



EFFECT OF CAPONISATION ON BONE DEVELOPMENT IN NATIVE MALE CHICKENS*

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Abstract

The aim of the study was to determine the effect of caponisation on the morphometric traits and mechanical parameters of tibial and femoral bones in Greenleg Partridge cocks. The experiment involved 200 cocks. At the age of 8 weeks, 100 birds were subjected to surgical castration. At week 24, the birds were slaughtered and tibial and femoral bones were collected from 10 non-caponised cocks and 10 capons. The caponisation surgery had no effect on the weight and length of any of the long bones but resulted in reduction ($P \leq 0.05$) of the ash content in both bones and Ca in the femur. It also influenced the geometric structure of the bones, i.e. there was an increase ($P \leq 0.05$) in the second moment of inertia in the tibial bone and the cross-sectional area and mean relative wall thickness in the femoral bone of the capons. The three-point bending test revealed a negative effect of caponisation on the mechanical strength of the bone. Values characterising the highest bone material strength, i.e. yielding load (femur), maximum force moment (tibia) and yielding deformation, bending point resistance, and load-to-deformation (both bones), declined in the capons. The investigations demonstrated a negative effect of caponisation on the quality of long bones. The tibial bone seems to be slightly more sensitive to the caponisation effects than the femoral bone. It can be assumed based on the analysis of biomechanical traits that the bones of capons are more susceptible to deformations or fractures due to their modified geometry and mechanical brittleness.

Key words: caponisation, cocks, bone characteristic, Greenleg Partridge breed

*Study funded from Statutory Research of Institute of Animal Nutrition and Bromatology, University of Life Sciences in Lublin, no. ZIZ/DS-4.

The production of capons is deeply rooted in the local tradition of not only Asia but also Europe (Miguel *et al.*, 2008; Díaz *et al.*, 2010; Lin *et al.*, 2012; Guo *et al.*, 2015; Franco *et al.*, 2016). They are mostly produced from native or locally adapted breeds with a slow growth rate. Similarly, conservative breeds, i.e. Greenleg Partridge, Yellowleg Partridge, and Polbar are used for the production of capons in Poland (Gryzińska *et al.*, 2012, 2013; Kwiecień *et al.*, 2015, 2018; Sokołowicz *et al.*, 2016; Gesek *et al.*, 2017).

Castration is a hormonal intervention, which permanently influences metabolic processes in birds (Rikimaru *et al.*, 2011; Symeon *et al.*, 2013; Chen *et al.*, 2014). Androgen deficiency induces changes in birds' behaviour, and maturity regresses to the pre-caponisation stage (Chen *et al.*, 2005). Androgens exert strong anabolic effects in various tissues, stimulating muscle, bone, and connective tissue growth (Pederson *et al.*, 1999). Capons are usually reared for up to 6 months; hence, they must have a strong skeleton to support the growing body weight. As shown in many investigations, caponisation influences production results (Chen *et al.*, 2006 a; Murawska and Bochno, 2007; Calik *et al.*, 2015; Gesek *et al.*, 2017; Kwiecień *et al.*, 2015, 2018), blood parameters (Lin and Hsu, 2011; Cheng-Yung *et al.*, 2012), and bone quality (Mahmud *et al.*, 2014; Muszyński *et al.*, 2017; Tomaszewska *et al.*, 2017; Zawacka *et al.*, 2018).

Induced by various factors (chemicals, testectomy operation, or age), androgen deficiency has a negative effect on bone growth and development in humans (Manolagas *et al.*, 2002). In turn, the effects of castration on bone parameters in poultry are ambiguous. In poultry, unfavourable changes are most frequently observed in the tibial bone. Many studies have been focused on measurements of the length, weight, or outer surface of bones, but there are no reports on bone biomechanics and histology. Investigations conducted by Chen *et al.* (2014) demonstrated the impact of castration on the weight, length, and histological parameters of bones. Other studies (Muszyński *et al.*, 2017) showed lower breaking strength of tibiotarsal bones in capons. Similarly, Tomaszewska *et al.* (2017) demonstrated that caponisation reduced mineral density and mechanical strength of femoral bone. Reduced tibial bone breaking strength after castration was also reported by Chen *et al.* (2007), Chen *et al.* (2014) and Lin *et al.* (2012). In turn, other studies did not show an effect of this treatment on bone parameters (Mahmud *et al.*, 2014). The age and sex of birds were found to have a significant impact on bone tissue parameters during postnatal development (Charuta *et al.*, 2012, 2013). A decline in the volumetric mineral density of tibiotarsal bones was demonstrated in 4-week-old broiler cocks (Charuta *et al.*, 2013) and in 9-week-old turkeys (Charuta *et al.*, 2012).

