Genetic Variation and Structure of native and introduced Casuarina equisetifolia (L. Johnson) Provenances

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Abstract

Casuarina equisetifolia is an important tree species in tropical/subtropical zones of Asia, the Pacific and Africa. In this study, 220 individuals from seven native provenances and eleven introduced provenances of $C.\ equisetifolia$ were analyzed to assess the genetic variation and structure using amplified fragment length polymorphism (AFLP) markers. A total of 465 bands were obtained by eight primer pairs, among which 153 were polymorphic. The mean Nei's gene diversity H=0.2113 calculated for 18 provenances and the total gene diversity $H_T=0.4065$ calculated for native provenances suggested abundant variation within provenances and species. High genetic divergence coefficient ($G_{ST}=0.4737$) and low gene flow ($N_m=0.5555$) detected among native provenances suggesting high differentiation of $C.\ equisetifolia$. An AMOVA analysis for native provenances revealed a high proportion (46.07%) of the total

genetic variation distributed among provenances. The UPGMA clustering (r = 0.8028) and the Mantel test (r = 0.0716) for native provenances showed there was no correlation among genetic relationships and geographical distribution. The genetic information provided important implications for the future conservation and breeding programs of C. equisetifolia.

Key words: Casuarina equisetifolia, AFLP markers, genetic variation, genetic structure, native range, introduced range.

Introduction

Casuarina equisetifolia (L. Johnson) has two subspecies: subsp. incana and subsp. equisetifolia (MIDGLEY et al., 1983; Wilson et al., 1989). The former occurs exclusively along the coast of Queensland and northern New South Wales of Australia and in Vanuatu. The latter has a wide native range, occurs on tropical and subtropical zones of Asia, the Pacific and Africa from throughout Malaysia to northern Australia, Melanesia, and Polynesia (Figure 1). As a fast-growing and nitrogen fixing tree of social, economic and environmental importance, it is commonly used in agroforestry systems, for soil stabilization, reclamation work, in coastal protection and rehabilitation. Though a littoral species in natural habitats it is well adapted to inland area, and has been introduced into southern China, India, East and West Africa, the United States of America, the

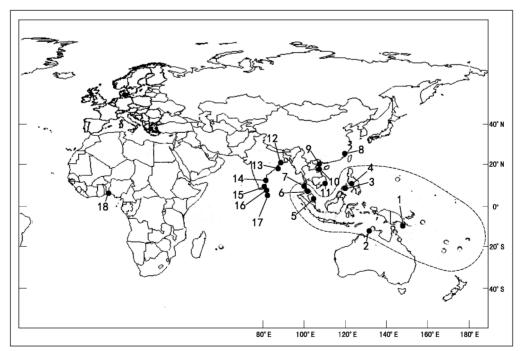


Figure 1. – The geographical distribution of 18 Casuarina equisetifolia provenances, within line is the native range of Casuarina equisetifolia ssp. equisetifolia.

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Caribbean, Vietnam, and some Middle East countries for more than a century. The provenances of *C. equisetifolia* exhibit significant of variation for growth characteristics, and grow on a variety of sites with very different annual precipitation and climatic conditions. Marked variation in growth and branching habit between different provenances, as well as individual trees within provenances, has been observed (PINYOPUSARERK and HOUSE, 1993; PINYOPUSARERK and WILLIAMS, 2000). However very little information is correctly available on the genetic variation of this species based on molecular technique despite its ecological and economic importance.

Information on the magnitude and structure of genetic variation within a species is an integral part of conservation and improvement of genetic resources. Amplified fragment length polymorphism (AFLP), which can be used to analyze large numbers of loci distributed throughout the genome, have proven a powerful tool in the assessment of genetic variation both within and among plant provenances without detailed prior knowledge of DNA sequences (Vos et al., 1995; Brown et al., 1990; Jones et al., 1998). AFLP markers have been successfully used for analyzing genetic variation and genetic structure (Barrett et al., 1998; Janssen et al., 1996; Xue et al., 2005; Stefenon et al., 2006).

The main goal in this study is to survey the levels and patterns of genetic variation of the provenances of *C. equisetifolia* using AFLP markers, and the genetic differentiation and genetic relationships among provenances were analyzed at the same time. Furthermore, we can compare the genetic diversity among native and introduced provenances. This work will provide valuable information for further genetic conservation and breeding program in *C. equisetifolia*.

