

Impact of cyclin D1 and DJ-1 on diagnosis, clinico-pathological features and outcome in prostate cancer and benign prostatic hyperplasia

Research Article

Amrallah A. Mohammed*¹, Hanna M. Ibrahim², Hanna A. Atwa²,
Ayman elshentenawy³, Amira Elwan⁴

¹Medical Oncology Department, Faculty of Medicine, Zagazig University, Egypt

²Department of Pathology, Faculty of Medicine, Zagazig University

³Kasr EL Einy center of clinical oncology and nuclear (NEMROCK), Cairo university, Egypt

⁴Clinical oncology Department, Faculty of Medicine, Zagazig University, Egypt

Received 3 August 2018; Accepted 21 July 2019

Abstract: Background: Disturbance in cell cycle regulatory genes is a common finding among many types of cancers. The aim of this study is to evaluate the role of cyclin D1 and DJ-1 in benign prostatic hyperplasia (BPH) and prostate cancer (PC).
Method: The current study enclosed 40 patients diagnosed with PC and 40 cases of BPH. The expression level of cyclin D1 and DJ-1 were evaluated by immunohistochemistry (IHC). Cyclin D1 scored depending on the percentage of stained nuclear tumor cells. While scoring of DJ-1 was based on intensity. The results were correlated with clinicopathological features and outcome.
Results: In the PC group, cyclin D1 was detected in 95% and overexpressed in 42.5%, DJ-1 was positively stained in 85% and overexpressed in 47.5%. Meanwhile, in the BPH group, cyclin D1 was not detected and DJ-1 stained in only 2.5%. There was a statistically significant difference in Gleason score (GS), tumor stage, size, and treatment failure ($p = < 0.001$). In the terms of PC diagnosis prediction, although cyclin D1 was more specific (100%), DJ-1 is more sensitive than cyclin D1 (80%, 70%, respectively) ($p = 0.000$).
Conclusions: Cyclin D1 and DJ-1 may emerge as a promising way for diagnosis of PC in certain circumstances, as the presence of insufficient tissue sampling, small foci of carcinoma or benign lesions mimic PC. This is in addition to the known role of cyclin D1 and DJ-1 in PC prognosis.

Keywords: Prostate carcinoma • benign prostatic hyperplasia • cyclin D1 • DJ-1

1. Introduction

Prostate cancer (PC) represents **19%** of the newly diagnosed male cancer cases of 2017 and is the second cause of **cancer-related deaths**.^[1] PC is not one **disease**, the heterogeneity is reflected through the different clinical behavior. This mandated the need for biomarkers that help in therapy selection and proper diagnosis in difficult circumstances.

Dysregulation of cell cycle regulating genes is believed to play a major part in cancers. Cyclin D1 is the key regulator during the G1 phase and **overexpression** was associated with malignant transformation.^[2]

DJ-1, also known as Parkinson disease protein 7, is encoded by the PARK7 gene. It acts **as a** negative regulator of PTEN (Phosphatase and Tensin homologue) gene, leading to tumor proliferation and invasion.^[3]

Although DJ-1 has been found to be **overexpressed** in multiple cancers, the expression pattern of PC needs more clarifications.

Accurate diagnosis is necessary to ensure the best and effective management. However, in some circumstances, for example, small foci and minimal (< 1 mm) needle tissue biopsy, the diagnosis is challengeable, and the IHC may be helpful.^[4,5]

* E-mail: amrallaabdelmoneem@yahoo.com

Moreover, there are data suggesting the relation between overexpression of DJ-1, cyclin D1 and androgen receptor (AR) status.^[6,7]

The aim of this study is to evaluate the expression level of cyclin D1 and DJ-1 in BPH and PC, correlation with clinicopathological features and assess the sensitivity and specificity of both as immune-markers in discerning some embarrassing cases.

2. Materials and methods

The current retrospective study involved 89 prostate needle biopsy specimens that were suspicious of cancer on abnormal rectal examination and/or elevated PSA during the period from January 2010 to March 2015 from the archives of the Department of Pathology, Medical Oncology Department and Clinical Oncology Department, Faculty of Medicine, Zagazig University. 9 samples were excluded due to insufficient data. They were diagnosed histopathologically with 40 cases of PC and 40 cases of BPH. The clinic-pathological and demographic features including age, pathological features, TNM stage, serum prostatic specific antigen (PSA) level, Gleason score, and followed up period were collected from the files of patients. The immune-histochemical analysis was done using cyclin D1 and DJ -1 antibody.

