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# An Evaluation of Serum Procalcitonin and C-Reactive Protein Levels as Diagnostic and Prognostic Biomarkers of Severe Sepsis

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#### **ABSTRACT**

**Background**: Recommendations have been made, following the multicenter Surviving Sepsis Campaign study, to standardize the definition of severe sepsis with reference to several parameters such as haemodynamic stability, acid-base balance, bilirubin, creatinine, International Normalized Ratio (INR), urine output and pulmonary functional value of the ratio between arterial oxigen partial pressure and inspiratory oxigen concentration. Procalcitonin (PCT) is considered to be a gold standard biomarker for the inflammatory response, and recent studies have shown that it may help to discover whether a seriously ill person is developing sepsis. C-reactive protein (CRP) is also used as a marker of inflammation in the body, as its blood levels increase if there is any inflammation in the body. The aim of this study was to evaluate serum procalcitonin and C-reactive protein levels as diagnostic and prognostic biomarkers of severe sepsis.

**Material and method**: Sixty patients, diagnosed as being "septic", were admitted to the intensive care unit (ICU). Based on laboratory results and clinical findings a diagnosis of "severe sepsis" was made, and correlated with PCT and CRP values. The APACHE II, SAPS II and SOFA severity scores were calculated, analyzed and correlated with PCT and CRP.

**Results**: Fifty two patients (86.67%) presented with criteria for severe sepsis. Multivariate correlation analysis indicated a significant positive association between procalcitonin and all severity scores (APACHEII p<0.0001, SOFA p<0.0001, SAPS II p<0.0001). CRP proved to be significantly correlated only with the SAPS II score (p=0.0145). Mortality rate was high, with 48 patients (80%) dying. There was no significant correlation between the levels of the PCT and CRP biomarkers and severe sepsis (p=0.2059 for PCT, p=0.6059 for CRP).

**Conclusions**: The procalcitonin levels are highly correlated with the severity scores (APACHE II, SAPS II, SOFA) regularly used in ICUs and therefore can be used for determining the severity of the septic process. Quantitive procalcitonin and C-reactive protein analysis was not shown to be useful in diagnosing severe sepsis. However, PCT and CRP can be used to predict the fatal progression of the septic patient.

**Keywords**: severe sepsis, procalcitonin, C-reactive protein, prognostic biomarkers

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#### INTRODUCTION

Sepsis is a systemic inflammatory response (SIRS) of the host to several infections. It can lead to severe sepsis which remains a principal cause of death in Intensive Care Units (ICU) worldwide, with one in four patients dying of severe sepsis or septic shock [1-5]. The early diagnosis and decided management can lead to lower mortality rates and improved outcomes [6].

Recent studies have focused on the use of an ideal biomarker for the diagnosis and prognostic of sepsis [7-9]. Of the many studied biomarkers, procalcitonin

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remains the gold standard for sepsis diagnosis, and is considered by some clinicians to be an indicator of the severity of clinical sepsis [10].

C reactive protein is another biomarker used to diagnose both chronic and acute inflammatory responses. In patients diagnosed as having sepsis or septic shock, it may be used to establish the severity of these conditions and in the same time to establish a prognosis [11,12].

In 2013 a large multicenter study (Surviving Sepsis Campaign) was conducted targeting the septic patient, with the aim of establishing new definitions and management for sepsis. The Surviving Sepsis Campaign lists the main criteria for defining both sepsis and severe sepsis. According to these guidelines, sepsis is defined as "the presence (probable or documented) of infection together with systemic manifestations of infection", while severe sepsis is defined as "sepsis plus sepsis-induced organ dysfunction or tissue hypoperfusion" [6,13,14].

For the diagnosis of sepsis, the systemic inflammatory response criteria remain mostly unchanged including general variables such as fever (>38.3°C) or hypothermia (<36°C), tachycardia >90/min, tachypnea, altered mental status, significant edema or positive fluid balance (>20 mL/kg over 24 hr). The laboratory criteria used include hyperglycemia (>140 mg/dL), leukocytosis (>12,000/ $\mu$ L) or leukopenia (<4000/ $\mu$ L), C-reactive protein and procalcitonin levels, both more than twice the normal value [6].