Sex steroid hormones have a substantial impact on bone metabolism; yet, the paracrine mediators of the androgen effect on bones are not fully known. There are not many reports on the effect exerted by androgens on the quality of bones in native breeds in Poland (Muszyński *et al.*, 2017; Tomaszewska *et al.*, 2017). Since other studies report a reduction in the weight, length, or biomechanics of bones in capons in comparison with non-caponised roosters, the present study assumes that, through the decrease in testosterone levels, the castration treatment of Greenleg Partridge cocks may also reduce bone structural integrity, inhibiting bone development. To obtain information on this issue, the investigations were focused on determination

of the effect of caponisation performed at 8 weeks of age on the physicochemical, morphometric (geometric and cortical), and strength parameters of tibia and femur in Greenleg Partridge chickens.

Material and methods

All procedures used during the study were approved by the Local Ethics Committee for Animal Testing at the University of Life Sciences in Lublin, Poland (Approval No. 33/2013 of 16 April 2013).

Experimental design

The experiment was conducted on 200 Greenleg Partridge (old traditional Polish chicken breed) cockerels, which were individually weighed and labelled. One-day-old birds were weighed, marked with wing tags, and randomly distributed to pens. A hundred 8-week-old birds were surgically castrated by a qualified veterinarian in accordance with Commission Regulation (EC) No. 543/2008, as described in detail by Tomaszewska et al. (2017). Afterwards, the cocks were divided into two groups: control group – cocks (50 birds in 10 cages with 5 birds in each) and a caponised experimental group (50 birds in 10 cages with 5 birds in each). The birds were reared to 24 weeks of age, fed commercial diets *ad libitum*, and provided with continuous veterinary care. The feed mixes were prepared based on cereal meal middlings (maize, wheat, oat), post-extraction soya meal, and sunflower seeds (Table 1). Capons and non-caponised cocks were fed with standard complete feed mixes for general-purpose hens adequate for respective breeding periods (weeks 1–8, weeks 8–18, and over 18 weeks of life). The nutritive value of the diets is presented in Table 2. The birds were kept in the same zoohygienic conditions.

Table 1. Composition (g·kg⁻¹) of the diet fed during the trial

Ingredients (g·kg ⁻¹)	1–8 wk old	8–18 wk old	> 18 wk old
Corn	441.5	438.0	281.9
Wheat	200	200	200
Oat	50	100	200
Soybean meal	200	100	100
Sunflower meal	50	100	150
Soybean oil	24	25	20
Monocalcium phosphate	13	14	19
Limestone	7.6	8.9	15
NaHCO ₃	1.2	1.4	1.6
NaCl	2.7	2.7	2.5
Mineral-vitamin premix	10*	10**	10**

*1 kg of premix for a period of 1–8 weeks contained: vit. A 12 000 IU, vit. D₃ 2 500 IU, vit. E 25 mg, vit. K₃ 3 mg, vit. B₁ 2 mg, vit. B₂ 6 mg, vit. B₆ 5 mg, vit. B₁₂ 0.02 mg, nicotinic acid 30 mg, pantothenic acid 15 mg, folic acid 2 mg, biotin 0.2 mg, choline 700 mg, Fe 70 mg, Zn 60 mg, Mn 70 mg, Cu 8 mg, I 1 mg, Se 0.3 mg.

