Sildenafil alters biogenic amines and increases oxidative damage in brain regions of insulin-hypoglycemic rats

DAVID CALDERÓN GUZMÁN¹
NORMA OSNAYA BRIZUELA¹
MARIBEL ORTÍZ HERRERA²
ARMANDO VALENZUELA PERAZA¹
GERARDO BARRAGÁN MEJÍA¹
HUGO JUÁREZ OLGUÍN^{3,4}
FRANCISCA TRUJILLO JIMÉNEZ³

¹ *Laboratory of Neurosciences
Instituto Nacional de Pediatría (INP)
Mexico*

² *Laboratory of Experimental Bacteriology
INP, Mexico*

³ *Laboratory of Pharmacology, INP
Mexico*

⁴ *Department of Pharmacology
Faculty of Medicine, Universidad
Nacional Autónoma de México, Mexico*

The aim of the present study was to determine the effect of sildenafil on dopamine, 5-hydroxyindol acetic acid (5-HIAA) and selected biomarkers of oxidative stress in the brain of hypoglycemic rats. The animals were treated intraperitoneally as follows: group 1 (control), saline solution; group 2, insulin (10 U per rat or 50 U kg⁻¹); group 3, insulin + single dose of sildenafil (50 U kg⁻¹ + 50 mg kg⁻¹); group 4, insulin + three doses of sildenafil every 24 hours (50 U kg⁻¹ + 50 mg kg⁻¹). In groups 2, 3 and 4, insulin was administered every 24 hours for 10 days. Blood glucose was measured after the last treatment. On the last day of the treatment, the animals' brains were extracted to measure the levels of oxidative stress markers [H₂O₂, Ca²⁺, Mg²⁺-ATPase, glutathione and lipid peroxidation (TBARS)], dopamine and 5-HIAA in the cortex, striatum and cerebellum/medulla oblongata by validated methods. The results suggest that administration of insulin in combination with sildenafil induces hypoglycemia and hypotension, enhances oxidative damage and provokes changes in the brain metabolism of biogenic amines. Administration of insulin and sildenafil promotes biometabolic responses in glucose control, namely, it induces hypoglycemia and hypotension. It also enhances oxidative damage and provokes changes in the brain metabolism of biogenic amines.

Keywords: sildenafil, rat, hypoglycemia, biogenic amines, glutathione, H₂O₂, lipid peroxidation

Accepted March 30, 2019
Published May 20, 2019

Obesity increases the risk of atherosclerosis, renal diseases, vesicular lithiasis and diabetes. In the last few decades, type 2 diabetes mellitus has reached epidemic proportions due to bad lifestyles (1). In recent times, multiple drugs for effective management of obesity have emerged but most of them confer only modest benefits for patients. The majority of patients require insulin to control diabetes; however, insulin may provoke hypoglycemic conditions in patients with this pathology (2). Erectile dysfunction (ED) associated with type 2 diabetes is a severe problem, which requires effective treatment (3). Most adult patients, irrespective of age, will require sildenafil as an option for this dysfunction and it

* Corresponding author; e-mail: juarezol@yahoo.com

has recently become the preferred treatment of choice (4). Effects of sildenafil on the pulmonary vasculature and on erectile dysfunction are increasingly well understood (5); nevertheless, its mechanism in hypoglycemic patients is still unknown.

On the other hand, obesity and overweight have been associated with an increase in free radicals (FR) (6), which can put the cells of the central nervous system (CNS) in danger of oxidative damage caused by excessive production of oxygen reactive species involved in many pathological processes (7). Oxidative stress dysregulates energy and glucose homeostasis in different brain regions and disrupts brain circuits such as the dopaminergic system (8).

Plasma membrane phospholipids in neurons are in close contact with structural proteins embedded in the lipid bilayer, from which Na^+, K^+ -ATPase is responsible for keeping the ionic interchange through this bilayer by stimulating Na^+ and K^+ flows. Inhibition of Na^+, K^+ -ATPase activity alters the ionic interchange within the CNS (9).