To arrive at a diagnosis of severe sepsis the presence of the following criteria were advocated: sepsis-induced hypotension, metabolic acidosis with high lactate, urine output <0.5 mL/kg/h, acute lung injury with PaO2/FIO2 <200, creatinine >2.0 mg/dL, bilirubin >2 mg/dL, platelet count <100,000 / $\mu$ L, international normalized ratio INR >1.5 [7].

Procalcitonin (PCT) and C reactive protein (CRP) values are considered to be of paramount importance for a diagnosis of sepsis, but there is lack of evidence about the role of these two markers in severe sepsis and in arriving at a prognosis following a diagnosis of severe sepsis, with reference to the new definitions as stated in the Surviving Sepsis Campaign study.

# Aim of study

The aim of this study is to investigate the utilization of procalcitonin and C reactive protein levels in the diagnosis of severe sepsis and to determine their potential prognostic roles.

# ■ MATERIAL AND METHODS

A prospective study was conducted in the Clinic of Intensive Care of the County Clinical Emergency Hospital Tîrgu Mureş from January to November 2014. The approval of the Ethics Committee of the University of Medicine and Pharmacy of Tîrgu Mureş was obtained prior to the commencement of the study. Written informed consent was obtained from all participants or their relatives before including them into the study.

Inclusion criteria: All adult (>18 years) admitted to the ICU with SIRS, presenting with at least two positive criteria for sepsis, and who had not received any surgical intervention in the previous seventy two hours or had cardiac arrest or trauma, were admitted into the study.

Exclusion criteria: Adult (>18 years) patients with non-infectious SIRS, admitted to the ICU during the same time period.

Patients were diagnosed with sepsis using the following standard clinical criteria: temperature: fever (>38.3°C) or hypothermia (<36°C), heart rate over 90/min, respiratory status: tachipnoe over 24 resp/min or mechanically ventilated, haemodynamic status: systolic blood pressure under 90 mmHg after fluid resuscitation and leukocyte count: leukocytosis (>12,000 /µL) or leukopenia (<4000 /µL).

Sepsis was defined as present when at least two of the criteria for the systemic inflammatory response syndrome (SIRS) and clinical or microbiological evidence infection were present.

Sixty eight patients presented with a diagnose of SIRS and of these sixty, aged between 20 and 90 years, were diagnosed with a proven infection. Eight patients were excluded due to lack of confirmation of an infectious etiology of SIRS. Out of sixty patients, thirty one (51.67%) were females and twenty nine (48.33%) males.

On admission to the ICU, a medical history was taken, the clinical status was monitored and recorded, including blood pressure, diuresis, SpO2, ECG and ventilation parameters such as current volume, respiratory rate and ventilation mode. Laboratory tests included leucocytes, platelets counts, creatinine, urea, INR, bilirubin and bacteriological determinations. Blood cultures were performed. The APACHE II (Acute Physiology and Chronic Health Evaluation), SOFA (Sequential Organ Failure Assessment) and SAPS II (Simplified Acute Physiology Score) severity scores were calculated.

The criteria for the severe sepsis in the study group are presented in Table 1.

Within twelve hours of being admitted to the ICU, blood samples were obtained via vascular puncture for PCT and CRP analysis and stored in serum tubes without anticoagulant. The samples were centrifuged and the serum were collected and frozen at -70°C for later processing. Blood cultures were attained using separate vials for aerobic (Standard SA) and anaerobic (Standard SN) organisms. Analyses of the blood cultures were processed using the BacT/Alert 3D (Biomerieux, France) automated hemoculture system.

PCT, CRP and the sepsis-related organ failure assessment (SOFA) were determined using the immuneturbidimetry method (Cobas 6000, Roche Diagnostics, Germany) and Promo Kinekits (PromoCell GmbH, Heidelberg, Germany) for the detection, elimination and prevention of cell culture contamination.

The antropometric characteristics of the patients were measured and the body mass index (BMI) was calculated. The correlations between BMI and both PCT and CRP levels were determined, as was the correlation between BMI, death and severe sepsis presence.

Surviving patients were seen six months after admission to the ICU and the outcomes were recorded.

Statistical analysis was performed using Microsoft Excel (Microsoft, Washington, USA), GraphPad (GraphPad Software, Inc., California, USA) and MedCalc (MedCalc Software, Ostend, Belgium).

