cortisol, immunoglobulin A, and creatine kinase responses in male students

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Summary

Study aim: To investigate the effects of consecutive aerobic and resistance exercise on Cortisol, Immunoglobulin A (IgA), and Creatine kinase (CK) responses of male students.

Material and methods: Eight subjects (mean age 21.0 ± 1.5 years) completed two trail, aerobic ergometer exercises (60% VO2max; 45-min) and resistance (80% 1RM; 45-min) exercise concurrently; each subject was his own control. Blood samples were collected pre-exercise, post-exercise, and 3 hours after exercise. One-way ANOVA with repeated measure and LSD post-hoc tests were used to evaluate changes in Cortisol, IgA, and CK during baseline and exercise periods.

Results: During baseline, there were no differences between exercise day (ED) values and those obtained at rest day (RD) for Cortisol, IgA, and CK. After exercise, Cortisol concentration in ED was significantly higher than RD (p<0.05); however, changes in IgA and CK responses were not significant.

Conclusions: This type of consecutive exercise didn't increase susceptibility to upper respiratory tract infection and muscle damage. Therefore, it can be useful for the preparation phase of training for athletes.

Key words: Immunoglobulin – Mucosal immune – Stress hormone – Consecutive exercises

Introduction

The immune system is influenced by physiological and psychological stressors [12], including physical activity. The effects of exercise on the immune system are mediated through interactions between the nervous, endocrine, and immune systems [35].

Salivary immunoglobulin A (IgA) provides a valuable defense against potential pathogens by preventing colonization and replication on the mucosal surfaces of the upper respiratory tract [22]. Secretory IgA as an important component of saliva is produced in local plasma cells. The secreted IgA prevents antigens and microbes from adhering to and penetrating the epithelium (immune exclusion), interrupts replication of intracellular pathogens during transcytosis through epithelial cells (intracellular neutralization), and binds antigens in the lamina propria, thus facilitating their excretion through the epithelium back into the lumen (immune excretion) [11,19]. Reduced concentrations or reduction in the release of Salivary IgA may allow for increased pathogenesis via the mucosal surface [34]. It has been shown that lower concentrations of IgA or chronic IgA deficiencies are associated with an increased frequency of upper respiratory tract infection (URTI) episodes [10,11], recurrent URTI [15], or reduced protection against certain infections [2].

There are reports demonstrating that IgA concentrations are suppressed in response to high intensity exercise [10,39] remain either unaltered [26,42] or are elevated in response to moderate-low intensity exercise [20]. However, several studies have also shown a stable secretion rate of IgA following tennis drills [31], a soccer match [5], or cycling [6]. Jemmott and McClelland [16] concluded from a meta-analysis of nine studies that the level of IgA secretion might indicate vulnerability toward URTI. Mackinnon and Hooper [24] further suggested that the protective effect might depend not only on IgA concentration, but also on salivary flow rate.

Although the physiological mechanisms underlying the temporary suppression of various aspects of the immune function after high-intensity exercise are still unclear, it is likely that both neural and endocrine factors influence the immune response to exercise [8,35]. Cortisol is the end product of the neuroendocrine stress response in humans. Although high levels of Cortisol have been demonstrated to inhibit antibody production in vitro [1],

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Material and Methods

Participants: Eight male physical education students took part in the study. They were not specifically endurance or resistance trained but were actively involved in various kinds of sports for 6-8 hours weekly on average. Characteristics of the subjects are reported in Table 1. Before the start of the study, subjects provided written informed consent. They were asked not to perform any strenuous exercise or consume medication for 2 days before each trial. Each subject was his own control. The protocol was approved by the Guilan University Ethics Committee.

Preliminary measurements: Maximal aerobic capacity (\(\text{VO}_{2\text{max}}\)) was estimated using the Fox protocol (heart rate response to 5 min of cycle ergometry at 150 W (6300 - 19.26 \times (HR5\text{min}) = \text{VO}_{2\text{max}}; [9]). Also, muscular strength was measured at the beginning of the study. Strength was assessed by one repetition maximum (1-RM) for bench press, leg press, leg extension, leg curl, lats pull-down, and biceps curl.

