



DOI: 10.2478/rrlm-2014-0034

A glance on recent progresses in diagnosis and treatment of primary immunodeficiencies

Progrese recente în diagnosticul și tratamentul imunodeficiențelor primare

Peter J. Späth

Institute of Pharmacology, University of Berne, CH-3010 Berne, Switzerland

Abstract

Primary immunodeficiencies (PIDs) belong to the group of rare diseases which need more awareness by the relevant medical disciplines. Below a review on recent progresses in diagnosis and treatment of PIDs is given. Reducing the regrettable delay in diagnosis of PIDs (worldwide) is possible only when awareness is increased by doctors who may encounter patients with PID. This review shall serve this purpose. Progresses in understanding what the link might be between one genetic defect presenting in various phenotypes or how various gene defects may manifest by very similar PID phenotypes helps building awareness. Knowledge of PID favours early diagnosis, a cornerstone of optimal, sometimes life-long care at justifiable costs. The complexity of PIDs calls for clinical laboratory and clinical diagnostic performed by experts only. Exciting laboratory diagnostic progresses in early diagnosis of the most severe forms of PID are reviewed below. Progresses in curative therapies for PIDs, such as hematopoietic stem cell transplantation and gene therapies, are mentioned in short. About 80% of PID patients suffer from an antibody deficiency syndrome and can profit from non-curative replacement therapies with human immunoglobulin G concentrates. Modes of application, safety and hints for dosing of replacement therapies to reduce frequencies of severe infections are mentioned below. Thanks to the increasing quality of care, patients survive adolescence. A glance is given on the problems of transition to the adult medicine setting.*

** for abbreviations please consult the list of abbreviations at the end of the manuscript*

Keywords: *Awareness, early diagnosis, gene therapy, hematopoietic stem cell transplantation, immunoglobulin G concentrates, new-born screening, primary immunodeficiencies, replacement therapy*

Received: 6th July 2014; **Accepted:** 25th August 2014; **Published:** 6th September 2014.

*Corresponding author: Peter J Späth , Universitat Bern Bern, Switzerland, e-mail: peter.spaeth@pki.unibe.ch

Progress in understanding the pathophysiology of primary immunodeficiency diseases

Primary immunodeficiencies (PIDs)*, in their majority, relate to an impaired capacity of patient's immune system to fight infections, recurrent ones or infections with a very narrow range of pathogens (1). Some forms, when untreated, result in death early in life; others, in chronic debilitating infections and tissue remodelling. In addition, genetic defects identified more recently may result in an autoimmune-inflammatory phenotype of PID. Over 200 different genetic defects leading to PIDs have been described so far. With the progress of knowledge, the classification of PIDs has been updated regularly. However, the most forefront classification also has become an issue of debates because for the everyday clinical work such classification has become a topic able to be overlooked by much specialised clinicians and researchers only (2-4).

Patients with recurrent, severe or unusual infections steadily are fuelling clinical research, thereby continuously increasing the number of genetic defects underlying PIDs (5;6). Understanding the pathogenesis of PID is essential for optimal therapy of PID patients and cost effectiveness of care. National and international registries are excellent sources for important information. Due to the increasing numbers of patients included to the various registries, more and more precise statements can be made on how an early, optimal and effective treatment of PID patients might become possible.

Definitions of rare disease in the EU and in the USA are: an incidence of $\leq 1/2000$ (EU) or $\leq 200,000$ patients (USA). PIDs are rare diseases with a prevalence of 5-6 patients per 100,000 inhabitants in France (highest report in the EU) (7). A higher prevalence has been reported from countries with consanguine marriages (8). CVID

is the most common phenotype of PIDs and embraces a heterogeneous group of patients which either suffer from infections only or in addition from inflammation, granuloma, autoimmunity, colitis, cytopenia and malignancy (9-12). Ten to twenty percent of the CVID phenotype have a familial trait and in about 3% of CVID 10 different gene defects or polymorphisms have been identified in the last 10 years (deficiencies of BAFF-R, CD19, CD20, CD21, CD81, ICOS, LRBA, MSH5, NF- κ B2, PI3K δ , and TACI) (13;14). In addition to CVID, haematological malignancies, autoimmune-like, inflammatory, and malignant conditions are comorbidities of many other PID phenotypes (15).

Beside the classical PIDs, the non-classical PIDs are Mendelian conditions which are characterized by a very narrow spectrum of opportunistic infections – sometimes limited to one microbial genus or species – while presenting a normal development of the principal leukocyte subsets. These non-classical PIDs comprise susceptibility to mycobacteria (10 genetic defects identified so far), predisposition to neisserial infections (e.g. complement deficiencies) and invasive bacterial disease, predisposition to invasive fungal infections and chronic mucocutaneous candidiasis, predisposition to HPV infection, HSV encephalitis and EBV infection.

The recognition and diagnosis of PID form beside of a clinician's observation of an individual clinical and/or immunological phenotype remains difficult. Therefore, attempts to provide guidelines for clinicians at the bedside were published recently (16).

PID diagnosis - As early as possible

Shortening the delay in diagnosis of PID is the key for cost-effective therapies and is helping to keep health care costs reasonable without restricting patients' access to the relevant therapies (17;18). The platform for early diagnosis is

medical awareness. To increase awareness in all medical disciplines that may encounter patients with PID and the wide spreading of the ‘warning signs of primary immunodeficiency’ can be an effective tool (19;20). However, it has to be stressed that for taking the correct actions the widely promulgated “warning signs” have to be interpreted by experts (21). Other tools to raise awareness might be medical discipline-specific reviews (22-26) or data mining in national or international registries. National registries may not have the power of international registries (<http://esid.org/Working-Parties/Registry/ESID-Online-Registry>, accessed April 2014).

Hints for PID need a follow-up by standard diagnostic measures, e.g. serum immunoglobulin isotype levels plus specific response to vaccination, small lymphocyte panel plus B cell panel plus CD45RA CD4 T cells (+ T cell proliferation) (27). If the results are not conclusive, the next level might include specific membrane protein detection by flow cytometry (28-30). Established diagnostic criteria for SCID variants and recently published guideline for new-born screening are available (31;32).

Combined immunodeficiencies (CIDs) need to be recognized and separated from CVID. CIDs are T-cell impairments and can manifest as infections by intra/extracellular “opportunistic agents”. Even though sharing common clinical features, the discovery of new causative gene alterations led to the identification of novel complex clinical phenotypes of CID, with manifestations of autoimmunity/inflammation, allergy and lymphoma. CIDs represent about 20% of PIDs with approximately 80 different gene alterations (prevalence according to the French registry 1.1/100,000 or ~1/20,000 births). The detection of these alterations relies on nucleic acid sequencing methods and can prevent the death of affected children.