In view of the above, the purpose of the present study was to evaluate the effect of sildenafil on the levels of dopamine and 5-HIAA monoamines, and selected oxidative stress markers in brain regions of young rats with insulin induced hypoglycemia.

EXPERIMENTAL

The study included twenty-eight male Wistar rats of 7 weeks of age and 200 g mean body mass. During the study, all animals were kept in closed boxes under a light:dark period of 12:12 h. Animals were divided into four groups of 7 rats each and treated intraperitoneally as follows: group 1 (control), 0.9 % NaCl; group 2, insulin (10 U per rat or 50 U kg^{-1}); group 3, insulin (50 U kg^{-1}) + single dose of sildenafil (50 mg kg^{-1}); group 4, insulin (50 U kg^{-1}) + three doses of sildenafil (50 mg kg^{-1}) every 24 hours. Treatment groups received insulin every 24 hours for 10 days. Groups 2–4 were treated every day with insulin for 10 days. Group 3 received sildenafil once, on day 10. Group 4 received sildenafil three times, namely, on days 8, 9 and 10, each 24 hours. Sixty minutes after receiving the last treatment, animals' blood was taken from the tail and used to measure their glucose levels. Animals were sacrificed by decapitation without anesthesia. Brains were extracted and immediately placed in a 0.9 % cold NaCl solution. They were later divided into the cortex, cerebellum/medulla oblongata and striatum. Tissues from these brain regions were then homogenized in a 0.05 mol L^{-1} phosphate buffer solution (PBS) and used to quantify 5-hydroxyindol acetic acid (5-HIAA), dopamine, $\text{Ca}^{2+}, \text{Mg}^{2+}$ -ATPase, H_2O_2 , lipid peroxidation and glutathione (GSH) using previously validated methods (10). All samples were frozen at -20°C until analysis.

Animal experiments were carried out in strict compliance with the Guidelines for Ethical Control and Supervision in the Care and Use of Animals and all experimental procedures were conducted following national and international rules. Also, the study was approved by the Laboratory Animals Use and Care Committee of the National Institute of Pediatrics, Ciudad de México, México.

Statistical analysis

Two-way ANOVA and Tukey's test were used in the analysis of the results while the statistical Dunnett's test was employed for Kramer HSD (honestly significant difference)

for comparison of all pairs. For all tests, values of $p < 0.05$ were considered statistically significant. Statistical Software JMP version 8.0.0 for academic purposes was used.

RESULTS AND DISCUSSION

Administration of insulin alone or in combination with sildenafil in single or repeated doses decreased the levels of glucose in a significant way ($p < 0.0001$) compared to the control group. Comparison among the experimental groups showed that the group treated with insulin + sildenafil in repeated doses had the biggest decrease in glucose levels, which in the group treated with insulin alone showed to be significantly different (Table I).

Table I. Levels of glucose in the blood of animals treated intraperitoneally either with insulin, sildenafil or both

Group/treatment	Glucose (mg per 100 mL)
Control (saline)	110.50 ± 7.56
Insulin (50 U kg ⁻¹)	57.83 ± 4.96*
Insulin (50 U kg ⁻¹) + sildenafil (50 mg kg ⁻¹) – single dose	48.57 ± 7.50*
Insulin (50 U kg ⁻¹) + sildenafil (50 mg kg ⁻¹) – 3 doses	46.57 ± 4.04**

Average ± SD, $n = 7$.
Statistically significant difference: * $p < 0.0001$ vs. control, ** $p = 0.017$ vs. insulin.

In the cortex, administration of insulin alone or combined with sildenafil produced a significant decrease in the levels of lipid peroxidation (TBARS – thiobarbituric acid reactive substances) compared to the control group (Table II).

A similar pattern was observed in the striatum; however, only in group 4 treated with repeated doses of sildenafil, TBARS level was significantly different from the control group although it was not different at all from the groups treated with insulin or a single dose of insulin + sildenafil (Table III).