Materials and Methods

Plant materials and DNA extraction

A total of 220 plants belong to 18 provenances were collected from an international provenance trial garden of *C. equisetifolia* subsp. *equisetifolia* established at

Zhangzhou in Fujian province of China. Some introduced provenances were included in the samples because we want to know the variation and the genetic information are needed for future management of these provenances since they have been identified as potential useful provenances in the south of China (ZHONG et al., 2001). The provenances in the introduced range are the result of provenance trail plantations. The geographic and ecological parameters of provenances are shown in Figure 1 and Table 1. Genomic DNA of each trees were extracted from fresh branchlets using a modified CTAB extraction (DOYLE, 1991). DNA concentrations were estimated and standardised against known concentrations of lambda DNA on 1% agarose gels.

AFLP analysis

AFLP analysis was carried out by the method of Vos et al. (1995) with some modifications. Approximately 300 ng of total genomic DNA per samples were digested with EcoRI and MseI and then ligated with EcoRI adaptor and MseI adaptor. Including 64 primer combinations with three selective bases were screened in a preliminary test, eight primer (Table 2) combinations found to give good amplifications were selected for further study. Pre-amplifications were performed in a total volume of 20 ul containing 2 ul of ligation mixture, running on GeneAmp PCR 9700 using the following temperature conditions: denaturation at 94°C for 3 min, 30 cycles of denaturation at 94°C for 30 s, followed by annealing at 56 °C for 30 s, ending with extension at 72 °C for 60 s, a final step at 72°C for 5 min. Selective amplifications were performed in a 20 ul volume containing 4 ul of preamplification products (diluted 50 times in TE). Amplifications were performed using the following temperature conditions: one cycle of denaturation at 94°C for 30 s, followed by annealing at 65°C for 30 s, ending with extension at 72°C for 60 s, followed by 12 cycles under the conditions listed above, except for a 0.7°C lower annealing temperature for each cycle, and finally 30 cycles of 94°C for 30 s, 56°C for 30 s, 72°C for 60 s. Following the selective amplification, the reaction products were mixed with an equal volume of loading buffer (98% formamide, 10 mM EDTA, 0.1% xylene cyanol, and 0.1%

Table 1. - The information of 18 provenances of Casuarina equisetifolia ssp. equisetifolia investigated for AFLP variation.

Pro. No.	CSIRO Seedlot No.	Provenance name Country Sample size		Latitude	Longitude	Altitude (m)		
Native	Native provenances							
1	18153	Ela Beach	Papua New Guinea	15	09° 05′ S	148° 17′ E	10	
2	18008	Darwin, Northern Territory	Australia	10	12° 25′ S	130° 50′ E	20	
3	18357	Narra, Palawan	Philippines	10	09° 19′ N	118° 29′ E	10	
4	18154	Aklan, Panay Island	Philippines	10	11° 31′ N	122° 30′ E	30	
5	18348	Kuantan, Pehang	Malaysia	15	06° 30′ S	99° 45′ E	30	
6	18298	Had Chao Mai, Trang	Thailand	10	07° 33′ N	100° 37′ E	2	
7	18297	Ban Kam Phuam,Ranong	Thailand	15	09° 21′ N	98° 27′ E	10	
Introduc	ed provenances							
8	Local	Dongshan, Fujian	China	12	23° 40′ N	117° 28′ E	4	
9	18128	Non Nuoc	Vietnam	10	16° 06′ N	106° 20′ E	2	
10	18086	Hai Thinh, Ha Nam Ninh	Vietnam	13	20° 22′ N	106° 21′ E	1	
11	18152	Ninh Chu, Ninh Thuan	Vietnam	10	11° 33′ N	108° 59′ E	2	
12	18015	Chandipur, Balasore, Orissa	India	15	21° 30′ N	86° 54′E	2	
13	18014	Balukhanda, Orissa	India	15	19° 50′ N	85° 53′E	10	
14	18120	Chengai Anna, Tamil Nadu	India	10	12° 36′ N	79° 48′ E	50	
15	18119	Rameswaram, Tamil Nadu	India	10	09° 15′ N	79° 20′ E	5	
16	18288	Madagama	Sri Lanka	15	08° 06′ S	80° 15′ E	80	
17	18287	Hambantota	Sri Lanka	10	06° 08′ N	81° 07′ E	16	
18	18355	Cotonou	Benin	15	06° 23′ S	02° 13′ E	8	

Table 2. – Number of fragments, number of polymorphic fragments, percentage of polymorphic fragments, and size of fragments for each primer pair used for AFLP analysis on Casuarina equisetifolia.