Population-based new-born screening (NBS) is a promising technique to detect from Guth-

rie cards severe combined immunodeficiency (SCID) or combined immunodeficiency (CID) based on the T-cell receptor (rearrangement) excision circles (TREC) assay (33). At present, the cut-off of the test is a problem because a wide spectrum of detected non-SCID disorders with T-cell lymphopenia. Although an evaluation of the population based SCID-NBS outside the US has started, and despite the proof of cost effectiveness of SCID-NBS, issues such as costs, ethics, moral, social, legal and policy remain the main barrier against wide implementation in countries other than the USA. Performing the TREC assay only might miss some forms of severe PIDs (34).

The potency of the population-based NBS for PIDs can be enhanced by introduction of the κ -deleting recombination excision circles (KREC) assay which allows for additional detection of B cell lymphopenia (35;36). The combination of both tests considerably expands the diagnostic range at a cost increase of about 0.10 Euro only. Furthermore, information from of TREC and KREC levels can also be used as a surrogate marker of lymphocyte output in acquired immunodeficiencies and it also might allow to follow therapy success (37). Laboratory methods for population-based dried spot analyses are expanding rapidly. Extension of the mass screening for PIDs not detectable by TREC and KREC assay might come at low cost from dried blood sample analysis by mass spectrometry (38)

The clinical and immunologic heterogeneity of CVID remains a diagnostic challenge (39;40). Recently a heterogeneous group of patients was classified according to B cell phenotype, KREC analysis, and SHM pattern. Five B-cell patterns were identified each reflecting an immunologically homogenous patient group with proposed unique pathophysiology. Another diagnostic challenge is the differential diagnosis between particular forms of CVID and CID. Time will

show whether the TREC, KREC assay or genome wide array analyses might be helpful in understanding clinical heterogeneity of CVID and dissect it from CID (14;41).

In 2006 the generation of “induced pluripotent stem cells” (iPSC) was reported for the first time by epigenetically reprogramming of somatic cells through the exogenous expression of transcription factors. The technique then could be adapted to human fibroblasts (42;43). Induced pluripotent stem cells (iPSCs) offer a unique potential for understanding the molecular basis of diseases and disease development. The study of primary immunodeficiencies (PIDs) has largely been based on animal models, in-vitro assays, and was suffering from the limited access to disease-specific tissue. Application of this technique one day might become an additional tool for functional investigation of human PIDs and other disorders as well as it might provide new medicines (44).

Optimizing replacement therapies at high safety levels

Immunoglobulin replacement therapies are the therapy of choice for the majority (~80%) of PID patients. Replacement therapy is not curative but can ensure over long-term acceptable Quality of Life. Depending on the country, there is a more or less broad offer of various immunoglobulin concentrates available (<http://www.ipopi.org/index.php?page=immunoglobulin-companies>; accessed May 2014).

Treatment of PID, as any other rare disease, requires expertise. A recent study in the northern hemisphere compared management of patients dependent on replacement therapies. US physicians with >10% of their practice devoted to primary immunodeficiency managed PID therapy very similarly to their colleagues in EU, while in US physicians whose clinical practice was composed of <10% of PID patients management pro-

ocols differed from the other two groups (45). Low percentage of PID patients treated in clinic or practice may not optimally comply with patients' need.

According to the American Academy of Allergy, Asthma and Immunology (AAAAI), replacement therapy is definitively beneficial for PIDs with absent B cells and PIDs with hypogammaglobulinaemia and impaired specific antibody production; it is probably beneficial for PIDs with normogammaglobulinaemia and impaired specific antibody production; and unlikely to be beneficial for isolated IgA and IgG4 deficiency (46). According to the EU PID Consensus Conference outcomes, the indication for replacement therapy is given for all patients with IgG of < 2g/L - with the exception of children without severe infectious complications (physiological hypogammaglobulinaemia of childhood); at levels of 2-5 g/L when associated with recurrent infections; and with IgG of > 5g/L when there is a deficiency in the formation of antibodies to specific antigens and serious or recurrent infections (http://ec.europa.eu/health/ph_projects/2005/action1/docs/action1_2005_exs_01_en.pdf; accessed April 2014).

In an environment of ever increasing use of immunoglobulin concentrates it has to be ascertained that PID patients have now and in future a continuous and sufficient access to this plasma derived medicinal product (47). Treatment based on evidence based practice guidelines (48;49) and Prioritisation Plans, of which the most have been compiled into a European consensus proposal (50), are valuable tools for an appropriate use and the best prioritisation of Ig concentrates in and outside the PID field. These documents can be adapted e.g. to the needs of Romanian patients and care givers.

The proportion of patients receiving SCIG is continuously increasing: new products, less frequent adverse events, sustainably higher immunoglobulin levels in the circulation and the

option of easy to perform home therapy thereby possibly reducing costs, at least in certain countries, support this development (51-54). SCIG can be applied with the help of a pump e.g. every two to three days, weekly, and three weekly to monthly facilitated by hyaluronidase or daily by "rapid push" (55-57). Availability of IVIG and SCIG allows for adapting replacement therapy to the clinical situation and the preference of the treating physician and/or how it is best suitable for the patient's individual needs (58). However, SCIG should be applied with care when risk factors exist (e.g. severe thrombocytopenia, bleeding disorders or anticoagulation therapy).

Dosing levels in replacement therapy are an eternally discussed topic. As a rule it is accepted that increasing the dose reduces risk for severe infections and this is true for the i.v. and the s.c. route of application of immunoglobulins (59;60). In a naïve patient, or a patient not having received replacement therapy for more than 3 months, the aim is to increase circulating IgG rapidly and this is achieved best by initially slow infusion via the i.v. route with doses starting at 0.4-0.6 g/kg b.w./3-4 weeks. Furthermore, evidence-based medicine data indicate as a rule that trough levels above 5-6 g IgG/L being "effective". However, there are patients who are not able to cope with these rules. Parameters identified so far for not being able to cope with the rules are (i) patient's residual levels of IgA, (ii) serum concentration of mannose-binding lectin, (iii) the polymorphism of the neonatal FcRn receptor, and (iv) B cell defects (61-65). In some patients the optimisation of replacement therapy might require measurement of e.g. anti-pneumococcal antibodies. An optimal maintenance therapy of PID patients is "individualized" and does not follow through levels and is not weight based, i.e. a biological level should be achieved and this level might need adaptation in winter-versus summer-time (66).