**1 kg of premix for a period of nine weeks from the end of the rearing contained: vit. A 10 000 IU, vit. D₃ 2 000 IU, vit. E 25 mg, vit. K₃ 2 mg, vit. B₁ 2 mg, vit. B₂ 4 mg, vit. B₆ 4 mg, vit. B₁₂ 0.02 mg, nicotinic acid 25 mg, pantothenic acid 15 mg, folic acid 1 mg, biotin 0.2 mg, choline 300 mg, Fe 50 mg, Zn 50 mg, Mn 60 mg, Cu 7 mg, I 0.7 mg, Se 0.2 mg.

Table 2. Nutrient composition of basal diet

Ingredients	1–8 wk old	8–18 wk old	> 18 wk old
ME (MJ kg ⁻¹)	11.20	11.30	11.00
Crude protein (%)	17.32	15.24	16.72
Crude fat (%)	4.00	4.00	3.40
Crude fibre (%)	5.50	6.20	7.15
Crude ash (%)	4.45	4.52	5.00
Na (%)	0.18	0.18	0.17
Ca (%)	1.00	1.30	4.00
Available P (%)	0.35	0.32	0.40
Lysine (%)	0.80	0.55	0.88
Methionine (%)	0.30	0.25	0.35

At the age of 24 weeks, 10 birds were selected from each group and slaughtered. During simplified dissection, tibial and femoral bones of the left leg of the birds were collected. Before slaughter, the birds did not receive feed but had permanent access to water. All birds were clinically healthy.

Bone collection and analysis

Carcasses were chilled for ca. 18 h at a temperature of 4°C. Afterwards, tibias and femurs were collected, cleaned from soft tissue remnants, and subjected to basic measurements (bone weight and length). Relative bone weight was calculated from the ratio of bone mass to body weight. Based on the measurements, the bone density index was determined showing changes in bone mineralisation calculated as bone weight (in mg) to length (in mm) (Ziaie *et al.*, 2011). This simple bone density index was introduced for the first time by Seedor *et al.* (1991). The higher the bone weight/length index, the denser is the bone and indicates changes in bone mineralization. After the measurements, the bones were wrapped individually in gauze soaked in physiological saline and kept frozen at a temperature of –25°C for further analysis.

Bone morphometric parameters

The measurements of the external and internal horizontal and vertical diameter of the bone shaft cross-section at the fracture site performed with the use of a digital calliper were used for calculation of the following geometric parameters: cross-sectional area, the second moment of inertia and the mean relative wall thickness, thickness of the cortical layer, cortical surface, cortical index, and cortical parameters: the cortical surface index (Ferretti *et al.*, 1993; Kwiecień *et al.*, 2016; Tomaszewska *et al.*, 2016).

Bone mechanical properties

The mechanical properties were determined after 3-hour thawing at room temperature. The three-point bending test of bone mid-diaphysis was performed on

a Zwick Z10 universal testing machine (Zwick/Roell, Ulm, Germany). Prior to the analysis, the bone was placed horizontally on two rounded support bars. The distance between the supports was set in each case at 40% of the total bone length. The load was applied in the anterior-posterior plane of bone with a displacement rate of 10 mm·min⁻¹ until fracture. The maximum elastic strength and the ultimate strength were determined as described previously (Ferretti et al., 1993; Kwiecień et al., 2016; Tomaszewska et al., 2016). Based on the measurements, the strength parameters of bones were determined: the value of deflection, bending point resistance, and load-to-deformation ratio (Ferretti et al., 1993; Kwiecień et al., 2014). Additionally, the value of strength parameters of the tibial and femoral bones as well as the physical properties of the bone material determining its strength: yielding load towards bone weight, yielding load towards body weight, maximum force moment towards bone weight, and maximum force moment towards body weight were determined.

Mineral composition of bones

The femurs and tibias were defatted, dried to constant weight, and mineralised (AOAC, 2000). The Ca content was determined with atomic absorption spectrophotometry on the Unicam 939/959 apparatus, and total P was determined with the colorimetric method (Polish standard PN-76/R-64781, 1976) using the Helios a-Unicam apparatus with a molybdenum-vanadium reagent (NH₄VO₃, (NH₄)₆Mo₇O₂₄·H₂O, H₂O) and converted into mineral content in crude ash.