3. Steps of preparation

Four-five-micron sections from the blocks were cut into positive-charged slides; air dried overnight, deparaffinized in xylene, hydrated through a series of graded alcohol and washed in distilled water and 0.01 PBS. The avidin–biotin–complex (ABC) method was used for the immunohistochemistry staining. Endogenous peroxidase activity was blocked with 3% hydrogen peroxide in methanol for 10 minutes. The sections were then treated with microwave radiation for 10 min for antigen retrieval, and, to block intrinsic antibody binding, they were reacted with normal serum (mouse IgG) for 10 min at room temperature. The sections were incubated overnight with a solution of primary antibodies to: Rabbit monoclonal anti-Cyclin D1 antibody (Cat. from Thermo Scientific/Lab Vision Corporation, Fermont, USA, and clone: EPR2764. 0.09% sodium azide. Dilution 1:100) and Rabbit monoclonal anti DJ1 antibody (Cat. from Thermo Scientific/Lab Vision Corporation, Fermont, USA, and clone: EPR2359. 0.09% sodium azide. Dilution 1:100) with appropriate negative and positive controls, they were reacted with biotinylated anti-

mouse antibody (secondary antibody) for 10 min and with ABC for another 10 min, with intervening washes. Diaminobenzidine tetrahydrochloride was used as the final chromogen, and sections were counterstained with Mayer's hematoxylin before mounting. Positive controls were cancer breast and kidney tissue for cyclin D1 and DJ1 respectively. Negative control was employed by substituting primary antibody. Cyclin D1 scored as: **negative**, **1+ (weak)** = less than 10%, **2+ (moderate)** = 11 to 50% and **3+ (strong)** = more than 50% nuclear tumor cells stained positive,^[8] while the scoring of DJ-1 was based on intensity. Four areas per tissue were evaluated using the following scale: **0**, no staining; **1**, faint staining; **2**, moderate intensity staining; and **3**, and intensity staining.^[9] Positive stain referred to the intensity or positive expression, while overexpression means increase in the percent of stained cells (the extent of staining).

4. Statistical analysis

Categorical variables were expressed as a number (percentage). Percent of categorical variables were compared using Pearson's Chi-square test or Fisher's exact test when appropriate. **Trends of change in the distribution of relative frequencies between ordinal data** were compared using the Chi-square test for trend. All tests were **two-sided**. We estimated the survival rates during the entire follow-up period by the Kaplan–Meier method. All the statistical analyses were performed using IBM SPSS 22.0 for Windows (SPSS Inc., Chicago, IL, USA) and Microsoft Office Excel 2010 for Windows (Microsoft Cor., Redmond, WA, USA), **with** a p-value < 0.05 indicating statistical significance.

5. Results

A total of 80 eligible patients were included, 40 cases of BPH and the other 40 cases had PC. In BPH group, the median age was 65.5 years (range, 53–65 years) and median serum PSA level was 11.6 ng/ml (range 3.1–20 ng/ml).

6. PC group

The median age was 65.5 years (range, 50–81 years) and median serum PSA level was 50.5 ng/ml (range, 10–91 ng/ml). Table 1 shows the main clinicopathological features and outcome of 40 patients with PC.

Table 1: Clinicopathological features and outcome of 40 patients with prostatic carcinoma.

Characteristics	Prostatic Carcinoma (N=40)		Characteristics	Prostatic Carcinoma (N=40)	
	No.	%		No.	%
Gleason score			Relapse		
< 7	7	17.5%	Absent	16	40%
= 7	11	27.5%	Present	24	60%
> 7	22	55%			
T			Distant metastasis		
T1	5	12.5%	Absent	16	40%
T2	14	35%	Present	24	60%
T3	21	52.5%			
Stage			Hormone Refractory Relapse (N=24)		
Stage I	3	7.5%	Absent	7	29.2%
Stage IIa	6	15%	Present	17	70.8%
Stage IIb	10	25%			
Stage III	21	25%			

Categorical variables were expressed as number (percentage).

7. IHC results

Regarding the staining, cyclin D1 and DJ-1 were positive in 38 patients (95%) and 34 patients (85%) in PC, respectively. While no nuclear staining was detected for cyclin D1, the DJ-1 was positive in only 1 sample (2.5%) in the BPH group (Figure 1).