When shifting from IVIG to SCIG difference exist what US and European authorities consider as adequate (67;68). In daily practice and in general, mean actual levels of circulating IgG in patients has risen over the decades, and due to several factors tend to be higher when on SCIG. Furthermore, steadily increasing mean levels of IgG in the circulation of PID patients are driven by deeper insight into pathophysiology. Particularly autoimmune and inflammatory conditions associated with antibody deficiency syndromes may support orientation towards treatment doses corresponding to those applied for immunomodulation. Some caution is advised when discussing trough levels. Because of the difference in application intervals and doses, pharmacokinetics differs and trough levels might not be directly comparable.

Therapy of antibody deficiency syndromes-associated non-infectious complication

Beside recurrent respiratory and gastroenteric infections, autoimmune-like, inflammatory and malignant conditions in a part of PID patients are associated with increased mortality and pose major clinical challenges. The background for these complications remains unknown. For CVID differences in peripheral T- and B-cell compartments of patients might have some influence, i.e. disease-related complications appear to be more frequent in patients with naïve CD4⁺ T-cell defects (69). Optimal treatment of disease-related complication in CVID remains an unmet medical need (9;70). Treatment needs expertise to take the decision for watchful waiting vs active therapy if available at all. Some therapy options might be optimal IgG substitution, supportive therapy (parenteral nutrition), antibiotics, corticosteroids / Budesonide, immunosuppressive therapy (Methotrexate, Azathioprine, biosimilars (Rituximab, Infliximab), and transplantation. It remains unclear how far the com-

plications are infection-related/triggered. Multi-centre (observational) studies and data mining in registries are required to obtain answers on how best to treat these conditions.

Transition of PID patients to the clinic for adults

Transition, i.e. the purposeful, planned movement of adolescents and young adults with chronic physical and medical conditions from child-centred to adult-oriented health care systems, is a long-haul interdisciplinary project and remains an unresolved problem (71). Particular centres for PID/rare diseases might be best suited in supporting patients on their sometimes difficult journey into adulthood by e.g. establishing a “transition clinic”. After the transition phase or PIDs with late onset diseases/late diagnosed disease, patients might be confronted with difficulties to access specialists and obtain optimal therapy. Very recently, a scoring system was presented how to establish replacement therapy in adult PID (72).

Pathogen safety of replacement therapy

Measures requested by authorities such as current Good Manufacturing Practice (cGMP) and voluntary quality standards implemented by the plasma fractioning industry have pushed pathogen safety of immunoglobulin concentrates high and this is indicated by no reports of transmission of emerging viruses (SARS CoV, West Nile Virus and others), zoonotic pathogens or the agent of variant Creutzfeldt-Jacob disease in the last decade (73;74).

Adverse events related to replacement therapy

Adverse events to IVIG and less frequently to SCIG in their majority are mild to moderate in intensity and largely self-limiting. Adverse events to therapeutic immunoglobulin concentrates might be related to the active component

IgG, to impurities or to excipients in the preparations. The active component-related adverse events are inherent to replacement therapy. Inherence in part is due to the nature of our immune system, which on one hand serves host defence and on the other hand it also is part of the peripheral immunologic homeostasis network and serves removal of altered and senescent self. IgG concentrates derive from the immune system of several thousand healthy donors. Their application in patients results in the desired recognition and subsequent removal of possible pathogens but inevitably also results in the recognition of the recipient's variable-region connected immune network (75) and the recognition of altered or senescent tissue in an alloimmune fashion (76). This type of adverse events, as a consequence, is more patient than product dependent. Frequency of adverse events after IVIG infusion usually can be kept low by low infusion rates and avoiding some excipients in IVIGs (77). Severe adverse events are acute renal failure mainly due to osmotic nephrosis (predominantly by excipient), anaphylaxis, i.e. interaction of co-fractionated IgA in the product (impurity) with anti-IgA very rarely present in the plasma of the patient, aseptic meningitis, haemolysis i.e. due to anti-blood group antibodies (impurity), TEEs i.e. due to FXIa (impurity) and transfusion-related acute lung injury (78). Reports on increasing numbers of haemolysis and TEEs with severe consequences are a recent focus of concern. Both types of AEs are strongly related to the dose applied. TEEs have been found to be associated with elevated levels of activated coagulation factor XI (FXIa) and kallikrein (79) and resulted in the batch-wise testing and removal of FXIa (http://www.web-medcentral.com/article_view/2002; accessed April 2014). Despite all measures, patient- and administration-related factors for TEE remain; risk factors are age, underlying disease, co-morbidities, high protein doses and rapid infusion.

Haemolysis is a long recognized complication of IVIG therapy occurring at an apparent frequency of <1/100,000 patients treated and has been observed with intramuscular, subcutaneous and predominantly with intravenous preparations. In the last decade an increase in haemolysis rates were observed when infusing certain IVIG brands at immunomodulatory doses (80) while in a review of IVIG replacement trials an increase in direct Coombs test was observed without evidence of haemolysis (81) (for the courtesy of the reader: Gammagard liquid (US) corresponds to KOIVIG (EU)). Haemolysis can manifest as haemolytic anaemia or in isolated cases as haemolysis-related renal dysfunction/renal failure or disseminated intravascular coagulation and death. Isoagglutinins, e.g. A, B and AB have been attributed to mediate these higher frequencies (82) although release criteria of US & EU products include specifications for anti-A & anti-B antibodies of $\leq 1:64$ in the direct haemagglutinin assay. As IVIG-associated haemolysis occurs although products meet licensed specifications risk factors other than isoagglutinins are supposed, e.g. inflammatory conditions and rapid infusion. Meanwhile the plasma fractionating industry has taken measures to reduce isoagglutinin levels in immunoglobulin concentrates, e.g. by withholding from pooling high-titre donations and/or by introducing a polishing step which reduces isoagglutinins by immunoaffinity chromatography.

Vaccination and PID

Vaccination can be an excellent diagnostic tool in PID. Otherwise, PID vaccination cannot have a universal role because the difference in the various forms of PIDs. In patients with low antibody production vaccination might be considered in order to minimize the recurrence of episodes of infection using killed pathogens or subunits (refrain from the vaccination with (at-

tenuated) live vaccines!). There are only very few controlled studies reported with most data generated in children. The grade of treatment recommendation drawn from these studies remains C (possible effective; 2nd or 3rd line treatment) and in rare cases it might reach recommendation grade B (possibly effective, use as alternative to other therapy options). After having consulted a PID specialist, vaccination may be performed (www.cipo.ca/Vaccines.doc; accessed June 2014). Thus, dead influenza vaccine is safe, can be applied for seasonal prophylaxis but may not work well in patients with CVID while patients with IgA deficiency might profit from it. However, clinical improvement might be observed even in absence of detectable increase of protective antibody levels and this has resulted in best practice statement from the UK (http://www.rcpch.ac.uk/sites/default/files/asset_library/Publications/I/Immunocomp.pdf; accessed June 2014) and uncertainties together with the growing neglect of societal adherence to routine immunizations has prompted the Medical Advisory Committee of the Immune Deficiency Foundation (US) to issue recommendations based on published literature and the collective experience of the committee members (83). The prophylactic vaccination of household members of these patients groups is highly recommended.