Blood analysis

At 24 weeks of rearing, the birds were fasted for 10 h before blood sampling and slaughter. Plasma for analysis of the biochemical parameters was obtained by centrifugation of whole blood at 3000 rpm (603 × g) for 15 min in a laboratory centrifuge (MPW-350R; MPW Medical Instruments, Warsaw, Poland) at a temperature of 4°C. Plasma without signs of haemolysis was analysed within 4 h after sampling and the contents of testosterone, the activity of alkaline phosphatase, and the contents of Ca and P were determined. Assays for the concentrations of plasma testosterone were carried out with an ELISA microtiter reader (MRX Dynex Technologies, USA), using ELISA kits (NEOGEN Testosterone ELISA kit, USA). The activity of the indicator enzyme, i.e. alkaline phosphatase, and the contents of Ca and P were determined in blood plasma with colorimetric methods using reagent kits according to the manufacturer's protocol (BioMaxima, Lublin, Poland; Hydrex Diagnostics, Warsaw, Poland) and a random access biochemical analyser Metrolab 2300 GL (Metrolab SA, Buenos Aires, Argentina). The analysis procedures were verified with the use of multiparametric control plasma (BioCal) as well as control plasma with a normal level (BioNorm) and a high level (BioPath) of indices (BioMaxima, Lublin, Poland; Hydrex Diagnostics, Warsaw, Poland).

Statistical analysis

The results tests were statistically analysed using Statistica software ver. 10 (Statsoft Inc., Tulsa, USA). The normality of data distribution was tested using the

Shapiro-Wilk test. One-way analysis of variance was only used in the case of the assessment of the bone parameters, where the effect of the breed was examined:

$$y_{ijk} = \mu + ai + e_{ijk}$$

where:

y_{ijk} – k th observation from the i th and j th groups,

μ – mean value of the trait in the population,

ai – effect of the i th group,

e_{ijk} – error = effect related to individual variability and measurement error.

Significant statistical differences were set at the level of $P \leq 0.05$. The significance of the differences between mean values in the respective diets was assessed using Duncan's multiple range test.

Results

Physical properties and geometric characteristics of bones

The caponisation surgery was shown to exert an impact on the final body weight, which was significantly higher in the capon group (Table 3). It did not change the weight of both bones; however, given the change in the final capon body weight, the proportion of the tibial and femoral bones in the body weight, i.e. the relative bone weight, decreased ($P \leq 0.05$) in the capons (Table 3 and 4). The caponisation treatment did not change the femur and tibia length (Table 3 and 4). The value of the second moment of inertia was increased by approx. 14% ($P < 0.05$) in the femur (Table 4) and by approx. 7% in the tibia of the capons (Table 3), with simultaneous reduction of the values of traits indicating the content of cortical tissue in the femur, i.e. the thickness of the cortical layer and the cortical surface (Table 3). Additionally, a ca. 7% and 12% increase in the cross-sectional area and mean relative wall thickness parameters, respectively, was noted in the femurs of the capons, in comparison with the non-caponised birds (Table 4).

Mechanical properties of bones

The caponisation treatment also exerted an effect on the femur and tibia strength parameters (Table 3 and 4). It led to a reduction of bone yielding load by 8.5%, bending point resistance by 6.9%, yielding deformation by 28%, and load-to-deformation ratio by 17% in the capons' femur (Table 4). In turn, the value of the maximum force moment, bending point resistance, yielding deformation, and load-to-deformation ratio decreased by 12%, 25%, 12%, and 23%, respectively, in the tibial bone of the capons (Table 3). The value of the strength parameters of the bone material (i.e. yielding load towards body weight, maximum force moment towards bone weight, and maximum force moment towards body weight) in the femur was significantly lower ($P \leq 0.05$) in the capons (Table 3 and 4).