In the cerebellum/medulla oblongata, a significant decrease was observed in the groups treated with a combination of insulin and sildenafil, both in single and repeated doses, compared to the control (Table IV). Similarly to what was observed in the cortex, the group that received repeated doses of insulin + sildenafil showed a larger decrease in the level of TBARS, which was significantly different from the group treated with a single dose of insulin + sildenafil (Table II).

In the cortex, groups 2 and 3 showed a significant decrease in the levels of H_2O_2 ; in the striatum, however, a discrete increase which was not significant was observed in the experimental groups compared to the control group. With regard to cerebellum/medulla oblongata, all groups showed a significant decrease in the levels of H_2O_2 compared to the control. Again, group 4 treated with repeated doses of sildenafil manifested a greater decrease in H_2O_2 levels, which was significant compared to the group treated with insulin alone.

Table II. Levels of biomarkers of oxidative stress and biogenic amines in the cortex of animals treated intraperitoneally either with insulin, sildenafil or both

Group/treatment	TBARS ($\mu\text{mol malondi-}$ aldehyde g^{-1}) ^a	H ₂ O ₂ ($\mu\text{mol g}^{-1}$) ^a	GSH (nmol g^{-1}) ^a	5-HIAA (nmol g^{-1}) ^a	Dopamine (nmol g^{-1}) ^a	ATPase ($\mu\text{mol Pi g}^{-1} \text{ min}^{-1}$) ^a
Control (saline)	2.47 ± 0.33	0.017 ± 0.005	603.03 ± 112.83	2.25 ± 0.41	22.08 ± 2.53	32.29 ± 3.85
Insulin (50 U kg ⁻¹)	2.05 ± 0.21*	0.009 ± 0.006*	704.93 ± 234.70	2.57 ± 0.48	23.11 ± 2.48	40.67 ± 7.59*
Insulin (50 U kg ⁻¹)+sildenafil (50 mg kg ⁻¹) – single dose	2.07 ± 0.30*	0.015 ± 0.014	581.651 ± 72.66	2.19 ± 0.39**	21.40 ± 2.73	43.39 ± 7.16*
Insulin (50 U kg ⁻¹)+sildenafil (50 mg kg ⁻¹) – 3 doses	1.78 ± 0.34***	0.010 ± 0.005*	571.13 ± 128.89	1.94 ± 0.32*	20.41 ± 2.91*	45.55 ± 7.18*

Average ± SD, $n = 7$; ^aWet mass (tissue) basis; Statistically significant differences: TBARS: * $p < 0.009$ vs. control, ** $p = 0.04$ vs. insulin, *** $p = 0.015$ vs. insulin + sildenafil single dose; H₂O₂: * $p < 0.005$ vs. control, ** $p = 0.04$ vs. insulin; Dopamine: * $p < 0.03$ vs. insulin; ATPase: * $p < 0.0009$ vs. control.

Table III. Levels of biomarkers of oxidative stress and biogenic amines in the striatum of animals treated intraperitoneally either with insulin, sildenafil or both

Group/treatment	TBARS ($\mu\text{mol malondi-}$ aldehyde g^{-1}) ^a	H ₂ O ₂ ($\mu\text{mol g}^{-1}$) ^a	GSH (nmol g^{-1}) ^a	5-HIAA (nmol g^{-1}) ^a	Dopamine (nmol g^{-1}) ^a	ATPase ($\mu\text{mol Pi g}^{-1} \text{ min}^{-1}$) ^a
Control (saline)	3.00 ± 0.70	0.023 ± 0.016	755.31 ± 288.32	2.47 ± 0.42	21.66 ± 2.80	80.62 ± 8.94
Insulin (50 U kg ⁻¹)	2.60 ± 0.51	0.042 ± 0.027	832.19 ± 239.21	2.64 ± 0.40	20.32 ± 1.69	86.98 ± 10.15
Insulin (50 U kg ⁻¹)+sildenafil (50 mg kg ⁻¹) – single dose	2.56 ± 0.42	0.035 ± 0.028	888.39 ± 187.00	2.63 ± 0.77	20.45 ± 2.49	93.03 ± 9.19*
Insulin (50 U kg ⁻¹)+sildenafil (50 mg kg ⁻¹) – 3 doses	2.50 ± 0.25*	0.025 ± 0.017	861.97 ± 172.75	2.73 ± 0.36	20.95 ± 2.97	89.89 ± 11.41

Average ± SD, $n = 7$; ^aWet mass (tissue) basis; Statistically significant differences: TBARS: * $p < 0.03$ vs. control, ATPase: * $p < 0.01$ vs. control.