Curative therapies for PID

Haematopoietic stem cell transplantation / bone marrow transplantation

Between 1968 and 2012 more than 2250 transplantations for SCID and more than 5000 for non-SCID PIDs have been performed (84). The methods applied are bone marrow transplantation, cord blood transplantation and machinery supported collection of HSCs. Very early transplantation (indicated by family history or population-based NBS) significantly improves out-

come irrespective of donor choice, conditioning regimen used, and underlying genetic diagnosis. Other factors of steadily and incrementally improving HSCT outcome encompass reduced intensity condition regiment, increasing expertise in ablation of certain donor cell populations in order to provide a balance between engraftment and graft-versus-host disease (GvHD), low dose serotherapy new drugs allowing reduction of veno-occlusive disease, close viral and fungal surveillance, and more sophisticated supportive care (85-87). Very recent US NBS data indicate that a prolonged time window for transplantation might be gained when timely TREC-NBS prompt immediate measures to avoid infections and organ damage. Transplantation in some CID can pose a considerable challenge as infection, autoimmunity/inflammation, allergy and lymphoma have to be taken into account.

Post HSCT viral infections are (severe) complications and also pose a major therapeutic challenge. Emerging therapies are Rituximab for EBV and CMX010 for CMV infections.

Patient-derived induced pluripotent stem cells

The iPSC technique allows the access to previously inaccessible diseased tissue and in turn allows the development of novel treatment strategies such as has been reported for X-linked CGD (88).

Gene therapy

Gene therapies are curative measures for PIDs (89-91). They belong to the emerging group of “personalised” therapies. Site-specific insertion and the development of safe and effective self-inactivating viral vectors have reduced the risk for post-transplant malignancies (insertional oncogenesis). The process involves the isolation of autologous haematopoietic stem cells (HSC), ex vivo introduction of the gene of interest by viral vector, expansion of cells and

‘transplantation’ of the gene-corrected HSC (92). The recent vectors are retroviruses, SIN retroviruses and lentiviruses engineered to have no potency for insertional oncogenesis (91;93). PID conditions treated so far or being in clinical studies are SCIDX1, ADA-SCID, CGD, and WAS. For the future all PID patients identified with genetic defects are potential candidates for gene therapy (94). This includes those 10 gene defects with the ‘CVID’ phenotype which have been identified in the last 10 years (see above). Attempts for scientific assessment for marketing authorization of ‘advanced therapy medicinal products’, e.g. of viral vectors based gene therapies, have been initiated (http://www.ema.europa.eu/ema/index.jsp?curl=pages/about_us/general/general_content_000126.jsp&mid=WC-0b01ac05800292a5; http://ec.europa.eu/health/human-use/advanced-therapies/developments/index_en.htm; <http://www.genetherapy.net.com/europe.html>; all accessed June 2014).

Conclusions

Developments in understanding pathogenesis, diagnosis and therapy of PIDs have progressed rapidly in the recent years. Particular note has to be given to the new screening techniques allowing diagnosis at a time point where no organ damage has occurred yet. Nevertheless, some problems remain such as the low awareness for PID or the emergence of complex ethical problems due to the new diagnostic tools. I hope this short review helps raising awareness for a rare disease where for a large part of the patients satisfying therapy options exist.

List of abbreviations

BAFF-R: B cell activating factor receptor (of the tumour necrosis factor family); tumour necrosis factor receptor superfamily member 13C; TNFRSF13C

BMT: bone marrow transplantation

CD: cluster of differentiation
 CID: combined immunodeficiency
 cGMP: current good manufacturing practice
 CMV: cytomegalovirus
 CVID: common variable immunodeficiency
 EBV: Epstein-Barr virus
 ESID: European Society for Immunodeficiencies
 FcRn: Fc receptor of the newborn; Brambell receptor
 GvHD: graf-versus-host disease
 HPV: human papilloma virus
 HSCT: hematopoietic stem-cell transplantation
 HSV: herpes simplex virus
 ICOS: inducible (T-cell) co-stimulator
 iPSC: induced pluripotent stem cells
 IVIG: human immunoglobulin G concentrate applicable via the intravenous route
 KREC: κ -deleting recombination excision circle
 LRBA: lipopolysaccharide-responsive and beige-like anchor protein
 MSH5: human homolog of Escherichia coli MutS 5
 NBS: new-born screening
 NF- κ B: nuclear factor kappa-light-chain-enhancer of activated B cells
 PI3K δ : phosphatidylinositol-3-OH kinase PI(3)K catalytic subunit delta
 PID: primary immunodeficiency
 PIDD: primary immunodeficiency disease
 SARS-CoV: severe acute respiratory syndrome corona virus
 SCID; severe combined immunodeficiency
 SCIG: human immunoglobulin G concentrate applied via the subcutaneous route
 SHM: somatic hyper mutation
 SIN: self-inactivating
 TACI: transmembrane activator and calcium modulator cyclophilin ligand interactor; tumor necrosis factor receptor superfamily member 13B; TNFRSF13B;
 TEE: thromboembolic event
 TREC: T-cell receptor (rearrangement) excision circle