Primer pairs	Number of fragments	Number of polymorphic fragments	Percentage of polymorphic fragments	Size of fragments (bp)
M-CAC/E-ACT	45	19	42,22%	75-600
M-CTA/E-AAG	80	24	30%	50-620
M-CTA/E-ACA	65	23	35.38%	115-820
M-CTC/E-AAC	60	25	41.67%	40-530
M-CTC/E-ACT	50	20	40%	40-375
M-CTG/E-AAG	48	22	45.83%	100-400
M-CTT/E-ACC	64	9	14.06%	40-680
M-CTT/E-AGC	53	11	20.75%	102-410
Total	465	153		
Mean	58.13	19.13	32.90%	

bromophenol blue). The mixtures were denatured for 5 min at 94°C then directly placed on ice. About 6 µl of the mixtures were run in 6% (v/v) polyacrylamide gels with a DNA sequencer. The gels were silver stained using the protocol published by STREIFF et al. (1998).

Genetic analysis

Each AFLP band of unambiguous pattern was given a score of 1 for presence or 0 for absence across all the polymorphic loci to create a binary matrix. POPGENE, version 1.31 (Yeh et al., 1999) was used to calculate genetic parameters within the 18 sampled provenances, including the number of polymorphic loci (NP), percentage of polymorphic loci (P), observed number of alleles per locus (N_a) , effective number of alleles per locus (N_a) , gene diversity (H) (NEI, 1973), Shannon's information index (I) (Lewontin, 1972). The total gene diversity (H_T) within species and within provenances (H_S) , differentiation coefficient (G_{ST}) and gene flow (N_m) were calculated only for native provenances assuming Hardy-Weinberg equilibrium. The unbiased measure of genetic identity (GID) and genetic distance (GD) among native provenances was calculated using Nei's (1978), and the genetic distance matrix was used to construct dendrogram using unweighted pair group method (UPGMA) of SHAN in NTSYS software (ROHLF, 1994). In addition, a

Mantel test among genetic distances and geographic distances among native provenances was carried out by TFPGA 1.3 (Mantel, 1967). The binary matrix of native provenances was used to generated a distance matrix with the AMOVA-PREP version 1.01 (Miller, 1998) and the resulting distance matrix was subjected to an analysis of molecular variance (AMOVA) using WINAMOVA version 1.55 (Stewart and Excoffier, 1996). The hierarchical analysis of the genetic variation partitioned among regions, as well as among provenances within regions was not carried out for some regions only have one or two provenances.

Results

Genetic variation within native and introduced provenances

The amplified results by each primer pair were shown in *Table 2*. Eight primer pairs revealed a total of 465 DNA fragments ranging from 40 bp to 820 bp, among which 153 (32.90%) of these bands were polymorphic loci either among or within provenances. Primer pairs were significantly different in their capacity to detect polymorphisms.

The genetic parameters for each of 18 *C. equisetifolia* provenances summarized in *Table 3* indicated a tenden-

 $Table\ 3$. – Genetic variation parameters of Casuarina equisetifolia provenances detected by AFLP markers. The mean values were calculated across provenances. NP, number of polymorphic loci; P, percentage of polymorphic loci; Na, observed number of alleles per locus; Na, effective number of alleles per locus; Na, Na gene diversity; Na, Na shannon's information index.