Table 3. The effect of caponisation on mechanical properties and dimensions as well as bone mineral density of tibia, obtained from 24-week-old Greenleg Partridge birds (mean±SE)¹

Item	Greenleg Partridge		P-value
	non-caponised cocks	capons	
1	2	3	4
Body weight (g)	1642 b±164.8	1808 a±129.8	0.022
Tibia weight (g)	14.4±1.33	14.7±1.36	0.697
Relative tibia weight (g/100 g of body weight)	0.88 a±0.05	0.81 b±0.08	0.037
Tibia length (mm)	94.5±3.91	94.5±4.09	0.995
Bone density index (mg×mm ⁻¹)	152.6±8.94	155.2±10.2	0.56
Yielding load (N×mm)	191.0±22.0	191.8±42.2	0.960
Maximum force moment (N×mm)	284.7 a±53.2	254.0 b±34.5	0.015
Bending point resistance (N×mm×mm ⁻²)	14.6 a±5.16	11.7 b±1.62	0.010
Yielding deformation (mm)	2.67 a±0.59	2.38 b±1.23	0.041
Load-to-deformation ratio (N×mm×mm ⁻¹)	91.8 a±32.9	74.5 b±17.6	0.016
Yielding load towards bone weight (N×mm×g ⁻¹)	13.3±2.85	13.2±2.22	0.900
Yielding load towards body weight (N×mm×1000×kg ⁻¹)	116.9 a±22.7	106.2 b±15.1	0.023
Maximum force moment towards bone weight (N×mm×g ⁻¹)	19.7 a±3.30	17.5 b±2.77	0.012
Maximum force moment towards body weight (N×mm×1000×kg ⁻¹)	172.7 a±23.0	140.9 b±17.3	0.003
Geometrical characteristics of the bone			
Horizontal internal diameter (mm)	5.47±0.66	5.41±0.48	0.831
Horizontal external diameter (mm)	7.72±0.57	7.79±0.50	0.770
Vertical internal diameter (mm)	4.80±0.41	5.07±0.45	0.185
Vertical external diameter (mm)	6.90±0.68	7.13±0.41	0.373

Table 3 – contd.

1	2	3	4
Second moment of inertia (mm ⁴)	98.4 b±33.6	105.0 a±25.9	0.043
Cross-sectional area (mm ²)	21.3±6.73	22.1±3.71	0.760
Mean relative wall thickness	0.44±0.17	0.43±0.08	0.900
Features of cortical bone			
Thickness of cortical layer (mm)	2.38 a±0.33	2.25 b±0.84	0.036
Cortical surface (mm ²)	31.4 a±5.09	29.5 b±10.5	0.043
Cortical index (%)	7.09±0.50	7.00±0.63	0.730
Cortical surface index (%)	60.4±7.84	59.3±8.28	0.770

¹ – Means of 10 replicates (samples), SE – standard error.

a, b – mean values in rows with different letters differ significantly at $P \leq 0.05$.

Table 4. The effect of caponisation on mechanical properties, dimensions as well as bone mineral density of femur, obtained from 24-week-old Polbar and Greenleg Partridge birds (mean \pm SE)¹

Item	Greenleg Partridge			P-value
	non-caponised cocks	capons		
1	2	3	4	
Femur weight (g)	11.9 \pm 1.11	12.0 \pm 0.71	0.867	
Relative femur weight (g/100 g of body weight)	0.73 a \pm 0.04	0.66 b \pm 0.04	0.002	
Femur length (mm)	50.6 \pm 3.12	51.0 \pm 2.74	0.738	
Bone density index (mg \times mm ⁻¹)	235.4 \pm 14.3	235.0 \pm 9.77	0.939	
Yielding load (N \times mm)	209.5 a \pm 48.9	193.0 b \pm 20.6	0.038	
Maximum force moment (N \times mm)	315.2 \pm 64.7	303.5 \pm 57.7	0.675	
Bending point resistance (N \times mm \times mm ⁻²)	12.3 a \pm 2.58	11.5 b \pm 3.39	0.042	
Yielding deformation (mm)	4.44 a \pm 1.52	3.46 b \pm 0.92	0.038	
Load-to-deformation ratio (N \times mm \times mm ⁻¹)	60.7 a \pm 21.6	51.7 b \pm 19.5	0.034	
Yielding load towards bone weight (N \times mm \times g ⁻¹)	16.3 b \pm 1.70	17.5 a \pm 3.70	0.036	
Yielding load towards body weight (N \times mm \times 1000 \times kg ⁻¹)	118.4 \pm 14.7	115.9 \pm 25.9	0.799	
Maximum force moment towards bone weight (N \times mm \times g ⁻¹)	25.5 \pm 4.27	26.3 \pm 5.16	0.709	
Maximum force moment towards body weight (N \times mm \times 1000 \times kg ⁻¹)	184.9 a \pm 29.5	174.3 b \pm 34.1	0.046	
Geometrical characteristics of the bone				
Horizontal internal diameter (mm)	5.96 \pm 0.70	6.13 \pm 0.41	0.522	
Horizontal external diameter (mm)	8.63 \pm 0.68	8.74 \pm 0.32	0.651	
Vertical internal diameter (mm)	5.44 \pm 0.70	5.56 \pm 0.64	0.689	
Vertical external diameter (mm)	7.85 \pm 0.66	7.67 \pm 0.28	0.456	