If we use cyclin D1/DJ-1 extent, both positive (+/+) was detected in 34 patients (85%) in the PC group and it was not detected in PBH group (0%). The same finding when used in cyclin D1 intensity/DJ-1 intensity, was statistically significant ($p < 0.001$). Table 2 shows the comparison between PC and BPH as regard cyclin D1 and DJ1 (staining and overexpression). Moreover, there were statistically significant differences between cyclin D1 and DJ-1 intensity/score/overexpression and clinicopathological features in the terms of GS, tumor size, TNM staging ($p = 0.001$).

Relapse and hormone refractory were statistically significantly correlated with overexpression of cyclin D1 and DJ-1. Although this positive correlation was maintained in DJ-1 extent; it was lost in cyclin D1 extent ($p = 0.154$) (Table 3, 4) (Figure 2, 3).

8. The relationship between the overexpression/staining of cyclin D1 and DJ-1 and outcome

The clinicopathological parameters in the form of high GS, large tumor size and higher stage were statistically significantly associated with disease relapse ($p < 0.001$). However, this association was lost in the case of cyclin D1 extent ($p < 0.15$). Moreover, the same findings were

detected in the case of overexpression and intensity of both cyclin D1 and DJ-1 ($p < 0.001$) (Table 3).

In addition, there was no statistically significant association with GS, tumor size, stage and hormone refractory ($p = 1.000$). Meanwhile, those associations were regained with the overexpression and intensity of both cyclin D1 and DJ-1.

9. Performance of the markers for PC diagnosis

DJ-1 was more sensitive than cyclin D1 in predicting PC diagnosis (80%, 70%, respectively). However, cyclin D1 was more specific (100%) ($p = 0.000$) (Figure 4, 5).

10. Discussion

Both environmental and genetic factors may be implicated in increasing the diagnosis of PC. The clinical course ranges from indolent behavior to highly aggressive that ultimately causes significant morbidity and even death.^[10] To achieve the best treatment, we need proper histopathology diagnosis via an adequate tissue sample.

The cell cycle regulators have been involved in many types of cancer including PC and associated with tumor aggressiveness and poor prognosis.^[11]

Cyclin D1 is a nuclear protein that is involved in shortening the G1 (growth) –S (synthesis) transition. Its overexpression is considered an oncogene, as it leads to uncontrolled cell growth and transformed into a malignant phenotype. As we know that cyclin D1 can

Table 2. Comparison between PC and BPH as regard cyclin D1 and DJ-1 (staining and overexpression)

	Prostatic Carcinoma (N=40)		Prostatic Hyperplasia (N=40)		p-value‡
	No.	(%)	No.	(%)	
Cyclin D1 extent					
Negative	2	(5%)	39	(97.5%)	< 0.001
Positive	38	(95%)	1	(2.5%)	
DJ1 extent					
Negative	6	(15%)	39	(97.5%)	< 0.001
Positive	34	(85%)	1	(2.5%)	
Cyclin D1 extent /DJ1 extent					
Negative/Negative	2	(5%)	38	(95%)	< 0.001
Negative/Positive	0	(0%)	1	(2.5%)	
Positive/Negative	4	(10%)	1	(2.5%)	
Positive/Positive	34	(85%)	0	(0%)	
Cyclin D1 intensity					
0	2	(5%)	39	(97.5%)	< 0.001
1	5	(12.5%)	1	(2.5%)	
2	16	(40%)	0	(0%)	
3	17	(42.5%)	0	(0%)	
DJ1 intensity					
0	6	(15%)	39	(97.5%)	< 0.001
1	4	(10%)	0	(0%)	
2	14	(35%)	1	(2.5%)	
3	16	(40%)	0	(0%)	
Cyclin D1 intensity/DJ1 intensity					
0/0	2	(5%)	38	(95%)	< 0.001
0/1	0	(0%)	0	(0%)	
0/2	0	(0%)	1	(2.5%)	
1/0	1	(2.5%)	1	(2.5%)	
1/1	4	(10%)	0	(0%)	
1/2	0	(0%)	0	(0%)	
2/0	3	(7.5%)	0	(0%)	
2/1	0	(0%)	0	(0%)	
2/2	10	(25%)	0	(0%)	
2/3	3	(7.5%)	0	(0%)	
3/2	4	(10%)	0	(0%)	
3/3	13	(32.5%)	0	(0%)	
Cyclin D1 overexpression					
Negative	23	(57.5%)			
Positive	17	(42.5%)			
DJ-1 overexpression					
Negative	21	(52.5%)			
Positive	19	(47.5%)			

Categorical variables were expressed as number (percentage); ‡ Chi-square test; p< 0.05 is significant.