Table IV. Levels of biomarkers of oxidative stress and biogenic amines in the cerebellum/medulla oblongata of animals treated intraperitoneally either with insulin, sildenafil or both

Group/treatment	TBARS ($\mu\text{mol malondi-}$ aldehyde g^{-1}) ^a	H ₂ O ₂ ($\mu\text{mol g}^{-1}$) ^a	GSH (nmol g^{-1}) ^a	5-HIAA (mmol/g) ^a	Dopamine (nmol g^{-1}) ^a	ATPase ($\mu\text{mol Pi g}^{-1} \text{ min}^{-1}$) ^a
Control (saline)	2.97 ± 0.64	0.050 ± 0.015	534.39 ± 93.51	2.24 ± 0.33	20.96 ± 1.48	67.71 ± 6.07
Insulin (50 U kg ⁻¹)	2.68 ± 0.30	0.035 ± 0.017*	571.34 ± 104.46	2.36 ± 0.21	21.96 ± 2.19	71.04 ± 9.36
Insulin (50 U kg ⁻¹)+sildenafil (50 mg kg ⁻¹) – single dose	2.49 ± 0.37*	0.023 ± 0.014*	476.96 ± 162.73	2.10 ± 0.56	20.22 ± 2.98	75.34 ± 10.48
Insulin (50 U kg ⁻¹) + sildenafil (50 mg kg ⁻¹) – 3 doses	2.06 ± 0.37****	0.021 ± 0.012***	428.21 ± 111.89***	2.16 ± 0.37	20.34 ± 1.86	84.73 ± 11.60*****

Average ± SD $n = 7$; ^a Wet mass (tissue) basis; Statistically significant differences: TBARS: * $p < 0.03$ vs. control, ** $p < 0.0001$ vs. insulin, *** $p = 0.01$ vs. insulin + sildenafil single dose; H₂O₂: * $p < 0.02$ vs. control, ** $p < 0.03$ vs. control, *** $p < 0.0001$ vs. insulin; GSH: * $p < 0.02$ vs. control, ** $p < 0.0001$ vs. control, *** $p = 0.0002$ vs. insulin, **** $p = 0.01$ vs. insulin + sildenafil single dose.

In the cortex and striatum, statistical analysis of the effects of insulin alone or combined with single or repeated doses of sildenafil on the levels of GSH did not reveal significant changes compared to the control group. In the cerebellum/medulla oblongata, a significant increase was observed in the levels of this biogenic amine in the groups treated with insulin alone and group 4 with repeated doses of sildenafil, compared to the control group.

Statistical analysis of the data revealed a significant increase in the levels of 5-HIAA in the cortex of group 4 treated with insulin + repeated doses of sildenafil compared to the control. Groups treated with the combination of insulin + sildenafil in both treatment schemes showed a decrease in the levels of 5-HIAA compared to the group that received insulin alone. In the striatum and cerebellum/medulla oblongata, the analysis did not reveal any significant changes in the concentration of 5-HIAA. Administration of insulin + sildenafil in repeated doses produced a decrease in the concentration of dopamine (DA) in the cortex, which was statistically significant only compared to the group that received insulin alone. In other regions of the brain, no changes in the concentration of DA were observed.

In the case of ATPase activity, the administration of insulin alone or combined with sildenafil increased significantly the activity of amine in the cortex and striatum. However, in the cerebellum/medulla oblongata, this increase was statistically significant only for group 4 treated with insulin + repeated doses of sildenafil compared to the control and the group that received a single dose of insulin + sildenafil.