Pro. No.	NP	P	Na	Ne	Н	I
Native provenances						
1	117	76.47	1.7647	1.4569	0.2669	0.3990
2	62	40.52	1.4052	1.2651	0.1533	0.2263
3	87	56.86	1.5686	1.3701	0.2125	0.3140
4	91	59.48	1.5948	1.4481	0.2466	0.3568
5	87	56.86	1.5686	1.4328	0.2369	0.3411
6	72	47.06	1.4706	1.3066	0.1737	0.2561
7	104	67.97	1.6797	1.3515	0.2077	0.3175
Introduced provenances						
(plantations)						
8	20	13.07	1.1307	1.0817	0.0472	0.0701
9	85	55.56	1.5556	1.4030	0.2248	0.3266
10	92	60.13	1.6013	1.3307	0.1972	0.2986
11	91	59.48	1.5948	1.4258	0.2359	0.3430
12	99	64.71	1.6471	1.4026	0.2328	0.3458
13	105	68.63	1.6863	1.5186	0.2854	0.4121
14	81	52.94	1.5294	1.3909	0.2186	0.3173
15	80	52.29	1.5229	1.3261	0.1871	0.2774
16	93	60.78	1.6078	1.4113	0.2355	0.3460
17	97	63.40	1.6340	1.4316	0.2457	0.3609
18	78	50.89	1.5098	1.3475	0.1963	0.2878
Mean	85.61	55.95	1.5596	1.3723	0.2113	0.3109

Table 4. – Analysis of molecular variance (AMOVA) for seven natural provenances of Casuarina equisetifolia. d.f., Degrees of freedom; SSD, Sum of squares; MSD, Mean squared deviation; % Total variance, the percentage of the total variance.

Source of variation	d. f.	SSD	MSD	Variance component	% Total variance	P value
Among provenances	6	1175.8804	195.980	14.814	46.07%	< 0.0002
Within provenances	78	1352.5667	17.341	17.340	53.93%	< 0.0002

Table 5. – NEi's (1978) unbiase measures of genetic identity (above diagonal) and genetic distance (below diagonal) for seven natural provenances Casuarina equisetifolia.

Pro. No.	1	2	3	4	5	6	7
1	****	0.6606	0.7192	0.7355	0.7288	0.6991	0.7747
2	0.4146	****	0.7096	0.6848	0.7538	0.6948	0.6824
3	0.3296	0.3430	****	0.7077	0.8157	0.7949	0.7462
4	0.3072	0.3786	0.3457	****	0.7019	0.6753	0.7112
5	0.3164	0.2826	0.2038	0.3540	****	0.7472	0.6967
6	0.3580	0.3641	0.2295	0.3926	0.2914	****	0.7538
7	0.2553	0.3821	0.2927	0.3408	0.3614	0.2827	****

cy for genetic variation of the Ela Beach native provenance in Papua New Guinea and Balukhanda, Orissa introduced provenance in India were higher than other provenances. The former was highest in number of polymorphic loci (NP) 117, percentage of polymorphic loci (P) 76.47% and observed number of alleles per locus (N_a) 1.7647. The latter was highest in effective number of alleles per locus (N_e), gene diversity (H) and Shannon information index (I) 1.5186, 0.2854 and 0.4121, respectively. The Dongshan, Fujian provenance of China was lowest in all of the values, there were NP=20, P=13.07%, $N_a=1.1307$, $N_e=1.0817$, H=0.0472 and I=0.0701, respectively. The mean gene diversity was 0.2113 and Shannon information index was 0.3109.

Genetic structure among native provenances

The POPGENE analyses for native provenances showed an abundant diversity ($H_T = 0.4065 \pm 0.0128$) at the species level, partitioning which $H_S = 0.2139 \pm 0.0093$ distributed within provenances and the rest among provenances. A high coefficient of genetic differentiation $G_{ST} = 0.4737$ and low gene flow $N_m = 0.5555$ [$N_m = 0.5(1 - G_{ST})/G_{ST}$] indicated the genetic divergence among C. equisetifolia provenances was substantial. An analysis of molecular variation (AMOVA) with 5000 permutations showed variance component 46.07% among

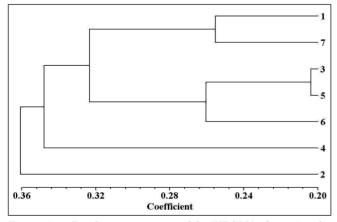


Figure 2. – Dendrogram generated by UPGMA clustering for seven native provenances of Casuarina equisetifolia, based on the Nei's (1978) genetic distances.

provenances and 53.93% within provenances (*Table 4*), both were highly significant (p < 0.0002).