Analysis included specificity and sensibility of PCT and CRP for diagnosis of severe sepsis, and correlation with death. Graphic representation of receiver-operating-characteristic (ROC) were used, with deter-

Table 1. Severe sepsis criteria

Criteria	N (%)
Without severe sepsis	8 (13.33%)
Severe sepsis	52 (86.67%)
Hypotension*	18
Metabolic acidosis **	23
Urine output <0.5 mL/kg/hr	17
Acute lung injury with PaO2/FIO2 <200	30
Creatinine >2.0 mg/dL,	22
Bilirubin >2 mg/dL,	3
Platelet count <100,000 /μL,	5
International normalized ratio INR >1.5	18

<sup>\*</sup> Hypotension = MAP <70 Hgmm

mination of area under the curve (AUC). Quantitative variables were tested for normal distribution using the Kolmogorov-Smirnov test and Bartlett's Test for equal variances. Where applicable the Mann-Whitney or Pearson tests were used for variable correlations. Pearson's chi2 test with Fisher or Yates correction were used to compare the distribution of nominal values. A significance level of alpha ( $\alpha$ ) =0.05 was applied.

# RESULTS

The demographic and clinical characteristics of the group are represented in Table 2.

Sixty patients met the criteria for sepsis and of these fifty two (86.67%) presented criteria for severe sepsis.

Thirty patients (57.69%) with severe sepsis were further identified with acute lung injury, twenty three patients (44.23%) with metabolic acidosis, twenty two (42.3%) with elevated serum creatinine levels, and eighteen patients (34.61%) were hypotensive.

The mean serum PCT level in the severe sepsis group (86.67%) was  $1.367\pm2.437$  ng/mL, and the mean serum CRP level  $172\pm124.1$ ng/mL.

No significant correlations were found between laboratory assays (PCT, CRP,INR, PLT, Bi) and severe sepsis (Table 3).

Positive blood cultures were found in 16.66% of patients. The commonest site of infection was the lungs (41 patients) (Table 4).

According to multivariate correlation analysis, there was a significant positive association between procalcitonin serum levels and all severity scores (APACHE II p<0.0001, SOFA p<0.0001, SAPS II p<0.0001). There was no significant correlation between CRP and SOFA score (p=0.2889), but there was a significant positive association with APACHE II (p=0.0058) and SAPS II (p=0.0145).

Mortality rate was high for all septic patients, forty eight patients deceased within 7 weeks of being admitted to the study (80%). The patients presented, too, several associated pathologies (Table 5). The survival rate was higher for those patients without severe sepsis criteria, 62.5% of patients without severe sepsis criteria surviving versus 13.46% of patiets with severe sepsis criteria. (Figure 1).

No significant correlation was found between the serum levels of PCT (p=0.2059) or CRP (p=0.6059) and severe sepsis.

<sup>\*\*</sup> pH<7.25 and NaHCO<sub>3</sub><22 mmol/l

Table 2. Demographic and clinical characteristic of the septic patients

	Minimum	Maximum	Mean	Std. Deviation	Skewness	Kurtosis
A = a (1,100m)						
Age (years)	20	90	69.27	15.28	-1.326	1.86
BMI (kg/m2)	17.8	46.9	26.7	5.99	1.239	2.136
TI days	0	42	8.1	9.24	1.783	2.874
MV days	0	34	5.61	7.20	2.273	5.316
APACHE II	10	50	27.6	8.54	0.5398	0.1158
SOFA	0	16	8.15	3.45	0.1336	-0.333
SAPS II	15	90	47.68	18.93	0.5062	-0.4605
INR	0.99	4.4	1.49	0.59	3.288	12.76
Urea (mg/dl)	1.99	452	110.6	89.37	1.478	2.838
Creatinine (mg/dl)	0.24	14.55	2.43	2.64	2.405	7.316
WBC (1000 * count/μl)	2.8	57.6	19.8	11.43	1.525	2.498
Hct (%)	12.8	57.9	34.48	9.37	0.1685	-0.2682
Hgb (mg/dl)	4.41	18	11.07	2.87	0.1311	-0.2663
AST (mg/dl)	7	860	76.15	135.2	4.012	19.53
ALT (mg/dl)	10	492	79.55	100.6	2.613	6.927
CRP (ng/ml)	1.96	545.2	170	124.1	1.205	1.227
PCT (ng/ml)	0.00196	13.24	1.34	2.43	3.384	12.99
Bi (mg/dl)	0.25	4	0.94	0.80	2.809	8.5
Plt (1000 * count/μl)	75	555	238.8	119.9	0.8479	0.01687

