

Helminths in rodents from Wet Markets in Thailand

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Summary

Only a few surveys have ever been carried out of the helminths of the commensal rodents found in the traditional wet markets that play such an important part of daily life in South-east Asia. The potential of rodents as reservoirs of zoonoses including helminths is of great interest since in these markets humans and rodents come into closer contact than in other environments and food may be indirectly contaminated via rodent faeces. Helminths in a total of 98 rats belonging to two species (*Rattus norvegicus* and *Rattus exulans*) were surveyed in eight traditional wet markets in Udon Thani, Thailand. Thirteen species of helminths were recovered, seven of which are potentially zoonotic, with an overall prevalence of 89.8 %. Our results show that rodents in wet markets could pose a threat to human health as potential reservoirs of zoonotic helminthiasis.

Keywords: Norwegian rat; Pacific rat; zoonoses; wet markets

Introduction

In South-east Asia (SEA) the traditional wet markets that sell fresh and cooked products for human consumption are an important part of daily life. Stalls are enclosed in permanent roofed structures that provide a stable habitat – i.e. food is available all year – for the successful survival and reproduction of commensal rodents. The study of possible rodent-borne zoonoses is of interest as it has been reported previously that in the same study sites (Ribas, *et al.*, 2016) rodents are carriers and potential transmitters of *Salmonella* to humans. The role of rodents as reservoirs of helminth zoonoses in SEA has been reviewed by Chaisiri *et al.* (2015). The prevalence of helminths in humans in Thailand should not be underestimated as a prevalence of 18.1 % was found by the last National Survey (Wongsaroj, 2015); some of these helminths are associated with rodents, the host species that are the object of the present study. Surveys of helminths in wild rodents in SEA including Thailand

have become increasingly common in recent years. Yet, although surveys of the rodents in anthropized habitats such as villages and associated habitats (rice fields and plantations) have been conducted, data from urban environments is still very limited. To date, studies of rodents in wet markets in SEA are limited to just two studies. Firstly, five wet markets were studied in Kuala Lumpur in insular Malaysia by Paramasvaran *et al.* (2009), who detected 97 rodents belonging to three species: *R. r. diardii*, a taxon that, according Pagès *et al.* (2010), should be considered as a *Rattus* sp. pending a definitive name, *R. norvegicus* Berkenhout, 1769 and *R. exulans* (Peale, 1848). A second study in insular SEA in the Philippines by Claveria *et al.* (2005) is of limited interest as its sampling effort (twelve rats) was low and rats were not identified to species level (hosts were simply described as *Rattus* spp.); in addition these authors provided no details of the number of studied wet markets. Finally, a third study in Taiwan, outside SEA but still in Asia (Tung *et al.*, 2013), analyzed 51 rats (*R. norvegicus* and *R. rattus*) in traditional wet markets. Aside from these three surveys,

no other information was found despite an extensive literature review (Veciana, 2016). Also of interest is the study by Mohd Zain *et al.* (2012) of *R. norvegicus* and *R. rattus* Linnaeus, 1758 (probably *Rattus* sp. according Pagès *et al.* (2010)) in Kuala Lumpur; however in this latter study collection sites were not characterized. Thus, in light of this lack of previous information, the aim of the present study was to fill the gaps in the knowledge of the role of rodents as reservoirs of zoonotic helminthiases in traditional wet markets in SEA.