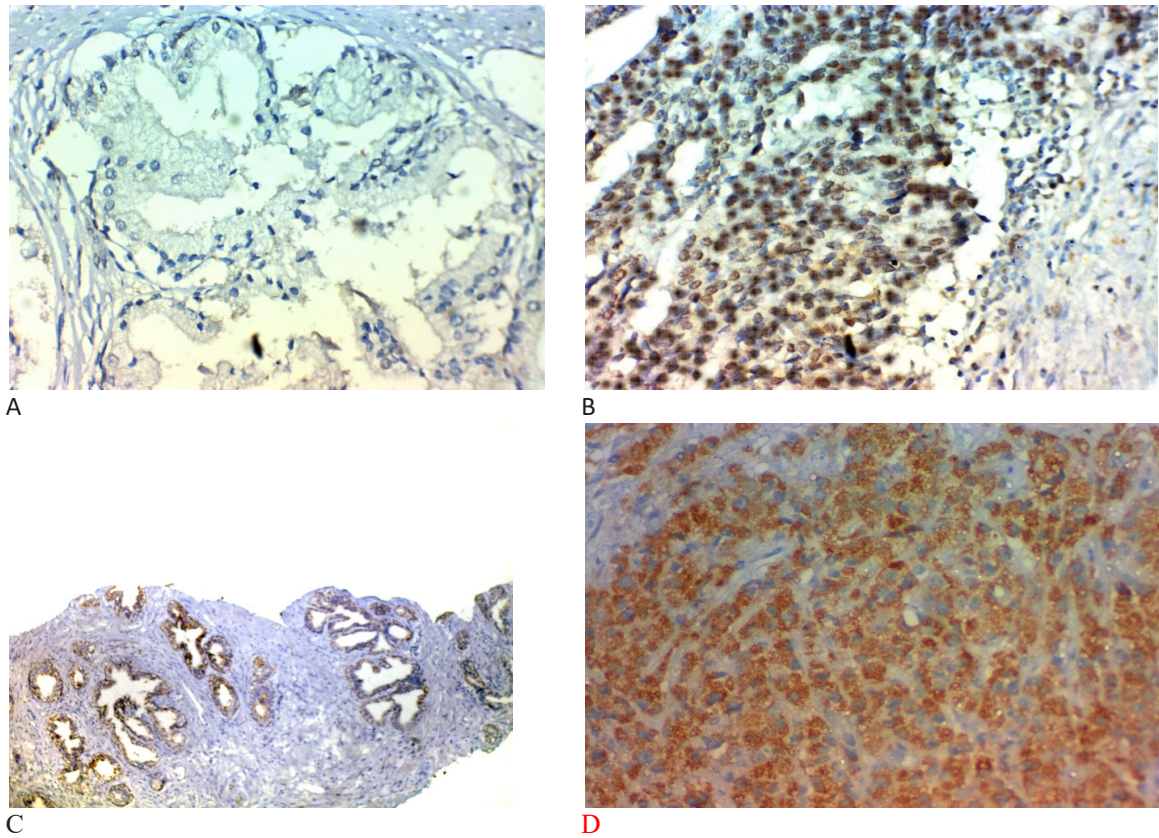


Figure 1. Staining of cyclin D1 and DJ-1 in benign prostatic hyperplasia and prostatic adenocarcinoma:
A) Benign prostatic hyperplasia showing negative cyclin D1 staining (ABC x 400)
B) High grade prostatic adenocarcinoma showing strong Cyclin D1 staining (ABC x 200)
C) Benign prostatic hyperplasia showing faint DJ1 intensity (ABC x 100)
D) High grade Prostatic adenocarcinoma with strong DJ1 intensity . (ABC x 400)