BMI= Body Mass Index TI days = days of intensive care MV days = mechanical ventilation days APACHE II = Acute Physiology and Chronic Health Evaluation SOFA = Sepsis-related Organ Failure Assessment

SAPS II = Simplified Acute Physiology Score INR = International normalized ratio WBC = white blood cell count Hct = hematocrit Hgb = hemoglobin

Hgb = hemoglobin
AST = aspartate aminotransferase
ALT = alanine aminotransferase
CRP = C-reactive protein

PCT = procalcitonin Bi = bilirubin Plt = platelet count

Table 3. Severe sepsis laboratory findings

Se	evere sepsis (n=52)	Without severe se	epsis (n=8)	_
Mean	Std. Deviation	Mean	Std. Deviation	p
0.98	0.84	0.66	0.42	0.1874
233.1	125.7	275.8	65.64	0.0970
1.53	0.63	1.25	0.15	0.2094
2.67	2.76	0.83	0.50	0.0102
172	124.9	156.7	125.8	0.6059
1.36	2.45	1.17	2.49	0.2059
	Mean 0.98 233.1 1.53 2.67 172 1.36	MeanStd. Deviation0.980.84233.1125.71.530.632.672.76172124.91.362.45	Mean         Std. Deviation         Mean           0.98         0.84         0.66           233.1         125.7         275.8           1.53         0.63         1.25           2.67         2.76         0.83           172         124.9         156.7           1.36         2.45         1.17	Mean         Std. Deviation         Mean         Std. Deviation           0.98         0.84         0.66         0.42           233.1         125.7         275.8         65.64           1.53         0.63         1.25         0.15           2.67         2.76         0.83         0.50           172         124.9         156.7         125.8           1.36         2.45         1.17         2.49

Table 4. Localization of infectious process

Site of infection	Patients (n)
Pulmonar	41
Abdominal cavity	8
Cutanat	5
Urinary tract	4
Central nervous system	1
Undefined	1

Table 5. List of associated pathology of our group f patients

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Pathology	N
hypertension	27
cardiac failure	19
chronic renal disfunction	14
diabetes	13
cerebral vascular disease	7
COPD	6
chronic hepatic disfunction	3

COPD = chronic obstructive pulmonary disease

Figure 1. Survival curve at 6 months for patients with severe sepsis vs sepsis

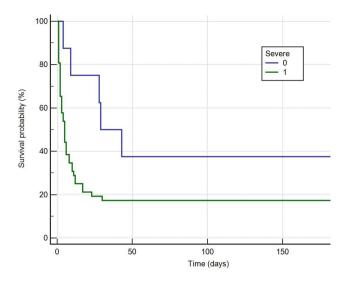
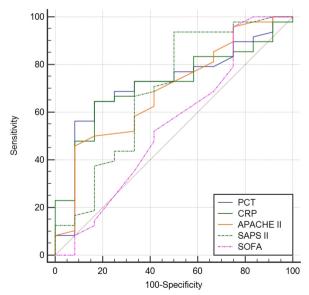


Figure 1. Survival curve at 6 months for patients with severe sepsis vs sepsis

A significant correlation existed between the death rate and serum levels of PCT (p=0.0180) or CRP (p=0.0195). A graphic interpretation of the comparative ROC curves for PCT, CRP and severity scores is represented by Figure 2.



	AUC	Sensitivity	Specificity	P value	Cut off value*	SE
PCT	0.720	64.58	83.33	0.0065	0.2576	0.0807
CRP	0.717	64.58	83.33	0.0036	121.65	0.0746
APACHE_II	0.689	45.83	91.67	0.0284	28	0.0863
SAPS_II	0.692	50.00	93.75	0.0543	27	0.0997
SOFA	0.546	25.00	95.83	0.6653	3	0.106

Figure 2. Comparative ROC curves for PCT, CRP, APACHE II, SAPS II and SOFA for predicting mortality

Correlation analysis showed no significant association between BMI and PCT levels (p=0.2473).

Neither was there a significant correlation found between BMI and severe sepsis (p=0.419) or between BMI and mortality (p=0.3512). However there was a significant correlation between BMI and CRP serum levels (p=0.02).

A positive correlations existed between the white blood cell count and CRP serum levels (p=0.0418), but not between the white blood cell count and PCT serum levels (p=0.3427).