A major issue in glycemic control in neurocritical care is the lack of continuous glucose monitoring in patients under insulin therapy. Thus, these patients are

associated with higher rates of hypoglycemia without a chance to improve their survival rate. This may probably be the reason why Bilotta and Rossa (11) recommended adequate nutrition before and during insulin infusion.

In this work, dopamine and 5-HIAA led to a significant decrease in the cortex regions. These findings coincide with that of Ammon-Treiber *et al.* (12) who suggest that decreased levels of DA and 5-HIAA can aggravate neurotoxic effects on the hypoxia/hypoglycemia model, and that their metabolites may increase oxidative damage to the brain.

Hypoglycemic animals exerted a decrease in the levels of glutathione, suggesting that insulin and sildenafil provide no protection from oxidative stress to the brain. Hence, such doses of insulin and sildenafil may make the CNS vulnerable to damage by oxidative stress caused by depletion of the glutathione antioxidant. NO production in selected neuron populations may have induced oxidative stress (13), which altered GSH metabolism in the brain.

Suppression of reactive oxygen species-dependent pathways in the endothelium restores insulin delivery to peripheral organs by preserving nitric oxide (NO) availability. The lipoperoxidation and H₂O₂ levels decreased as a consequence of sildenafil and insulin treatment. This result is in accord with what was previously reported in our laboratories where hypoglycemia was associated with increased oxidative damage (14).

Animals treated with insulin and sildenafil experienced an increase in total ATPase in the cortex and striatum. This finding agrees with the report of Filippov (15), who suggested an increase in the number of active molecules of ATPase under the influence of membrane structural changes in the hypoglycemic state. There was a decrease in blood glucose in the animals that received sildenafil, thus confirming the hypoglycemia model and a potential increase of oxidative damage with the use of this drug. This concurs with Mandosi *et al.* (16) who found out that sildenafil exerted a beneficial effect on glycometabolic control in type 2 diabetes by increasing insulin sensitivity (17). Moreover, the combination of sildenafil and insulin may offer an improved cerebrovascular response (18), due to its relaxation action on corpora cavernosa.

CONCLUSIONS

Administration of insulin and sildenafil promoted biometabolic responses in glucose control. Hypoglycemic animals showed a decrease in the levels of glutathione suggesting that insulin and sildenafil provided no protection from oxidative stress to the brain. Hence, such doses of insulin and sildenafil may make the CNS vulnerable to damage caused by oxidative stress due to depletion of the glutathione antioxidant.

REFERENCES

1. A. V. Raveendran, E. C. Chacko and J. M. Pappachan, Non-pharmacological treatment options in the management of diabetes mellitus, *Eur. Endocrinol.* **14** (2018) 31–39; <https://doi.org/10.17925/EE.2018.14.2.31>
2. A. Hussain, O. B. Latiwesh, F. Ali, M. Y. G. Younis and J. A. Alammari, Effects of body mass index, glycemic control, and hypoglycemic drugs on serum uric acid levels in type 2 diabetic patients, *Cureus* **10** (2018) e3158; <https://doi.org/10.7759/cureus.3158>