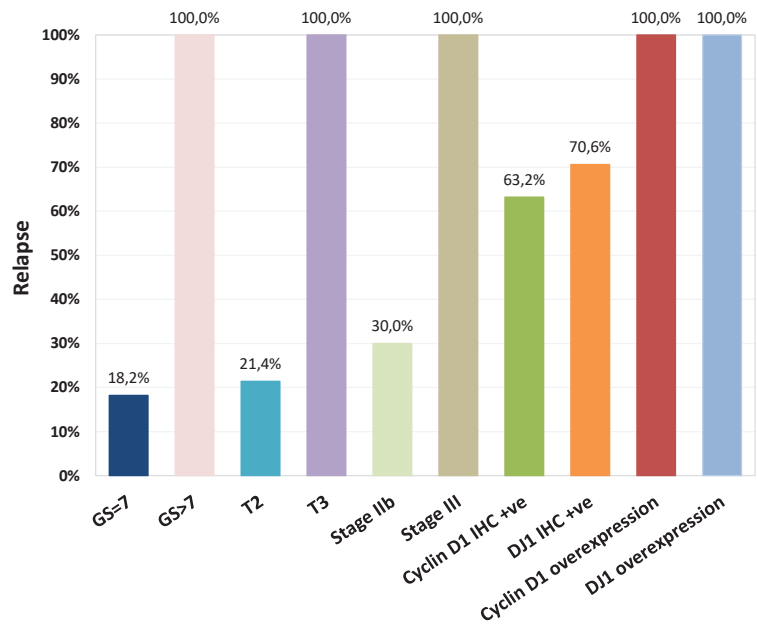


Figure 2. Bar chart shows percent of relapse among different studied subpopulation of prostatic carcinoma patients.

Table 3. Relation between cyclin D1 and DJ-1 (staining and overexpression), clinicopathological parameters and relapse in 40 PC patients

	Prostatic Carcinoma (N=40)		Relapse				p-value
			Absent (N=16)		Present (N=24)		
	No.	(%)	No.	(%)	No.	(%)	
Gleason score							
< 7	7	(17.5%)	7	(100%)	0	(0%)	< 0.001§
= 7	11	(27.5%)	9	(81.8%)	2	(18.2%)	
➤ 7	22	(55%)	0	(0%)	22	(100%)	
T							
T1	5	(12.5%)	5	(100%)	0	(0%)	< 0.001§
T2	14	(35%)	11	(78.6%)	3	(21.4%)	
T3	21	(52.5%)	0	(0%)	21	(100%)	
Stage							
Stage I	3	(7.5%)	3	(100%)	0	(0%)	< 0.001§
Stage IIa	6	(15%)	6	(100%)	0	(0%)	
Stage IIb	10	(25%)	7	(70%)	3	(30%)	
Stage III	21	(25%)	0	(0%)	21	(100%)	
Cyclin D1 extent							
Negative	2	(5%)	2	(100%)	0	(0%)	0.154‡
Positive	38	(95%)	14	(36.8%)	24	(63.2%)	
DJ1 extent							
Negative	6	(15%)	6	(100%)	0	(0%)	0.002‡
Positive	34	(85%)	10	(29.4%)	24	(70.6%)	
Cyclin D1 extent /DJ1 extent							
-ve/-ve	2	(5%)	2	(100%)	0	(0%)	0.003§
+ve/-ve	4	(10%)	4	(100%)	0	(0%)	
+ve/+ve	34	(85%)	10	(29.4%)	24	(70.6%)	
Cyclin D1 intensity							
0	2	(5%)	2	(100%)	0	(0%)	0.002§
1	5	(12.5%)	5	(100%)	0	(0%)	
2	16	(40%)	5	(31.3%)	11	(68.8%)	
3	17	(42.5%)	4	(23.5%)	13	(76.5%)	
DJ1 intensity							
0	6	(15%)	6	(100%)	0	(0%)	< 0.001§
1	4	(10%)	4	(100%)	0	(0%)	
2	14	(35%)	6	(42.9%)	8	(57.1%)	
3	16	(40%)	0	(0%)	16	(100%)	
Cyclin D1 intensity/DJ1 intensity							
0/0	2	(5%)	2	(100%)	0	(0%)	< 0.001§
1/0	1	(2.5%)	1	(100%)	0	(0%)	
1/1	4	(10%)	4	(100%)	0	(0%)	
2/0	3	(7.5%)	3	(100%)	0	(0%)	
2/2	10	(25%)	2	(20%)	8	(80%)	
2/3	3	(7.5%)	0	(0%)	3	(100%)	
3/2	4	(10%)	4	(100%)	0	(0%)	
3/3	13	(32.5%)	0	(0%)	13	(100%)	
Cyclin D1 overexpression							
Negative	23	(57.5%)	16	(69.6%)	7	(30.4%)	< 0.001‡
Positive	17	(42.5%)	0	(0%)	17	(100%)	
DJ1 overexpression							
Negative	21	(52.5%)	16	(76.2%)	5	(23.8%)	< 0.001‡
Positive	19	(47.5%)	0	(0%)	19	(100%)	