Table 4. – contd.

1	2	3	4
Second moment of inertia (mm ⁴)	140.8 b±18.9	160.7 a±55.5	0.029
Cross-sectional area (mm ²)	25.8 b±3.22	27.7 a±6.99	0.045
Mean relative wall thickness	0.41 b±0.08	0.46 a±0.15	0.039
Features of cortical bone			
Thickness of cortical layer (mm)	2.66±0.64	2.61±0.46	0.819
Cortical surface (mm ²)	38.9±9.65	38.7±6.59	0.967
Cortical index (%)	7.94±0.70	8.04±0.34	0.689
Cortical surface index (%)	74.4±11.7	76.0±5.61	0.706

¹ – Means of 10 replicates (samples); SE – standard error.

a, b – mean values in rows with different letters differ significantly at $P \leq 0.05$.

Composition of bone minerals

The ash content in both bones was significantly lower ($P \leq 0.05$) in the capons (Table 5). The caponisation surgery resulted in a 14% decline in the content of Ca in the femur ($P \leq 0.05$), whereas no effect of the treatment on the P content and the Ca:P ratio in both bones was detected (Table 5).

Table 5. Effect of caponisation on mineral composition of bones at 24 weeks (mean \pm SE)¹

Item	Greenleg Partridge		P-value
	Non-caponised cocks	Capons	
Tibia bone			
Bone ash (%)	43.6 a±2.43	40.6 b±1.53	0.004
Ca (g·kg ⁻¹)	527.9±47.0	512.2±14.5	0.327
P (g·kg ⁻¹)	185.1±7.73	192.2±12.1	0.135
Ca:P	2.85±0.05	2,66±0.03	0.537
Femur bone			
Bone ash (%)	39.8 a±1.93	34.9 b±1.58	<0.001
Ca (g·kg ⁻¹)	552.5 a±61.3	519.5 b±32.1	0.014
P (g·kg ⁻¹)	189.5±0.92	192.2±2.04	0.052
Ca:P	2.91±0.06	2.70±0.04	0.421

¹ – Means of 10 replicates (samples); SE – standard error.

a, b – mean values in rows with different letters differ significantly at $P \leq 0.05$.

Table 6. The effects of caponisation on blood characteristics at 24 weeks (mean \pm SE)¹

Item	Greenleg Partridge		P-value
	Non-caponised cocks	Capons	
Testosterone (ng/ml)	1.87 a \pm 0.042	0.17 b \pm 0.012	0.004
Ca (mmol \cdot l ⁻¹)	2.76 \pm 0.22	2.85 \pm 0.46	0.327
P (mmol \cdot l ⁻¹)	1.59 \pm 0.14	1.61 \pm 0.25	0.135
Alkaline phosphatase (U \cdot l ⁻¹)	681.5 \pm 221.3	720.9 \pm 178.8	0.781

¹ – Means of 10 replicates (samples); SE – standard error.

a, b – mean values in rows with different letters differ significantly at $P \leq 0.05$.

Level of testosterone, Ca, P, and activity alkaline phosphatase in the blood

At 24 weeks of rearing, there was a 11-fold decrease in testosterone level in capon birds. There was a numerical tendency towards an increase in the level of Ca and P as well as alkaline phosphatase activity in the capons (Table 6).