References

1. Casanova JL, Abel L. The genetic theory of infectious diseases: A brief history and selected illustrations. *Annu Rev Genomics Hum Genet.* 2013;14:215-43. DOI: 10.1146/annurev-genom-091212-153448
2. Pandolfi F, Milito C, Conti V, Pagliari D, Frosali S, Cianci R, et al. Common variable immunodeficiency - New insight into the pathogenesis and the quest for a workable classification. *J Biol Regul Homeost Agents.* 2013;27(2):285-9.
3. Maggina P, Gennery AR. Classification of primary immunodeficiencies: need for a revised approach? *J Allergy Clin Immunol.* 2013 Feb;131(2):292-4. DOI: 10.1016/j.jaci.2012.10.008
4. Al-Herz W, Bousfiha A, Casanova J-L, Chatila T, Conley ME, Cunningham-Rundles C, et al. Primary immunodeficiency diseases: An update on the classification from the International Union of immunological societies expert committee for primary immunodeficiency. *Front Immunol.* 2014;5(APR). DOI: 10.3389/fimmu.2014.00162
5. Chinen J, Notarangelo LD, Shearer WT. Advances in basic and clinical immunology in 2013. *J Allergy Clin Immunol.* 2014 Feb 28;133(4):967-76. DOI: 10.1016/j.jaci.2014.01.026
6. Hernandez-Trujillo V. New genetic discoveries and primary immune deficiencies. *Clin Rev Allergy Immunol.* 2014;46(2):145-53. DOI: 10.1007/s12016-013-8380-0
7. Gathmann B, Binder N, Ehl S, Kindle G. The European internet-based patient and research database for primary immunodeficiencies: Update 2011. *Clin Exp Immunol.* 2012;167(3):479-91. DOI: 10.1111/j.1365-2249.2011.04542.x
8. Bousfiha AA, Jeddane L, Ailal F, Benhsaien I, Mahlaoui N, Casanova JL, et al. Primary immunodeficiency diseases worldwide: more common than generally thought. *J Clin Immunol.* 2013 Jan;33(1):1-7. DOI: 10.1007/s10875-012-9751-7
9. Chapel H, Lucas M, Lee M, Björkander J, Webster D, Grimbacher B, et al. Common variable immunodeficiency disorders: division into distinct clinical phenotypes. *Blood.* 2008 Jul 15;112(2):277-86. DOI: 10.1182/blood-2007-11-124545
10. Wehr C, Kivioja T, Schmitt C, Ferry B, Witte T, Eren E, et al. The EUROclass trial: defining subgroups in common variable immunodeficiency. *Blood.* 2008 Jan 1;111(1):77-85. DOI: 10.1182/blood-2007-06-091744
11. Baldovino S, Montin D, Martino S, Sciascia S, Menegatti E, Roccetello D. Common variable immunodeficiency: crossroads between infections, inflammation and autoimmunity. *Autoimmun Rev.* 2013 Jun;12(8):796-801. DOI: 10.1016/j.autrev.2012.11.003
12. Gathmann B, Mahlaoui N, Gerard L, Oksenhendler E,

- Warnatz K, Schulze I, et al. Clinical picture and treatment of 2212 patients with common variable immunodeficiency. *J Allergy Clin Immunol*. 2014 Feb 27;Epub ahead of print. DOI: 10.1016/j.jaci.2013.12.1077
13. Eibel H, Salzer U, Warnatz K. Common variable immunodeficiency at the end of a prospering decade: Towards novel gene defects and beyond. *Curr Opin Allergy Clin Immunol*. 2010;10(6):526-33. DOI: 10.1097/ACI.0b013e328333fea1c
 14. Keller MD, Jyonouchi S. Chipping away at a mountain: Genomic studies in common variable immunodeficiency. *Autoimmun Rev*. 2013;12(6):687-9. DOI: 10.1016/j.autrev.2012.10.017
 15. Todoric K, Koontz JB, Mattox D, Tarrant TK. Autoimmunity in immunodeficiency. *Curr Allergy Asthma Rep*. 2013;13(4):361-70. DOI: 10.1007/s11882-013-0350-3
 16. Bousfiha AA, Jeddane L, Ailal F, Al HW, Conley ME, Cunningham-Rundles C, et al. A phenotypic approach for IUIS PID classification and diagnosis: guidelines for clinicians at the bedside. *J Clin Immunol*. 2013 Aug;33(6):1078-87. DOI: 10.1007/s10875-013-9901-6
 17. Modell V, Gee B, Lewis DB, Orange JS, Roifman CM, Routes JM, et al. Global study of primary immunodeficiency diseases (PI)-diagnosis, treatment, and economic impact: an updated report from the Jeffrey Modell Foundation. *Immunol Res*. 2011 Oct;51(1):61-70. DOI: 10.1007/s12026-011-8241-y
 18. Routes J, Abinun M, Al-Herz W, Bustamante J, Condino-Neto A, de la Morena MT, et al. ICON: The early diagnosis of congenital immunodeficiencies. *J Clin Immunol*. 2014 May;34(4):398-424. DOI: 10.1007/s10875-014-0003-x
 19. Modell F, Puente D, Modell V. From genotype to phenotype. Further studies measuring the impact of a Physician Education and Public Awareness Campaign on early diagnosis and management of Primary Immunodeficiencies. *Immunol Res*. 2009;44(1-3):132-49. DOI: 10.1007/s12026-008-8092-3
 20. Borte S, Borte M. Decreasing time-to-diagnosis in primary antibody deficiencies - The case for newborn screening programmes and awareness campaigns. *Eur Infect Dis*. 2012;6(2):99-104.
 21. MacGinnitie A, Aloï F, Mishra S. Clinical characteristics of pediatric patients evaluated for primary immunodeficiency. *Pediatr Allergy Immunol*. 2011;22(7):671-5. DOI: 10.1111/j.1399-3038.2011.01167.x
 22. Motamed F, Aghamohammadi A, Soltani M, Mansouri M, Rezaei N, Teimourian S, et al. Evaluation of liver diseases in Iranian patients with primary antibody deficiencies. *Ann Hepatol*. 2009 Jul;8(3):196-202.
 23. Dehkordy SF, Aghamohammadi A, Ochs HD, Rezaei N. Primary immunodeficiency diseases associated with neurologic manifestations. *J Clin Immunol*. 2012;32(1):1-24. DOI: 10.1007/s10875-011-9593-8
 24. Agarwal S, Mayer L. Diagnosis and treatment of gastrointestinal disorders in patients with primary immunodeficiency. *Clin Gastroenterol Hepatol*. 2013;11(9):1050-63. DOI: 10.1016/j.cgh.2013.02.024
 25. Hosseinverdi S, Hashemi H, Aghamohammadi A, Ochs HD, Rezaei N. Ocular involvement in primary immunodeficiency diseases. *J Clin Immunol*. 2014;34(1):23-38. DOI: 10.1007/s10875-013-9974-2
 26. Lehman H. Skin manifestations of primary immune deficiency. *Clin Rev Allergy Immunol*. 2014;46(2):112-9. DOI: 10.1007/s12016-013-8377-8
 27. Wood P. Primary antibody deficiencies: Recognition, clinical diagnosis and referral of patients. *Clin Med (Lond Engl)*. 2009;9(6):595-9. DOI: 10.7861/clinmedicine.9-6-595
 28. Abraham RS. Relevance of laboratory testing for the diagnosis of primary immunodeficiencies: A review of case-based examples of selected immunodeficiencies. *Clin Mol Allergy*. 2011;9:6. DOI: 10.1186/1476-7961-9-6
 29. de Vries E, European Society for Immunodeficiencies (ESID) members. Patient-centred screening for primary immunodeficiency, a multi-stage diagnostic protocol designed for non-immunologists: 2011 update. *Clin Exp Immunol*. 2012 Jan;167(1):108-19. DOI: 10.1111/j.1365-2249.2011.04461.x
 30. Locke BA, Dasu T, Verbsky JW. Laboratory diagnosis of primary immunodeficiencies. *Clin Rev Allergy Immunol*. 2014 Apr;46(2):154-68. DOI: 10.1007/s12016-014-8412-4
 31. Borte S, Von Döbeln U, Hammarström L. Guidelines for newborn screening of primary immunodeficiency diseases. *Curr Opin Hematol*. 2013;20(1):48-54. DOI: 10.1097/MOH.0b013e32832835a9130
 32. Shearer WT, Dunn E, Notarangelo LD, Dvorak CC, Puck JM, Logan BR, et al. Establishing diagnostic criteria for severe combined immunodeficiency disease (SCID), leaky SCID, and Omenn syndrome: The Primary Immune Deficiency Treatment Consortium experience. *J Allergy Clin Immunol*. 2014;133(4):1092-8. DOI: 10.1016/j.jaci.2013.09.044
 33. Chan K, Puck JM. Development of population-based newborn screening for severe combined immunodeficiency. *J Allergy Clin Immunol*. 2005 Feb;115(2):391-8. DOI: 10.1016/j.jaci.2004.10.012
 34. Janik DK, Lindau-Shepard B, Comeau AM, Pass KA. A multiplex immunoassay using the Guthrie specimen to detect T-cell deficiencies including severe combined immunodeficiency disease. *Clin Chem*. 2010 Sep;56(9):1460-5. DOI: 10.1373/clinchem.2010.144329
 35. Borte S, Von Döbeln U, Fasth A, Wang N, Janzi M, Winiarski J, et al. Neonatal screening for severe primary