Materials and Methods

In mid-July to mid-August 2014 traps for rodents were set in eight traditional wet markets in Udon Thani (Udon Thani province, Thailand) (see methodological details in Ribas *et al.*, 2016). Trapped animals were euthanized and dissected following international standards (American Veterinary Medical Association Council on Research). These protocols, which maximize animal care and prioritize the health and safety of field parasitologists, generate good-quality data (Herbreteau, *et al.*, 2011). The study was approved by the Udon Thani Rajabhat University Animal Care and Ethical Use Committee. The viscera (GI tract, liver and lungs) were separated and dissected under a binocular microscope to identify and count helminths. Cestodes were stained with Semichon acetocarmine, dehydrated in alcohol, cleared in xylene and mounted in Canada balsam. Nematodes and acanthocephalans were mounted temporarily in Amann lactophenol. All helminths were identified by their morphology and morphometry according to the literature. Parasitization by *Capillaria hepatica* (Bancroft, 1893) was diagnosed macroscopically by the observation of whitish livers and then confirmed microscopically (categorized as positive or negative) by the observation of the typically bi-polar eggs in lactophenol. Liver cysts were dissected to confirm the presence of *H. taeniaeformis* larvae, identified by their characteristic rostellar hooks (Lavikainen *et al.*, 2016). Some of the recovered helminth species have not yet been identified to species level since more material is still needed for full identification and comparison, and because our aim was essentially to study zoonotic helminths. The ecological terminology and quantitative parameters used followed Bush *et al.* (1997). Descriptive parameters and confidence intervals were calculated using Quantitative parasitology (Reiczigel & Rózsa, 2005). In addition to the morphological and metrical traits (Aplin *et al.*, 2003; Marshall, 1988), for host identification a piece of muscle tissue from each rodent was preserved in absolute ethanol to use the CO1 region for molecular identification (details of protocol in (Ribas *et al.*, 2016).

Results

A total of 98 rodents were collected from the eight sampled markets. The molecular study of the hosts confirmed the presence of two rat species, 88 Norwegian rats (*R. norvegicus*) and 10 Pacific

rats (*R. exulans*). The number of rodents collected in each market varied from seven to 21. The overall prevalence of gastrointestinal helminth infection was 89.8 % (95 % CI: 82.0 – 95.0). The thirteen helminth species recovered included four cestodes, seven nematodes and one acanthocephalan (see details in Table 1).

Discussion

The two studied rodents are synanthropic and act as reservoirs in the spread of several diseases (Morand *et al.*, 2015; Wells *et al.*, 2015). Thus, both species are likely to be found in cities in SEA. The aforementioned studies of SEA markets (Claveria *et al.*, 2005) (Paramasvaran *et al.*, 2009; Tung *et al.*, 2013), as well as in urban habitats (Mohd Zain *et al.*, 2012), report the presence of the same zoonotic helminths as recovered in the present study (see details in Table 1).

According to the review by Chaisiri *et al.* (2015), four of our recovered helminths (*H. diminuta*, *H. nana*, *Raillietina* sp. and *M. moniliformis*) are potentially zoonotic species. A fifth gastrointestinal helminth (*G. neoplasicum*), not studied by Chaisiri *et al.* (2015), is also a potentially zoonotic species and has been reported in humans in Thailand by Pasuralertsakul *et al.* (2008). Nevertheless, reported cases of infestation in humans by this species worldwide are limited to just 40 – 50 reports (Wilson *et al.*, 2001). We found a sixth zoonotic species, the lung nematode *A. cantonensis*, that was not included in the review by Chaisiri *et al.* (2015) given that these authors only studied gastrointestinal helminths. This nematode causes human eosinophilic meningitis, and human cases have been reported in Thailand, mostly in the northeastern provinces (Eamsobhana, 2013) such as our study area. Pipitgool *et al.* (1997) report the presence of *A. cantonensis* in snails in Udon Thani province and in rodents; nevertheless, the methodology used in these surveys is not described in detail (i.e. it is not possible to identify the surveyed habitats or the host to species level). The market where this lungworm was found backs onto railway tracks and an unpaved area with soil and vegetation that could allow snails to survive and act as intermediate hosts. A seventh potentially zoonotic species is the liver nematode *C. hepatica* that in humans can cause the rare hepatic capillariosis, a sanitary problem caused by consuming food contaminated with parasite eggs. In our study sites humans are usually infected after ingesting embryonated eggs in fecal-contaminated food (Tesana *et al.*, 2007). High prevalences of *C. hepatica* in wet markets in comparison with rodents in the wild are evident in SEA. Brown *et al.* (1975) surveyed 6,498 rodents in Indonesia and only 31 (0.5 %) were positive for *C. hepatica*. A study by Liat *et al.* (1977) in Malaysia analyzed 1,258 rodents and detected higher prevalences in *R. norvegicus* (8/26) captured in houses and lower prevalences in forest rodents (1.54 – 8 %). Nevertheless, the habitats in which captures were performed are not well defined and so it is not easy to compare these author's results with those of the present study. In the Brazilian Amazon, Camargo *et al.* (2010) used indirect