Discussion

Caponisation of cocks is aimed at achievement of better weight gains and delicate-flavoured meat, which is a result of the increasing content of intramuscular

fat (Hsieh *et al.* 2001). The results of our previous investigations of the Greenleg Partridge and Polbar conservative breeds demonstrated that surgical removal of testicles contributed to an increase in final body weight (Kwiecień *et al.*, 2015; 2018). Similarly, higher capon body weight was reported in other studies (Chen *et al.*, 2006 c; Chen *et al.*, 2007; Calik *et al.*, 2015; Adamski *et al.*, 2016; Zawacka *et al.*, 2017). Nevertheless, the results of the procedure and its effect on the body weight are contradictory. Investigations conducted by Shao *et al.* (2009) indicate a decrease in the weight of capons, while other studies show no significant effect of caponisation on the final weight (Symeon *et al.*, 2010, 2012; Franco *et al.*, 2016). This may be associated with the use of different bird breeds, changes in the rearing length, nutrition, or age at caponisation (Shao *et al.*, 2009). The intense weight gain in capons, mainly the weight of muscles, is associated with heavy loads carried by birds' leg bones. Hence, the bones are more susceptible to developmental and mineralisation disorders, which in turn may lead to deformities and even fractures (Leterrier and Nys, 1992). The risk of bone fracture may be associated with such factors as age and sex (Charuta *et al.*, 2012; Shim *et al.*, 2012) as well as hormonal disorders induced by testectomy. Steroids are indispensable for the development, growth, and maintenance of the skeleton. It has been shown that the increasing testosterone concentration stimulates the activity of osteoclasts in bone formation, hence their key role in the development of the skeleton in growing cocks (Gryzińska *et al.*, 2012). Additionally, androgens can promote proliferation and differentiation of osteoblasts, inhibit the recruitment of osteoclasts to osteoblasts, or influence calcium retention in bones, thereby accelerating their ossification (Falahati-Nini *et al.*, 2000; Gryzińska *et al.*, 2012). Investigations conducted by Lee *et al.* (2006) demonstrated increased bone resorption and reduced bone formation resulting from hypogonadal osteoporosis.

As expected, the testosterone concentrations in the plasma of intact male chickens increased significantly in the present study, which was confirmed in other studies (Chen *et al.*, 2005; Zawacka *et al.*, 2017). Our investigations demonstrated that the androgen deficiency in the capons (testosterone concentration of 0.17 ng/ml in the capons vs. 1.87 ng/ml in the non-caponised cocks) resulted in an increase in the abdominal fat (Kwiecień *et al.*, 2015) and body weight by 10%, compared with the non-caponised cocks. In contrast, it did not affect the bone weight and length, which may indicate that the skeleton growth in both groups was similar. However, in terms of the bone weight in relation to body weight, the relative bone weight in the capons was shown to be lower ($P \leq 0.05$). Similarly, the castration procedure described in other studies (Chen *et al.*, 2006 b; Lin *et al.*, 2012) did not influence the tibia weight but caused a reduction in the relative tibia weight. This indicates that tibial bones in caponised cocks must carry a greater weight load, which increases the risk of bone fracture. The caponisation-induced androgen deficiency changed the carcass tissue composition via intensification of anabolic processes and acceleration of the metabolic rate (Wink and Felts, 1980). However, it did not change the bone weight but reduced the bone proportions in the body weight. Therefore, the sex steroid deficiency during intensive skeletal development in young cocks may affect bone metabolism, resulting in bone loss and, consequently, osteoporosis.

Alkaline phosphatase and P content in the blood can be indicators of bone traits. The bone cell contains large amounts of alkaline phosphatase released into the blood during bone growth or degeneration. It is also associated with osteogenic activity and the P concentration (Galvanovskii et al., 1985). However, in the present study, the alkaline phosphatase activity and the P content did not change ($P>0.05$). As demonstrated by Lin and Hsu (2003), phosphorus can be released from the bone and enter the blood after caponisation, resulting in an increase in phosphorus ($P<0.05$) in the blood of 26-week-old capons. In turn, Ca retention should increase with its increasing amount in the blood (Shafty et al., 1990). In the present study, there was no effect of castration at 8 weeks of age on the plasma Ca content in the cocks at rearing week 24.