Relationships among native provenances

Genetic identity and genetic distances for each pair of native provenances were calculated and shown in Table 5. The genetic distances ranged from 0.2038 (between provenance 3 and provenance 5) to 0.4146 (between provenance 1 and provenance 2). The UPGMA cluster analysis based on Nei's (1978) unbiased genetic distances was performed to further show the genetic relationships among provenances (Figure 2). The cophenetic correlation coefficient of this cluster analysis was r = 0.8028 (p = 0.9994). Four groupings could be recognized from the dendrogram: the first group consisted of provenances 1 and 7 while the second group comprised provenance 3, 5 and 6. The provenance 4 and provenance 2 were segregated in third group and fourth group, respectively. A Mantel test with 1000 random permutations revealed no correlation between pair-wise genetic distances and geographical distances among provenances (correlation coefficient of r = 0.0716, p = 0.05).

Discussion

This study revealed that there was a large amount of genetic variation at provenance level and at species level of *C. equisetifolia* except the local Dongshan, Fujian provenance had extraordinarily low level variation. The mean gene diversity (0.2113) was higher than HO (2002) result of 12 *C. equisetifolia* provenances (0.1473) assessed by RAPD, this may be account by the different provenances used in the two studies and the more gene loci can be detected by AFLP markers. The average Shannon's information index (0.3109) and the total gene diversity (0.4065) were also higher than many other plants such as *Picea asperata* (0.227 and 0.237, respectively, XUE et al., 2005) based on AFLP.

The present result showed genetic variation among provenances (46.07%) was a little higher than HO (39.28%, 2002) result of 12 *C. equisetifolia* provenances based on RAPD markers, however, the same tendency of high proportion of total variation among provenances

has been tested by the two different makers. Such results are in contrast with expectations for woody, perennial, predominantly outcrossed species, which maintain most variations within provenances (HAMRICK, 1990). However, this may be due to the low sample size in the study. High variation among provenances was also found in another tropical tree Tectona grandis (43%, Shrestha et al., 2005) by AFLP. Even more variations among provenances have been detected in Syringa oblate, (51.53%, MING and Gu, 2006), Moringa oleifera Lam. (59%, MULUVI et al., 1999) and Euterpe edulis Mart. (57%, CARDOSO et al., 2000) based on AFLP. The presence of such a high genetic variation among provenances could be attributed to the features of the wide distribution range of this species and the habitats where the sampled provenances occur. Its wide distribution is scattered through many islands, and therefore geographical isolation may occur. High degree of differentiation (0.4737) among provenances suggests that they have probably undergone random changes in genetic composition in response to prevalent environment conditions with limited gene flow (0.5555) between provenances. According to WRIGHT (1969), the critical value for N_m is 0.5. When N_m value is below 1, it means that provenances began to differentiate due to genetic drift. $N_{\rm m}$ value below 0.5 indicates that populations will diverge extensively as a result of drift (McDermott and McDonald, 1993). Overall, the calculated gene flow (N_m) indicates that the provenances were in the process of differentiation due to genetic drift.

It is interesting to find out that Balukhanda, Orissa introduced provenance was highest in genetic variation, reflecting the important implications on the introductions into this site. Multiple seed sources could have been collected to establish this plantation. Similarly, presumption that introduced provenances may have been derived from multiple seed sources can be seen from the higher variation (two Orissa provenance in India, two Sri Lanka provenances) than some native provenances. Although some of the introduced provenances existed abundant variation and could be used in the planting programs and breeding programs, the really problem is that there were no records of the origins of the seed which was first introduced to these places. Sampling of provenances in the native range must be much denser than presently undertaken with high resolution markers (such as AFLP markers) are likely to yield better results over the identities of the originating native provenances. In addition, chloroplast DNA pattern across the native range of the species might also enable increased resolution among the introduced provenances.

In general, if native provenances originated from the same hypothetical ancestors and underwent stable evolution, geographically close provenances should show the closest genetic relationships. However, the present result showed there was no distinct pattern of genetic distances correlated with geographic distances. This was consistent with Yang (1995) result that the variation of seed weight and seedling growth among provenances of *C. equisetifolia* was not continuous. However, the limited number of native provenances in this study may be

another factor on the discordance. Introduced provenances were not included in the analysis for these provenances were only one or two generations removed from their wild provenances and were not in Hardy-Weinberg equilibrium. In fact, many of the clusters in the dendrogram had very low relation values (XUE et al., 2005; EREN KANDEDMIR et al., 2004; LI and XIA, 2005; HONG et al., 2004; ZENG et al., 2003), suggesting that the clusters could be refined if more provenances and markers are included in future studies.