# DISCUSSIONS

Data are lacking on the precise role of PCT and CRP in diagnosing severe sepsis and for their potential use as a prognostic indicator in cases of the severe sepsis.

There was no significant difference of mean serum PCT and CRP values between those cases with severe sepsis versus patients with sepsis. This is in agreement with results of other studies [15].

The present study is also in agreement with previous data which shoes that respiratory infections are invariably the most common cause of sepsis and severe sepsis [15]. Ventilator acquired pneumonia is a common cause of intensive care infections worldwide [16]. Cutaneous infections were also common in our study (5 patients, 8.3%), patients presenting several pressure ulcers. One of the most important determinant factor for pressure ulcers is the difficulty of adequate mobilization of the patients due to several associated conditions such as mechanical ventilation, pathologies which requires immobilization, deep coma or high sedation.

Haemocultures revealed a high rate of nosocomial infections among our group. Seven patients (70%) out of 10 positive haemocultures were found to have Staphyloccocus Methicilin Resitant (MRSA) (n=3), Acinetobacter (n=2), KlebsiellaPenumonie (n=3) and Pseudomonas Aeruginosa (n=2). One patient was identified to present simultaneously with positive haemocultures of Staphylococcus Aureus MRSA (Methilicilin resistant) and Pseudomonas Aeruginosa, one patient with Staphylococcus Aureus MRSA and Klebsiella Pneumonie and another one with Pseudomonas Aeruginosa and Klebsiella Pneumonieplus Acinetobacter. One of the most frequently observed severe infection was with MRSA, a fact also observed by other authors [17].

The data from the present study indicated that PCT and CRP serum levels have a prognostic value for death rate (PCT p=0.0180, CRP p=0.195). PCT is correlated with the severity scores APACHE II (p<0.0001), SOFA (p<0.0001) and SAPS II (p<0.0001), and is in agreement with previously recorded findings [18].

The high mortality rate in the present study may be explained by the severe status of the patients who had a mean APACHE II value 27.6±8.54, with fifty two of the patients (86.67%) above the value of 19, most of them presenting various severe associated pathology (Table 5).

The serum levels of PCT and CRP were measured within twenty four hours of admission and this corresponded with previous studies [19].

There was no significant correlation between PCT and CRP levels and the presence of severe sepsis, the ROC curve analysis revealed an AUC of 0.641 for PCT and of 0.558 for CRP, therefore the use of these biomarkers for establishing the diagnosis of severe sepsis should not be recommended. For predicting the prognosis of sepsis both biomarkers proved to be accurate, and our data indicated a cut off value of 0.2576 ng/ml for PCT and 121.65 ng/ml for CRP in predicting a fatal outcome for patients (Figure 2).

In the present study, the survival rate was higher for those patients not diagnosed with severe sepsis. Other studies indicated that patients with clinical criteria for severe sepsis but without positive blood cultures died, and patients with a documented infection are at similarly high risk of death [20].

Regarding the significant correlation between CRP and BMI (p=0.0201), this has been also reported previously in a group of young healthy individuals [21,22].

The serum levels of PCT and CRP are very highly correlated (p<0.0001) and as both of them are biomarkers of inflammation, this finding is to be expected. However the high correlation between PCT serum levels and APACHE II, SAPS II and SOFA severity scores in parallel with the lack of correlation between CRP serum levels and SOFA severity score, suggests the superiority of PCT for assessing the severity of the septic patients, and this is in agreement within previously stated conclusions [23,24].

# **■** CONCLUSIONS

The procalcitonin serum levels are highly correlated with APACHE II, SAPS II and SOFA severity scores

used in intensive care units and can therefore be used for determining the severity of the septic process.

Quantitive procalcitonin and C-reactive protein analysis did not indicate their usefulness in the diagnosis of severe sepsis, however procalcitonin can be used for predicting the fatal outcome of the septic patient.

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# **■** CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

## ABBREVIATIONS

APACHE II - Acute physiology and chronic health evaluation

AUC - Area under curve

BMI - Body mass index

COPD: chronic obstructive pulmonary disease

CRP - C reactive protein

ELISA - Enzyme-linked immunosorbent assay

MRSA - Methicilin resistant staphylococcus aureus

PCT - Procalcitonin

SAPS - Simplified acute physiology score

SIRS – Systemic inflammatory response syndrome

SOFA - Sequential Organ Failure Assessment

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