- ry immunodeficiency diseases using high-throughput triplex real-time PCR. *Blood*. 2012;119(11):2552-5. DOI: 10.1182/blood-2011-08-371021
36. Chiarini M, Zanotti C, Serana F, Sottini A, Bertoli D, Caimi L, et al. T-cell receptor and K-deleting recombination excision circles in newborn screening of T- and B-cell defects: Review of the literature and future challenges. *J Public Health Res*. 2013;2(1):9-16. DOI: 10.4081/jphr.2013.e3
37. Serana F, Chiarini M, Zanotti C, Sottini A, Bertoli D, Bosio A, et al. Use of V(D)J recombination excision circles to identify T- and B-cell defects and to monitor the treatment in primary and acquired immunodeficiencies. *J Transl Med*. 2013;11:119. DOI: 10.1186/1479-5876-11-119
38. la Marca G. Mass spectrometry in clinical chemistry: the case of newborn screening. *J Pharm Biomed Anal*. 2014 Apr 28;E pub ahead of print. DOI: 10.1016/j.jpba.2014.03.047
39. Ameratunga R, Woon ST, Gillis D, Koopmans W, Steele R. New diagnostic criteria for common variable immune deficiency (CVID), which may assist with decisions to treat with intravenous or subcutaneous immunoglobulin. *Clin Exp Immunol*. 2013;174(2):203-11. DOI: 10.1111/cei.12178
40. Seppänen M, Aghamohammadi A, Rezaei N. Is there a need to redefine the diagnostic criteria for common variable immunodeficiency? *Expert Rev Clin Immunol*. 2014;10(1):1-5. DOI: 10.1586/1744666X.2014.870478
41. Nijman IJ, Van Montfrans JM, Hoogstraat M, Boes ML, Van De Corp, Renner ED, et al. Targeted next-generation sequencing: A novel diagnostic tool for primary immunodeficiencies. *J Allergy Clin Immunol*. 2014;133(2):529-34. DOI: 10.1016/j.jaci.2013.08.032
42. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*. 2006 Aug 25;126(4):663-76. DOI: 10.1016/j.cell.2006.07.024
43. Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, Tomoda K, et al. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*. 2007 Nov 30;131(5):861-72. DOI: 10.1016/j.cell.2007.11.019
44. Pessach IM, Ordovas-Montanes J, Zhang SY, Casanova JL, Giliani S, Gennery AR, et al. Induced pluripotent stem cells: A novel frontier in the study of human primary immunodeficiencies. *J Allergy Clin Immunol*. 2011;127(6):1400-7. DOI: 10.1016/j.jaci.2010.11.008
45. Hernandez-Trujillo HS, Chapel H, Lo RI, V, Notarangelo LD, Gathmann B, Grimbacher B, et al. Comparison of American and European practices in the management of patients with primary immunodeficiencies. *Clin Exp Immunol*. 2012 Jul;169(1):57-69. DOI: 10.1111/j.1365-2249.2012.04588.x
46. Orange JS, Hossny EM, Weiler CR, Ballow M, Berger M, Bonilla FA, et al. Use of intravenous immunoglobulin in human disease: A review of evidence by members of the Primary Immunodeficiency Committee of the American Academy of Allergy, Asthma and Immunology. *J Allergy Clin Immunol*. 2006 Apr;117(4 Suppl):S525-S553. DOI: 10.1016/j.jaci.2006.01.015
47. Stonebraker JS, Farrugia A, Gathmann B, ESID Registry Working Party, Orange JS. Modeling primary immunodeficiency disease epidemiology and its treatment to estimate latent therapeutic demand for immunoglobulin. *J Clin Immunol*. 2014 Feb;34(2):233-44. DOI: 10.1007/s10875-013-9975-1
48. Robinson P, Anderson D, Brouwers M, Feasby TE, Hume H. Evidence-based guidelines on the use of intravenous immune globulin for hematologic and neurologic conditions. *Transfus Med Rev*. 2007 Apr;21(2 Suppl 1):S3-S8. DOI: 10.1016/j.tmr.2007.01.004
49. Shehata N, Palda V, Bowen T, Haddad E, Issekutz TB, Mazer B, et al. The use of immunoglobulin therapy for patients with primary immune deficiency: an evidence-based practice guideline. *Transfus Med Rev*. 2010 Jan;24(Suppl. 1):S28-S50. DOI: 10.1016/j.tmr.2009.09.011
50. Sewell WC, Kerr J, Behr-Gross ME, Peter HH. European consensus proposal for immunoglobulin therapies. *Eur J Immunol*. 2014 Jun 28. DOI: 10.1002/eji.201444700
51. Beauté J, Levy P, Millet V, Debré M, Dudoit Y, Le Mignot L, et al. Economic evaluation of immunoglobulin replacement in patients with primary antibody deficiencies. *Clin Exp Immunol*. 2010;160(2):240-5. DOI: 10.1111/j.1365-2249.2009.04079.x
52. Lingman-Framme J, Fasth A. Subcutaneous immunoglobulin for primary and secondary immunodeficiencies: An evidence-based review. *Drugs*. 2013;73(12):1307-19. DOI: 10.1007/s40265-013-0094-3
53. Torgerson TR, Bonagura VR, Shapiro RS. Clinical ambiguities - Ongoing questions. *J Clin Immunol*. 2013;33(2 SUPPL.):S99-S103. DOI: 10.1007/s10875-012-9851-4
54. Martin A, Lavoie L, Goetghebeur M, Schellenberg R. Economic benefits of subcutaneous rapid push versus intravenous immunoglobulin infusion therapy in adult patients with primary immune deficiency. *Transfus Med*. 2013;23(1):55-60. DOI: 10.1111/j.1365-3148.2012.01201.x
55. Shapiro R. Subcutaneous immunoglobulin therapy by rapid push is preferred to infusion by pump: a retrospective analysis. *J Clin Immunol*. 2010 Mar;30(2):301-7. DOI: 10.1007/s10875-009-9352-2
56. Wasserman RL, Melamed I, Stein MR, Gupta S, Puck J, Engl W, et al. Recombinant human hyaluronidase-fa-