This work provides a deep insight into the genetic variation and genetic structure of C. equisetifolia, and supplies fundamental genetic information for conservation and breeding program of this species. Conservation programs dealing with native provenances of C. equisetifolia must take into account that most of the variation was distributed within provenance (53.93%). Conservations of more family within a provenance could get important variation. Low intensity phenotypic selections within provenances are appropriate strategy to exploit the intra-provenance variation, offering a potential for selection of genetic material for tree improvement programs. On the other hand, high proportion of the total variation (46.07%) and extensive genetic differentiation (0.4737) among provenances suggesting each provenance should be included in the conservation program because they possess partially unique genetic characteristics. In such a situation loss any provenance could result in an important loss of variation of the species. Preservation plots earmarked within natural forests can be considered as in situ conservation stands. They can be expected to retain much of the original variation as a result of better protection than other areas. Some introduced provenances (Balukhanda, Orissa in India and Hambantota in Sri Lanka) can also be considered in conservation and breeding program in the south of china for these provenances possesses substantial genetic variation. In this circumstance, the results of provenances trials are expected to be useful in the selection of a suitable provenance for a particular environment.

Superior trees assembled in germplasm banks is the only source of ex situ conservation for C. equisetifolia. Clonal seed orchards have a good collection for static conservation. However, they represent only a fraction of total genetic variation present within the species. Seedling seed orchards can serve as dynamic ex situ conservation stands with sufficiently broad genetic base for a long breeding and conservation program. An appropriate number of superior plants (It was collected 30-50 in China) of each provenances should be considered for effective conservation of the total genetic variation. As in China, more families should be included for those superior provenances and high variation provenances of C. equisetifolia when considering ex situ conservation, however the Dongshan, Fujian provenance could be much less for it may not be a good base to be conserved and used in improvement programs.

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Annual Fertility Variation in Clonal Seed Orchards of Teak (*Tectona grandis* L.f.) and its Impact on Seed Crop

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Abstract

Fertility variation was studied in two clonal seed orchards (CSO) of teak in four consecutive years (2003–2006). Both orchards were raised in 1976 with grafts of phenotypes selected for growth and form. The seed orchards of CSO I (Topslip, Tamil Nadu State) and CSO II (Walayar, Kerala State) have 15 and 20 clones, respectively, with 13 common clones. The proportion of flowering ramets was generally low ranging from 16 to 53% across years. The best fruit yield during the study period was around 18 kg ha⁻¹ in CSO I and 17 kg ha⁻¹ in CSO II. Highly significant clonal variation and clone by year and clone by site interactions were observed for fertility traits. The clonal contribution was more skewed in poor flowering years than in abundant flowering years and in CSO II than in CSO I. Broad sense heritability for flower and fruit production per tree was low to moderate (0.16 to 0.55). Flower and fruit production by individual ramets in successive years were positively correlated. Correlations between reproductive and growth traits were generally low, but correlation was strong between flowering and fruiting. Fertility variation and group coancestry were higher in poor flowering years than in abundant years and in CSO II than CSO I. Restricting seed collection to abundant flowering years,

Key words: Flowering, fruit production, diversity, coancestry, heritability, status number.

Introduction

Teak (Tectona grandis L.f.) is a high value tropical species acclaimed world over for its timber quality. Increased demand for timber and restrictions on felling from natural forests has prompted large scale planting of teak throughout the tropics (PANDEY and BROWN, 2000). Large quantities of seed are required every year to establish new plantations. Clonal seed orchards (CSO) were planted in India and other countries for domesticating and breeding teak and produce genetically improved seed. The first seed orchard was established in India during 1962 and since then more than 1000 ha of CSOs have been planted in different parts of the country (Kedharnath and Matthews, 1962; Katwal, 2005). Seed production and germination has been generally low in most of these orchards (INDIRA and BASHA, 1999; KJÆR et al., 1999; MATHEW and VASUDEVA, 2003). As a result, seed requirement for new plantations is met from seed production areas and existing plantations. Teak breeding programmes have not moved beyond the first generation clonal orchards.

Low seed production in teak orchards has prompted many studies on the reproductive behaviour of teak dur-

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adjusting ramet number to balance contribution of clones and mixing of seeds from successive years are suggested to reduce relatedness among orchard progeny. The usefulness of low input breeding options for teak like seed production areas are also discussed.

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