3. G. T. Chen, B. B. Yang, J. H. Chen, Z. Zhang, L. L. Zhu, H. S. Jiang, W. Yu, Y. Chen and Y. T. Dai, Pancreatic kininogenase improves erectile function in streptozotocin-induced type 2 diabetic rats with erectile dysfunction, *Asian J. Androl.* **20** (2018) 448–453; https://doi.org/10.4103/aja.aja_23_18
4. L. Chen, S. E. Staubli, M. P. Schneider, A. G. Kessels, S. Ivic, L. M. Bachmann and T. M. Kessler, Phosphodiesterase 5 inhibitors for the treatment of erectile dysfunction: a trade-off network meta-analysis, *Eur. Urol.* **68** (2015) 674–680; <https://doi.org/10.1016/j.eururo.2015.03.031>
5. R. S. Calabrò, G. Polimeni and P. Bramanti, Current and future therapies of erectile dysfunction in neurological disorders, *Rec. Pat. CNS Drug Discov.* **6** (2011) 48–64.
6. S. J. Flora, Role of free radicals and antioxidants in health and disease, *Cell. Mol. Biol.* **53** (2007) 1–2.
7. J. T. Coyle and P. Puttfarcken, Oxidative stress, glutamate, and neurodegenerative disorders, *Science* **262** (1993) 689–695.
8. M. C. Vogt and J. C. Brüning, CNS insulin signaling in the control of energy homeostasis and glucose metabolism – from embryo to old age, *Trends Endocrinol. Metab.* **24** (2013) 76–84; <https://doi.org/10.1016/j.tem.2012.11.004>
9. G. D. Calderon, N. Osnaya Brizuela, M. Ortiz Herrera, O. H. Juárez, A. Valenzuela Peraza and G. Barragan Mejía, Effect of an antiviral and vitamins A, C, D on dopamine and some oxidative stress markers in rat brain exposed to ozone, *Arch. Biol. Sci. Belgrade* **65** (2013) 1371–1379; <https://doi.org/10.1186/s12868-018-0474-4>
10. D. Calderón Guzmán, N. Osnaya Brizuela, M. Ortiz Herrera, E. Hernandez Garcia, G. Barragan Mejía, H. Juárez Olguín, A. Valenzuela Peraza, J. Attilus and N. Labra Ruiz, Effect of cerebrolysin on dopaminergic neurodegeneration of rat with oxidative stress induced by 3-nitropropionic acid, *Acta Pharm.* **66** (2016) 443–448; <https://doi.org/10.1515/acph-2016-0027>
11. F. Bilotta and G. Rosa, Optimal glycemic control in neurocritical care patients, *Crit. Care* **16** (2012) 163–165; <https://doi.org/10.1186/cc11521>
12. S. Ammon-Treiber, D. Stolze, H. Schroder, H. Loh and V. Holtt, Effects of opioid antagonists and morphine in a hippocampal hypoxia/hypoglycemia model, *Neuropharmacology* **49** (2005) 1160–1169; <https://doi.org/10.1016/j.neuropharm.2005.06.016>
13. J. H. Jhamandas, K. H. Harris, T. Petrov and K. H. Jhamandas, Activation of nitric oxide-synthesizing neurons during precipitated morphine withdrawal, *Neuroreport* **7** (1996) 2843–2846.
14. G. D. Calderón, G. E. Hernández, M. G. Barragán, O. H. Juárez, G. J. Saldivar and R. N. Labra, Effect of morphine and lacosamide on levels of dopamine and 5-HIAA in brain regions of rats with induced hypoglycemia, *Pak. J. Biol. Sci.* **17** (2014) 292–296.
15. S. P. Filippov, ATPase activity of rat brain microsomal and synaptosomal fractions in insulin hypoglycemia and its treatment with glucose, *Probl. Endokrinol. (Moscow)* **37** (1991) 52–54; <https://doi.org/10.1134/S0022093006020062>
16. E. Mandosi, E. Giannetta, T. Filardi, M. Lococo, C. Bertolini, M. Fallarino, D. Gianfrilli, M. A. Venneri, L. Lenti, S. Morano and A. Lenzi, Endothelial dysfunction markers as a therapeutic target for sildenafil treatment and effects on metabolic control in type 2 diabetes, *Expert Opin. Ther. Target.* **19** (2015) 1617–1622; <https://doi.org/10.1517/14728222.2015.1066337>
17. C. E. Ramirez, H. Nian, Y. Chang, L. J. Gamboa, J. M. Luther, N. J. Brown and C. A. Shibao, Treatment with sildenafil improves insulin sensitivity in prediabetes: a randomized, controlled trial, *JCEM* **100** (2015) 4533–4540; <https://doi.org/10.1210/jc.2015-3415>
18. F. G. Al-Amran, A. A. Zwain, N. R. Hadi and A. M. Al-Mudhaffer, Autonomic cerebral vascular response to sildenafil in diabetic patient, *Diabetol. Metab. Syndr.* **4** (2012) 2–8; <https://doi.org/10.1186/1758-5996-4-2>