Categorical variables were expressed as number (percentage); ‡ Chi-square test; § Chi-square test for trend; p < 0.05 is significant.

Table 4. Relation between cyclin D1 and DJ-1 (staining and overexpression), clinicopathological parameters and hormone response in 40 PC patients

	Prostatic Carcinoma (N=24)		Hormone Refractory				p-value
			Absent (N=7)		Present (N=17)		
	No.	(%)	No.	(%)	No.	(%)	
Gleason score							
= 7	2	(8.3%)	0	(0%)	2	(100%)	1.000‡
➤ 7	22	(91.7%)	7	(31.8%)	15	(68.2%)	
T							
T2	3	(12.5%)	1	(33.3%)	2	(66.7%)	1.000‡
T3	21	(87.5%)	6	(28.6%)	15	(71.4%)	
Stage							
Stage IIb	3	(12.5%)	1	(33.3%)	2	(66.7%)	1.000‡
Stage III	21	(87.5%)	6	(28.6%)	15	(71.4%)	
Cyclin D1 extent							
Positive	24	(100%)	7	(29.2%)	17	(70.8%)	---
DJ1 extent							
Positive	24	(100%)	7	(29.2%)	17	(70.8%)	---
Cyclin D1 extent /DJ1 extent							
+ve/+ve	24	(100%)	7	(29.2%)	17	(70.8%)	---
Cyclin D1 intensity							
2	11	(45.8%)	0	(0%)	11	(100%)	0.006‡
3	13	(54.2%)	7	(53.8%)	6	(46.2%)	
DJ1 intensity							
2	8	(33.3%)	0	(0%)	8	(100%)	0.054‡
3	16	(66.7%)	7	(43.8%)	9	(56.3%)	
Cyclin D1 intensity/DJ1 intensity							
2/2	8	(33.3%)	0	(0%)	8	(100%)	0.006§
2/3	3	(12.5%)	0	(0%)	3	(100%)	
3/3	13	(54.2%)	7	(53.8%)	6	(46.2%)	
Cyclin D1 overexpression							
Negative	7	(29.2%)	7	(100%)	0	(0%)	<0.001‡
Positive	17	(70.8%)	0	(0%)	17	(100%)	
DJ1 overexpression							
Negative	5	(20.8%)	5	(100%)	0	(0%)	<0.001‡
Positive	19	(79.2%)	2	(10.5%)	17	(89.5%)	

Categorical variables were expressed as number (percentage); ‡ Chi-square test; p < 0.05 is significant.

control the mitogenic signaling, either the strength or duration. There is data available on the relationship between the low or undetectable level of cyclin D1 and the level of PSA. Thus, in PC, the potency of cyclin D1 to curb AR activity seems to be lost, reflecting the role of AR in tumor initiation and progression.^[12]

The relationships between PC and **overexpression** of cyclin D1 are challenging. Some studies have revealed that cyclin D1 **overexpression** in PC is rare, where others showed the association with aggressive disease behavior and **overexpression**.^[13–15]

In our study, cyclin D1 immunostaining was detected in 95% of PC group and was not detected in BPH group (0%), which is in agreement with a study done by Ueda et al.^[16] In another retrospective study on 100 prostatic specimens, which divided into 50 cases were BPH and 50 cases were PC, Qahtani et al. demonstrated that cyclin D1 **overexpression** was detected in 45 specimens out of 50 PC specimens (90%), while only 16% was focally positive in BPH group.^[17]

On the other hand, our results contradict some other studies that revealed cyclin D1 **overexpression** was