Table 1. Recovered helminths in the studied markets from Udon Thani, Thailand. Z: zoonotic helminth; R.e.: *Rattus exulans*; R.n: *Rattus norvegicus*; P%: general prevalence, CI: clopper-Pearson confidence interval; MI: general mean intensity; Z: zoonotic helminth. Comparison with previous studies of zoonotic helminths in wet markets: 1: Claveria et al., 2005; 2: Paramasavaran et al., 2009; 3: Tung et al., 2013.

| | Host | General prevalence | CI (Clopper-Pearson) | MI | Zoonotic | 1 | 2 | 3 |
|---|---------|--------------------|----------------------|-------|----------|---|---|---|
| <i>Gongylonema neoplasticum</i> (Fibiger & Ditlevson, 1914) Ransom & Hall, 1916 | R.e/R.n | 38.8 | 29.1 – 49.2 | 4.68 | Z | - | * | - |
| <i>Raillietina</i> sp. | R.n | 34.7 | 25.4 – 45.0 | 6.18 | Z | - | * | * |
| <i>Hymenolepis nana</i> (Siebold, 1852) | R.n | (8) 8.2 | 3.6 – 15.5 | 1.38 | Z | - | * | * |
| <i>Hymenolepis diminuta</i> (Rudolphi, 1819) Weinland, 1858 | R.n | (5) 5.1 | 1.7 – 11.5 | 1.60 | Z | * | * | * |
| Trichostrongylidae | R.n | (35) 35.7 | 26.3 – 46.0 | 36.86 | - | - | - | - |
| <i>Syphacia muris</i> (Yamaguti, 1935) | R.e/R.n | (2) 2.0 | 0.2 – 7.2 | 25.0 | - | - | - | - |
| Capillariinae | R.n | (6) 6.1 | 2.3 – 12.9 | 11.17 | - | - | - | - |
| <i>Pterygodromatites tani</i> (Hoepli, 1929) | R.n | (3) 3.1 | 0.6 – 8.7 | 3.67 | - | - | - | - |
| <i>Physaloptera gnoci</i> Le-Van-hoa, 1961 | R.n | 1 (1) | 0.0 – 5.6 | 2.0 | - | - | - | - |
| <i>Hydatigera taeniaeformis</i> L. (Batsch, 1786) Lamarck, 1816 | R.n | (28) 28.6 | 19.9 – 38.6 | 1.89 | - | - | - | - |
| <i>Moniliiformis moniliformis</i> (Bremser, 1811) | R.e | (1) 1.0 | 0.0 – 5.6 | 3.00 | Z | * | * | - |
| <i>Angiostrongylus cantonensis</i> (Chen, 1935) Dougerthy, 1946 | R.n | (3) 3.1 | 0.6 – 8.7 | 10.00 | Z | - | - | * |
| <i>Capillaria hepatica</i> (Bancroft, 1893) | R.e/R.n | (63) 64.3 | - | - | Z | * | * | * |

immunofluorescence techniques to establish an association between infected rodents and the incidence of hepatic capillariasis in workers in markets where there is a high incidence of *C. hepatica* in rodents. Although the high prevalence of *C. hepatica* in commensal rodents may increase the health risk in humans, to date human seroprevalence has not been examined in SEA.

Our study detected all the rodent-borne zoonotic nematodes and cestodes reported by Chaisiri *et al.* (2015) except *Cyclodontosomum purvisi*. Thus, there can be no doubt that in the studied wet markets there is a potentially high risk of rodent-borne transmission. The cockroaches that have been observed in the studied markets act as mechanical dispersers of helminth eggs (Young and Babero 1975; Chamavit *et al.*, 2011). In addition, cockroaches also probably play an essential role in maintaining the life cycles of several species of helminths in wet markets as the majority of isolated helminth species require an arthropod as an intermediate host. Thus, the elimination of cockroaches could be an indirect way of reducing the number of rodent-borne helminths in wet markets. In conclusion, our results highlight the need to intervene to reduce rodent populations in wet markets in Thailand and in other countries where these markets are commonplace given that the rats that inhabit them are potential reservoirs of zoonotic diseases.