Following warm-up for the 6 resistance exercise testing, subjects selected a weight with which they felt they could complete three repetitions. At this weight, they only performed one repetition. Subjects then selected a weight with which they felt would be their 1-RM and attempted one repetition with this weight. Following successful attempts, weight was increased by 2.5 kg for subsequent 1-RM attempts. The 1-RM was usually reached in less than 6 sets, including the warm-up set. There was 3 min of rest between attempts, and two assistants changed the weight on the bar between attempts.

Procedures: The subjects completed trials in a counterbalanced order. For the afternoon exercise trial, subjects reported to the laboratory at 14:00 after fasting from 22:30 the previous day to 10:30 a.m. They then performed 45 min cycling at 60% \(\text{VO}_{2\text{max}}\) and 45 min 6 free-weight resistance exercise in the following order: bench press, leg press, leg extension, leg curl, lats pull-down, and biceps curl. Exercises consisted of 3 sets of 8 repetitions (80% 1-RM), with a 2-min rest between sets and a 3-min rest between exercises. The laboratory temperature and relative humidity were 21.4 ± 0.4°C and 64 ± 3%, respectively. Blood and saliva samples were taken during baseline, immediately after exercise, and 3 hours after exercise. Samples were refrigerated until the end of the session and centrifuged, then refrozen at -20°C, and stored until all sessions for each subject were completed.

Salivary cortisol concentrations were measured in duplicate by using a commercially prepared ELISA kit (Diagnostics Biochem Canada, Inc.) with modified procedures suggested by the manufacturer. A lower limit of detection for saliva cortisol was 1.0 ng/dl. CK was assayed spectrophotometrically through the use of commercially available kits (PARS AMUN CO.TEHRAHAN, IRAN). The CV for CK was < 4%. IgA was assayed by immunodiffusion (SRID) method by use of IgA kit (The Binding Site Ltd., Birmingham, UK). The CV for IgA was < 3%.

Table 1. Characteristics of subjects (mean ±SD)

<table>
<thead>
<tr>
<th>Variable</th>
<th>Male students (n=8)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>21.0 ± 1.51</td>
</tr>
<tr>
<td>Body height (cm)</td>
<td>178.0 ± 5.0</td>
</tr>
<tr>
<td>Body mass (kg)</td>
<td>74.0 ± 8.8</td>
</tr>
<tr>
<td>Body fat (%)</td>
<td>12.3 ± 3.6</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>23.3 ± 1.0</td>
</tr>
<tr>
<td>(\text{VO}_{2\text{max}}) (ml/kg/min)</td>
<td>46.2 ± 6.7</td>
</tr>
</tbody>
</table>
**Statistical analysis:** All results are presented as mean values and standard deviation of the mean. Normality of data distribution was assessed by the Kolmogorov-Smirnov test. Data were analyzed using a repeated measure ANOVA and LSD post-hoc test. Statistical significance was set at $\alpha = 0.05$.

**Results**

Results of the analysis showed that during baseline, there were no differences between exercise day (ED) and rest day (RD) for Cortisol, IgA, and CK. After exercise, Cortisol concentration in ED were significantly higher than RD ($p<0.05$); however, IgA and CK responses were not significant between ED and RD. Also, 3 hours after exercise, the Cortisol concentration in ED was significantly higher than in RD ($p<0.05$); but, IgA and CK concentration 3 hours after exercise was similar in ED and RD (Table 2). In addition, there were no correlations between IgA and Cortisol responses to exercise in both days.

**Discussion**

This study focused on IgA, CK, and cortisol response to exercise. Consecutive aerobic and resistance exercise protocol were designed to present physiological stress similar to the preparation phase of training in most sports.

Our results shows that mean salivary concentration of IgA were unaffected by consecutive aerobic and resistance exercise. These results are in line with previous studies [4,28,42]. Some studies have reported decreases in IgA concentration following high-intensity exercise [21,38]. Their exercise protocols were undertaken during a period of overtraining [24]. However, a significant elevation in IgA concentration has also been reported after high-intensity exercise [6]. This increase seemed to result from the reduction of saliva flow rate. Previous studies have suggested that there is an intensity-dependent effect of acute exercise on IgA, i.e. strenuous exercise is suppressive and moderate exercise has no effect [23]. Therefore, the inconsistency in responses of IgA secretion rate to exercise may be attributed to different types of sympathetic nervous system stimulation during exercise [14]. It seems that neither aerobic exercise nor resistance exercise at the same moderate exercise intensity evokes sufficient stimulation of the sympathetic nervous system or of the hypothalamic-pituitary-adrenal axis to modify IgA transcytosis.