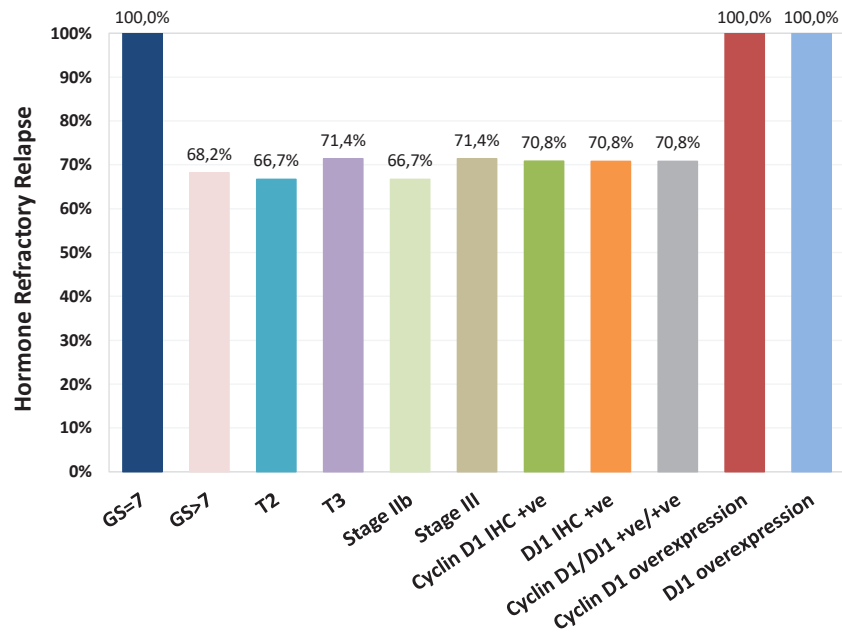


Figure 3. Bar chart shows percent of hormone refractory relapse among different studied subpopulation of prostatic carcinoma patients

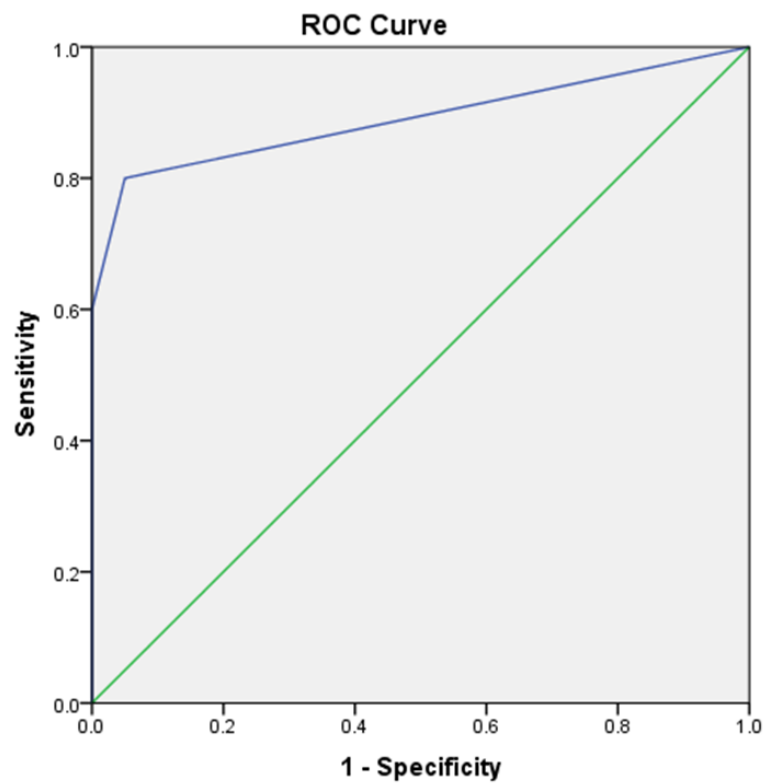


Figure 4. ROC curve for sensitivity and specificity of cyclin D1 in prediction of prostatic carcinoma