Androgens can contribute to Ca retention in bones, thus accelerating bone ossification and increasing their weight and ash content. With age, the testosterone levels increase, and this mainly stimulates the activity of osteoclasts in bone formation (Anderson et al., 1998; Gryzińska et al., 2012). In our study, the caponisation procedure significantly reduced the ash content in both bones and reduced the Ca content in the femoral bones. In turn, it exerted no effect on the P content and the Ca:P ratio. These results are in part consistent with the results reported by Chen et al. (2006 c) and Lin et al. (2012), which showed that caponisation reduced the ash content but did not change the Ca content. Another study (Lin and Hsu, 2003) demonstrated reduced Ca content in capons' bones, which did not influence the bone ash content. It can be assumed that the reduced content of bone ash and Ca in capons results from reduced deposition of Ca in bones caused by the deficiency of androgens (testosterone determining bone material maturity) induced by the caponization treatment (Mauras et al., 1999). Besides an inhibition or altered capacity of bone mineralization another possible interpretation is that bone turnover is increased (as suggested by increased alkaline phosphatase levels) and bone organic matrix does not have enough time to become fully mineralized (Rodriguez-Navarro et al., 2018).

Basically, the increased cross-sectional area of bone in capons is due to functional adaptation of bone to the increased weight of birds (Charuta et al., 2013). Although there were no statistical differences in our research, there was a tendency to increase the diameters (horizontal and vertical) of the cross-section at the bone fracture site in the capons, which was yet reflected in the geometric structure of the bones. The secondary moment of inertia and cross-sectional area of the femur increased, whereas only an increase in the secondary moment of inertia was noted in the tibial bone. The castration treatment induced a significant increase in the cortical layer thickness (mean relative wall thickness) in the femur, which determines the higher mechanical strength against applied forces. However, it did not affect the value of this trait in the tibial bone. Additionally, there was no impact of caponisation on the cortical index value. The castration procedure caused an increase in the medullary cavity, as confirmed by the increased diameters. The increase in the body weight of the capons increased the load and the mechanical wear of the bones.

Changes in mechanical parameters reflect alterations in bones occurring throughout life. Despite bone stiffness, bones exhibit some plasticity and flexibility and respond to the continuous or repetitive action of loading- and unloading-related

deformation forces by changes in their structure (Malcolm, 2002). In the present investigations, the bones of the non-caponised cocks were less mature and had smaller diameters. In contrast, they were better mineralised, which probably contributed to the increase in the mechanical parameters of the femoral and tibial bones. The mechanical tests carried out in the present study revealed that the bones of the capons were more susceptible to the risk of deformation; moreover, there was a greater risk of permanent bone deformation in the capons, as the threshold value of the load resulting in plastic bone deformation decreased, which may lead to bone damage. The decline in the bone mechanical values after the caponisation surgery is consistent with the results reported by Chen *et al.* (2006 b). Furthermore, the caponisation-induced reduction of the testosterone level probably contributes to a decline in the weight of capons' leg muscles (Symeon *et al.*, 2012; Kwiecień *et al.*, 2015). Additionally, the present study showed that the values of most of the measured strength parameters of tibial bone and physical characteristics of the bone material determining bone strength, i.e. yielding load towards bone weight, yielding load towards body weight, maximum force moment towards bone weight, and maximum force moment towards body weight, were lower in the capons.

The discrepancies between the results of various studies may be associated with the differences in the bird material analysed. An important effect is also exerted by the concentration of hormones, e.g. testosterone. At post-caponisation week 4, increased bone loss occurs, which is manifested by damage to bone cells, indicating a reduction of relative bone weight, breaking strength, cortical layer thickness, and ash, Ca, and P contents (Lin *et al.*, 2012).

Conclusions

In conclusion, our study demonstrates that castration has an adverse effect on the structural integrity of tibial and femoral bones and inhibits their development. Based on the analyses, it can be concluded that the bones of capons are more susceptible to deformations or fractures, which is reflected in the lower values of their geometrical and mechanical parameters. It seems that the tibia, which is burdened by a higher body weight load, is more susceptible to the effects of caponisation than the femoral bone, which increases the risk of tibia fracture.

Disclosure statement

No potential conflict of interest was reported by the authors.

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Received: 2 IV 2019

Accepted: 22 VIII 2019