- cilitated subcutaneous infusion of human immunoglobulins for primary immunodeficiency. *J Allergy Clin Immunol.* 2012;130(4):951-7. DOI: 10.1016/j.jaci.2012.06.021
57. Chapel H, Gardulf A. Subcutaneous immunoglobulin replacement therapy: The European experience. *Curr Opin Allergy Clin Immunol.* 2013;13(6):623-9. DOI: 10.1097/ACI.000000000000013
58. Espanol T, Prevot J, Drabwell J, Sondhi S, Olding L. Improving current immunoglobulin therapy for patients with primary immunodeficiency: Quality of life and views on treatment. *Patient Preference Adherence.* 2014;8:621-9. DOI: 10.2147/PPA.S60771
59. Orange JS, Grossman WJ, Navickis RJ, Wilkes MM. Impact of trough IgG on pneumonia incidence in primary immunodeficiency: A meta-analysis of clinical studies. *Clin Immunol.* 2010 Oct;137(1):21-30. DOI: 10.1016/j.clim.2010.06.012
60. Orange JS, Belohradsky BH, Berger M, Borte M, Hagan J, Jolles S, et al. Evaluation of correlation between dose and clinical outcomes in subcutaneous immunoglobulin replacement therapy. *Clin Exp Immunol.* 2012 Aug;169(2):172-81. DOI: 10.1111/j.1365-2249.2012.04594.x
61. Oksenhendler E, Gérard L, Fieschi C, Malphettes M, Mouillot G, Jaussaud R, et al. Infections in 252 patients with common variable immunodeficiency. *Clin Infect Dis.* 2008;46(10):1547-54. DOI: 10.1086/587669
62. Llobet MP, Soler-Palacin P, Detkova D, Hernandez M, Caragol I, Espanol T. Common variable immunodeficiency: 20-yr experience at a single centre. *Pediatr Allergy Immunol.* 2009 Mar;20(2):113-8. DOI: 10.1111/j.1399-3038.2008.00744.x
63. Quinti I, Soresina A, Guerra A, Rondelli R, Spadaro G, Agostini C, et al. Effectiveness of immunoglobulin replacement therapy on clinical outcome in patients with primary antibody deficiencies: Results from a multicenter prospective cohort study. *J Clin Immunol.* 2011;31(3):315-22. DOI: 10.1007/s10875-011-9511-0
64. Gregersen S, Aaløkken TM, Mynarek G, Fevang B, Holm AM, Ueland T, et al. Development of pulmonary abnormalities in patients with common variable immunodeficiency: associations with clinical and immunologic factors. *Annals of Allergy, Asthma and Immunology.* 2010;104(6):503-10. DOI: 10.1016/j.anai.2010.04.015
65. Gouilleux-Gruart V, Chapel H, Chevret S, Lucas M, Malphettes M, Fieschi C, et al. Efficiency of immunoglobulin G replacement therapy in common variable immunodeficiency: Correlations with clinical phenotype and polymorphism of the neonatal Fc receptor. *Clin Exp Immunol.* 2013;171(2):186-94. DOI: 10.1111/cei.12002
66. Bonagura VR, Marchlewski R, Cox A, Rosenthal DW. Biologic IgG level in primary immunodeficiency disease: the IgG level that protects against recurrent infection. *J Allergy Clin Immunol.* 2008 Jul;122(1):210-2. DOI: 10.1016/j.jaci.2008.04.044
67. Berger M, Rojavin M, Kiessling P, Zenker O. Pharmacokinetics of subcutaneous immunoglobulin and their use in dosing of replacement therapy in patients with primary immunodeficiencies. *Clin Immunol.* 2011;139(2):133-41. DOI: 10.1016/j.clim.2011.01.006
68. Wasserman RL. Progress in gammaglobulin therapy for immunodeficiency: from subcutaneous to intravenous infusions and back again. *J Clin Immunol.* 2012;32(6):1153-64. DOI: 10.1007/s10875-012-9740-x
69. Mouillot G, Carmagnat M, Gérard L, Garnier JL, Fieschi C, Vince N, et al. B-cell and T-cell phenotypes in CVID patients correlate with the clinical phenotype of the disease. *J Clin Immunol.* 2010;30(5):746-55. DOI: 10.1007/s10875-010-9424-3
70. Resnick ES, Moshier EL, Godbold JH, Cunningham-Rundles C. Morbidity and mortality in common variable immune deficiency over 4 decades. *Blood.* 2012;119(7):1650-7. DOI: 10.1182/blood-2011-09-377945
71. Rapley P, Davidson PM. Enough of the problem: a review of time for health care transition solutions for young adults with a chronic illness. *J Clin Nurs.* 2010 Feb;19(3-4):313-23. DOI: 10.1111/j.1365-2702.2009.03027.x
72. Agarwal S, Cunningham-Rundles C. Treatment of hypogammaglobulinemia in adults: A scoring system to guide decisions on immunoglobulin replacement. *J Allergy Clin Immunol.* 2013;131(6):1699-701. DOI: 10.1016/j.jaci.2013.01.036
73. O'Mahony B, Turner A. The Dublin Consensus Statement 2011 on vital issues relating to the collection and provision of blood components and plasma-derived medicinal products. *Vox Sang.* 2012 Feb;102(2):140-3. DOI: 10.1111/j.1423-0410.2011.01528.x
74. Späth PJ, Van Holten RW, Kempf C. Pathogen safety of immunoglobulin preparations. In: Wahn V, Orange J, editors. *Clinical Use of Immunoglobulins.* 2nd ed. Bremen - London - Boston: UNI-MED Verlag; 2013. p.26-50.
75. Vassilev TL, Bineva IL, Dietrich G, Kaveri SV, Kazatchkine MD. Variable region-connected, dimeric fraction of intravenous immunoglobulin enriched in natural autoantibodies. *J Autoimmun.* 1995 Jun;8(3):405-13. DOI: 10.1006/jaut.1995.0032
76. Späth PJ, Lutz HU. Naturally occurring antibodies/autoantibodies in polyclonal immunoglobulin concentrates. Lutz HU, editor. *Naturally Occurring Antibodies (NABs).* *Advances in Experimental Medicine and Biology*[750], 239-261. 16-4-2012. Austin, TX USA, Lan-