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References

- APLIN, K.P., BROWN, P.R., JACOB, J., KREBS, C.J., SINGLETON, G.R. (2003): *Field Methods for Rodent Studies in Asia and the Indo-Pacific*. Canberra, Australia, ACIAR Monograph, 100 pp.
- BROWN, R.J., CARNEY, W.P., VAN, PEENEN, P.F., CROSS, J.H., SAROSO, J.S. (1975): Capillariasis in wild rats of Indonesia. *Southeast Asian J. Trop. Med. Public Health*, 6(2): 219 – 222
- BUSH, A.O., LAFFERTY, K.D., LOTZ, J.M., SHOSTAK, A.W. (1997): Parasitology meets ecology on its own terms: Margolis *et al.* revisited. *J. Parasitol.*, 83(4): 575 – 583
- CAMARGO, L.M., CAMARGO, J., VERA, L.J., BARRETO, P., TOURINHO, E.K., DE SOUZA, M.M. (2010): Capillariasis (Trichurida, Trichinellidae, *Capillaria hepatica*) in the Brazilian Amazon: low pathogenicity, low infectivity and a novel mode of transmission. *Parasit. Vectors*, 3(11). DOI:10.1186/1756-3305-3-11
- CH AISIRI, K., SIRIBAT, P., RIBAS, A., MORAND, S. (2015): Potentially zoonotic helminthiasis of murid rodents from the Indo-Chinese Peninsula: impact of habitat and the risk of human infection. *Vector Borne and Zoonotic Dis.*, 15(1): 73 – 85. DOI:10.1089/vbz.2014.1619
- CHAMAVIT, P., SAHAISOOK P., NIAMNUY, N. (2011): The majority of cockroaches from the Samutprakarn Province of Thailand are carriers of parasitic organisms." *EXCLI Journal*, 10: 218 – 222
- CLAVERIA, F.G., CAUSAPIN, J., DE GUZMAN, M.A., TOLEDO, M.G., SALIBAY, C. (2005): Parasite biodiversity in *Rattus* spp. caught in wet markets. *Southeast Asian J. Trop. Med. Public Health*, 36(suppl 4): 146 – 148
- EAMSOBHANA, P. (2013): Angiostrongyliasis in Thailand: epidemiology and laboratory investigations. *Hawaii J. Med. Public Health*, 72(6 Suppl 2): 28 – 32
- HERBRETEAU, V., JITTAPALAPONG, S., RERKAMNUAYCHOKE, W., CHAVAL, Y., COSSON, J.F. MORAND, S. (2011): *Protocols for Field and Laboratory Rodent Studies*. Bangkok, Thailand, Kasetsart University Press, 56 pp.
- LAVIKAINEN, A., IWAKI, T., HAUKISALMI, V., KONYAEV, S.V., CASIRAGHI, M., DOKUCHAEV, N.E., GALIMBERTI, A., HALAJIAN, A., HENTTONEN, H., ICHIKAWA-SEKI, M., ITAGAKI, T., KRIVOPALOV, A.V., MERI, S., MORAND, S., NÄREAHÖ, A., OLSSON, G.E., RIBAS, A., TEREFE, Y., NAKAO, M. (2016): Reappraisal of *Hydatigera taeniaeformis* (Batsch, 1786) (Cestoda: Taeniidae) sensu lato with description of *Hydatigera kamiyai* n. sp. *Int. J. Parasitol.* DOI:10.1016/j.ijpara.2016.01.009
- LIAT, L.B., FONG, Y.L., KRISHNASAMY, M. (1977): *Capillaria hepatica* infection of wild rodents in Peninsular Malaysia. *Southeast Asian J. Trop. Med. Public Health*, 8(3): 354 – 58
- MARSHALL, J.T. (1988): Family Muridae: Rats and Mice. In: LEKAGUL, B., MC NEELY, J.A. (Eds) *Mammals of Thailand*, Bangkok, Thailand: Association for the Conservation of Wildlife, Sahakarnbhat Co, pp. 397 – 487
- MOHD ZAIN, S.N., BEHNKE, J.M., LEWIS, J.W. (2012): Helminth communities from two urban rat populations in Kuala Lumpur, Malaysia. *Parasit. Vectors*, 5(1): 47. DOI:10.1186/1756-3305-5-47
- MORAND, S., BORDES, F., CHEN, H.W., CLAUDE, J., COSSON, J.F., GALAN, M., CZIRJÁK, G.Á., GREENWOOD, A.D., LATINNE, A., MICHAUX, J., RIBAS, A. (2015): Global parasite and *Rattus* rodent invasions: the consequences for rodent-borne diseases no title. *Integr. Zool.*, 10(5): 409 – 423. DOI: 10.1111/1749-4877.12143
- PAGÈS, M., CHAVAL, Y., HERBRETEAU, V., WAENGSOOTHORN, S., COSSON, J.F., HUGOT, J.P., MORAND, S., MICHAUX, J. (2010): Revisiting the taxonomy of the Rattini tribe: a phylogeny-based delimitation of species boundaries. *BMC Evol. Biol.*, 10: 184. DOI:10.1186/1471-2148-10-184
- PARAMASVARAN, S., SANI, R.A., HASSAN, L., HANJEET, K., KRISHNASAMY, M., JOHN, J., SANTHANA, R., SUMARNI, M.G., LIM, K.H. (2009): Endo-parasite fauna of rodents caught in five wet markets in Kuala Lumpur and its potential zoonotic implications. *Trop. Biomed.*, 26(1): 67 – 72
- PASURALERTSAKUL, S., YAICHAROEN, R., SRIPOCHANGE, S. (2008): Spurious human infection with *Gongylonema*: nine cases reported from Thailand. *Ann. Trop. Med. Parasitol.*, 102(5): 455 – 457. DOI:10.1179/136485908X300869
- PIPITGOOL, V., SITHITHAWORN, P., PONGMUTTASAYA, P., HINZ, E. (1997): *Angiostrongylus* infections in rats and snails in northeast Thailand. *Southeast Asian J. Trop. Med. Public Health*, 28(Suppl 1): 190 – 193