We found that consecutive aerobic and resistance exercise was associated with increased salivary Cortisol concentrations immediately and 3 hours after exercise. Post-exercise Cortisol concentration changes seems to be affected by several mechanisms including stimulation of the sympathetic nervous system, stimulation of hypothalamic-pituitary-adrenal (HPA) secretion, increase of body temperature, changes in blood pH, hypoxia, lactate accumulation, and mental stress [30,37]. A study reported that Cortisol concentration increases due to continued exercises. These researchers reported that physical exercise could stimulate HPA, increase body temperature, increase Cortisol secretion, and the release of Cortisol from the carrier proteins [3]. Therefore, the high concentration of salivary Cortisol accompanied by increase of saliva viscosity is the indicator of sympathetic nervous system activation [3]. The findings of this study are in line with findings of Ben-Aryeh et al. [3] and Kaciuba-Uscilko et al. [17], who found that consecutive aerobic exercise and resistance exercise increase Cortisol concentration. Furthermore, the results of this study do not support the data given by O’Connor et al. [33] and Sari-Sarraf et al. [37]. The equivocal nature of these observations may be due to the difference in intensity, duration, and mode of exercise, the place of exercise, and the age of the subjects in each study.

We found a lack of correlation between IgA and Cortisol concentrations. Specifically, we observed that IgA concentration after consecutive aerobic and resistance exercise did not change, while Cortisol concentration changed. Our findings match those of Sari-Sarraf et al. [37], reporting that during heavy and moderate exercises Cortisol secretion had no relationship with the inhibition of salivary IgA.

**Table 2.** Mean values (±SD) of Cortisol, CK and IgA recorded pre-, post- and 3 hours post exercise during rest and exercise day in male physical education students

<table>
<thead>
<tr>
<th>Variable</th>
<th>Rest day</th>
<th>Exercise day</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Pre-exercise</td>
<td>Post-exercise</td>
</tr>
<tr>
<td>Cortisol (nmol/L)</td>
<td>1.26 ±0.3</td>
<td>0.67± 0.1</td>
</tr>
<tr>
<td>Creatine kinase (U/L)</td>
<td>200.1 ±12.5</td>
<td>196.2± 8.7</td>
</tr>
<tr>
<td>IgA (mg/dl)</td>
<td>72.2 ±8.3</td>
<td>67.4± 6.1</td>
</tr>
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Legend: IgA - Immunoglobulin A * Significantly (p<0.05) different from the rest day
When the subjects work at a fixed loading, CK is released from the muscle when the loading exceeds a certain limit of muscle ability [25]. Although numerous studies [18, 32, 40] have examined the effects of exercise on serum CK activity under various exercise conditions, to our knowledge no study has addressed CK release during consecutive aerobic and resistance exercise. In the present study, we found that consecutive aerobic and resistance exercise could not increase CK concentration. Previous studies reported that during exercise, muscle repeatedly contracts and uses energy substrates. When exercise intensity is within the normal range of metabolism, the muscle tissue is exercised without marked changes in membrane permeability. However, when exercise intensity exceeds this permissible range, the membrane permeability temporarily changes, resulting in CK release from the active muscle [25]. Rolffinger et al. [41] indicated the possibility of a threshold value for CK release, with a “distance” threshold for CK release.

Totsuka et al. [25] reported that CK response depends on individual fitness level, levels that are usually different between athlete and non-athlete. Our subjects were physical education students and maybe the intensity of consecutive aerobic and resistance exercise did not exceed the ability of their muscles, thus the CK concentration in serum did not increase due to consecutive aerobic and resistance exercise.

In conclusion, we showed that 90-min consecutive aerobic and resistance exercise led to increased salivary Cortisol, but there were no significant changes in salivary IgA and serum CK concentration. Therefore, we suggest that this type of consecutive aerobic and resistance exercise don’t show a wide range of deleterious effects on the immune system. Finally, it can only be stated that this kind of consecutive aerobic and resistance exercise was well tolerated; it has demonstrated that this sub-maximal exercise did not lead to muscle damage or altered salivary IgA levels.

References


Immune response to consecutive exercise


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