- des Bioscience and Springer Science+Business Media. *Advances in Experimental Medicine and Biology*.
77. Berger M. Adverse effects of IgG therapy. *J Allergy Clin Immunol. Pract.* 2013;1(6):558-66. DOI: 10.1016/j.jaip.2013.09.012
 78. Gürçan HM, Keskin DB, Ahmed AR. Information for healthcare providers on general features of IGIV with emphasis on differences between commercially available products. *Autoimmun Rev.* 2010;9(8):553-9. DOI: 10.1016/j.autrev.2010.03.003
 79. Etscheid M, Breiter-Ruddock S, Gross S, Hunfeld A, Seitz R, Dodt J. Identification of kallikrein and FXIa as impurities in therapeutic immunoglobulins: implications for the safety and control of intravenous blood products. *Vox Sang.* 2012 Jan;102(1):40-6. DOI: 10.1111/j.1423-0410.2011.01502.x
 80. Desborough MJ, Miller J, Thorpe SJ, Murphy MF, Misbah SA. Intravenous immunoglobulin-induced haemolysis: a case report and review of the literature. *Transfus Med.* 2013 Oct 25;E-pub ahead of print. DOI: 10.1111/tme.12083
 81. Schroeder HW, Jr., Dougherty CJ. Review of intravenous immunoglobulin replacement therapy trials for primary humoral immunodeficiency patients. *Infect.* 2012;40(6):601-11. DOI: 10.1007/s15010-012-0323-9
 82. Dhainaut F, Guillaumat PO, Dib H, Perret G, Sauger A, De Coupade C, et al. In vitro and in vivo properties differ among liquid intravenous immunoglobulin preparations. *Vox Sang.* 2013;104(2):115-26. DOI: 10.1111/j.1423-0410.2012.01648.x
 83. Medical Advisory Committee of the Immune Deficiency Foundation, Shearer WT, Fleisher TA, Buckley RH, Ballas Z, Ballow M, et al. Recommendations for live viral and bacterial vaccines in immunodeficient patients and their close contacts. *J Allergy Clin Immunol.* 2014 Apr;133(4):961-6. DOI: 10.1016/j.jaci.2013.11.043
 84. Worth AJJ, Booth C, Veys P. Stem cell transplantation for primary immune deficiency. *Curr Opin Hematol.* 2013;20(6):501-8. DOI: 10.1097/MOH.0b013e-328365a13b
 85. Cavazzana-Calvo M, Andre-Schmutz I, Fischer A. Haematopoietic stem cell transplantation for SCID patients: Where do we stand? *Br J Haematol.* 2013;160(2):146-52. DOI: 10.1111/bjh.12119
 86. Güngör T, Teira P, Slatter M, Stussi G, Stepensky P, Moshous D, et al. Reduced-intensity conditioning and HLA-matched haemopoietic stem-cell transplantation in patients with chronic granulomatous disease: a prospective multicentre study. *Lancet.* 2014 Feb 1;383(9915):436-48. DOI: 10.1016/S0140-6736(13)62069-3
 87. Lane JP, Evans PTG, Nademi Z, Barge D, Jackson A, Hambleton S, et al. Low-dose serotherapy improves early immune reconstitution after cord blood transplantation for primary immunodeficiencies. *Biol Blood Marrow Transplant.* 2014;20(2):243-9. DOI: 10.1016/j.bbmt.2013.11.005
 88. Zou J, Sweeney CL, Chou BK, Choi U, Pan J, Wang H, et al. Oxidase-deficient neutrophils from X-linked chronic granulomatous disease iPS cells: functional correction by zinc finger nuclease-mediated safe harbor targeting. *Blood.* 2011 May 26;117(21):5561-72. DOI: 10.1182/blood-2010-12-328161
 89. Cavazzana-Calvo M, Fischer A, Hacein-Bey-Abina S, Aiuti A. Gene therapy for primary immunodeficiencies: Part 1. *Curr Opin Immunol* 2012 Oct;24(5):580-4. DOI: 10.1016/j.coi.2012.08.008
 90. Aiuti A, Bacchetta R, Seger R, Villa A, Cavazzana-Calvo M. Gene therapy for primary immunodeficiencies: Part 2. *Curr Opin Immunol* 2012 Oct;24(5):585-91. DOI: 10.1016/j.coi.2012.07.012
 91. Fischer A, Hacein-Bey-Abina S, Cavazzana-Calvo M. Gene therapy of primary T cell immunodeficiencies. *Gene.* 2013;525(2):170-3. DOI: 10.1016/j.gene.2013.03.092
 92. Touzot F, Hacein-Bey-Abina S, Fischer A, Cavazzana M. Gene therapy for inherited immunodeficiency. *Expert Opin Biol Ther.* 2014 Jun;14(6):789-98. DOI: 10.1517/14712598.2014.895811
 93. Farinelli G, Capo V, Scaramuzza S, Aiuti A. Lentiviral vectors for the treatment of primary immunodeficiencies. *J Inher Metab Dis.* 2014 Mar 12;Epub ahead of print. DOI: 10.1007/s10545-014-9690-y
 94. Mukherjee S, Thrasher AJ. Gene therapy for PIDs: progress, pitfalls and prospects. *Gene.* 2013 Aug 10;525(2):174-81. DOI: 10.1016/j.gene.2013.03.098