- REICZIGEL, J., RÓZSA, L. (2005): *Quantitative Parasitology 3.0*. Budapest, Hungary
- RIBAS, A., SAIJUNTHA, W., AGATSUMA, T., PRANTLOVÁ, V., POONLAPHDECHA, S. (2016): Rodents as a source of *Salmonella* contamination in wet markets in Thailand." *Vector Borne Zoonotic Dis.* 16 (8): 537 – 540. doi:10.1089/vbz.2015.1894
- TESANA, S., PUAPAIROJ, A., SAESEOW, O. (2007): Granulomatous, hepatolithiasis and hepatomegaly caused by *Capillaria hepatica* infection: first case report of Thailand. *Southeast Asian J. Trop. Med. Public Health*, 38(4): 636 – 640
- TUNG, K.C., HSIAO, F.C., WANG, K.S., YANG, C.H., LAI, C.H. (2013): Study of the endoparasitic fauna of commensal rats and shrews caught in traditional wet markets in Taichung City, Taiwan. *J. Microbiol. Immunol. Infect.*, 46(2): 85 – 88. DOI:10.1016/j.jmii.2012.01.012
- VECIANA, M. (2016): *Noves dades parasitològiques en micro-mamífers (insectívors i rosegadors) del Sud-Est Asiàtic (Cambodja, Laos i Tailàndia)*. [New parasitological data on small mammals (insectivores and rodents) from South-east Asia (Cambodia, Laos and Thailand)]. PhD thesis, Spain, Barcelona: Universitat de Barcelona (In Catalan)
- WELLS, K., O'HARA, R.B., MORAND, S., LESSARD, J.P., RIBAS, A. (2015): The importance of parasite geography and spillover effects for global patterns of host-parasite associations in two invasive species. *Divers. Distrib.*, 21(4): 477 – 486. DOI:10.1111/ddi.12297
- WILSON, M. E., LORENTE, C. A., ALLEN, J. E., EBERHARD, M. L. (2001): *Gongylonema* infection of the mouth in a resident of Cambridge, Massachusetts. *Clin. Infect. Dis.*, 32(9): 1378 – 1380. DOI:10.1086/319991
- WONGSAROJ, T., NITHIKATHKUL, C., ROJKITIKUL, W., NAKAI, W., ROYAL, L., RAMMASUT, P. (2014): National survey of helminthiasis in Thailand. *Asian Biomed.*, 8(6): 779 – 783. DOI:10.5372/1905-7415.806.357
- YOUNG, P.L., BABERO, B.B. (1975): Studies on the transmission of helminth ova by cockroaches. *Proc Okla Acad Sci* 55: 169 – 174