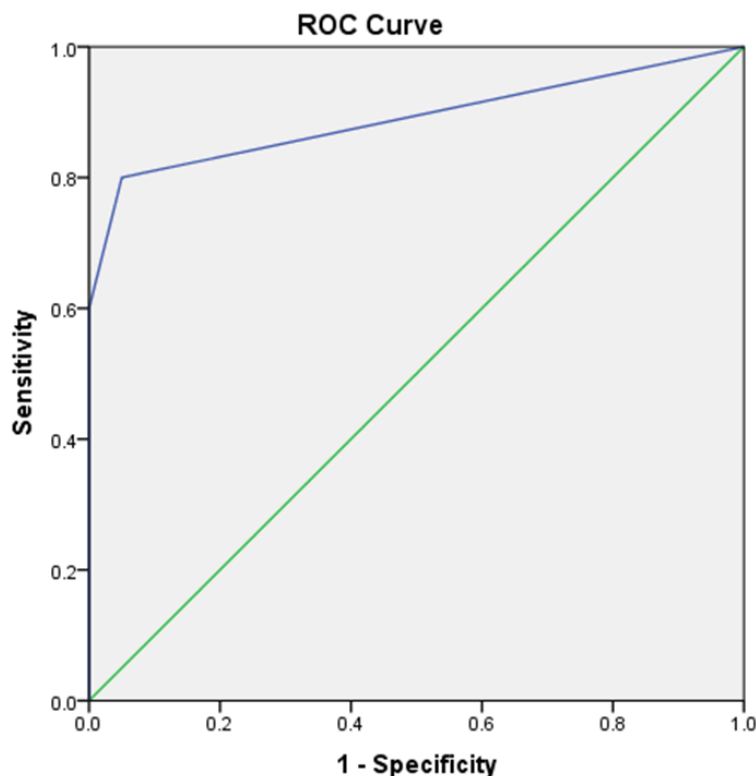


Figure 4. ROC curve for sensitivity and specificity of DJ-1 in prediction of prostatic carcinoma

detected in **the** range of 22% up to 11% of PC cases.
[18,19]

DJ-1 is a diverse signaling protein associated with multiple cellular processes, such as cellular transformation, response to oxidative stress, and androgen-receptor signaling.^[20] It acts as **a** negative regulator of PTEN gene, leading to tumor proliferation, invasion, **and** distance metastasis. It increased in response to increased ROS (reactive oxygen species) levels.^[21]

As regards DJ-1 immunostaining, our presenting results showed that 85% of PC was positive, compared to 2.5% was positive in BPH group; these **findings** are inconsistent with other earlier studies.^[22,23] The meta-analysis involved fourteen studies including 1,947 cancer patients and revealed that DJ-1 was an important biomarker in tumor evaluation and outcome.^[24]

The findings of our study provided that involvement in cyclin D1 and DJ-1, **and in** all the studied stages of PC compared to BPH. In addition, we found a significant association with the extent, the intensity, and **overexpression** of cyclin D1 and DJ-1 in PC, and BPH patients suggesting that they could be used as biomarkers for early detection of PC.

This is supported by **an** early study done by Lee et al., which showed that **the** mutation of many cell cycle regulating proteins was involved in the initiation of PC to late stage of disease progression.^[25] In addition, these results are close to the other previous studies.^[22,23]

In addition, we found a significant association with cyclin D1/DJ-1 overexpression with tumor size, stage, and GS > 7. Moreover, similar results validated the same association and were reported by many studies.^[17,26,27] On the other hand, this association was not proved in many earlier studies.^[9,14,16,24,28]

Sensible explanations were suggested to prove that these differences are sample size, technical varieties of biopsy taking, staining type (cytoplasmic vs nuclear) and scoring system.

Our data showed that **overexpression** of cyclin D1 and DJ-1 was associated with treatment failure in the form of relapse, hormonal refractory or distance metastasis. Moreover, the same results were obtained from a study done by Drobnjak et al., who reported that cyclin D1 **overexpression** was associated with bone metastasis in PC.^[13]

In the PC group, the sensitivity and specificity of cyclin D1 expression were 70% and 100%, respectively,

while in a retrospective study done by Atta et al., on 60 cases of PC were 93.3% and 86.6%, respectively. In addition, they were 80% and 95% respectively for DJ-1 expression, while by Osman et al., they were 93.33% and 86.67%, respectively.^[23]

Can we use these results of clinical practice to help in diagnosis in special cases? **Actually**, the answers need more verification and more investigations.

Based on **cross-talk** between antioxidant systems, targeting both DJ-1 and thioredoxin (antioxidant system found in all species) may be an effective new anticancer therapy.^[29] Recently, in 2016, Imrali et al. reported synergistic effect between rapamycin and cisplatin in the presence of high cyclin D1 level in PC.^[30]

From a practical point of view and owing to their **role in tumor** initiation and progression, targeting cyclin D1 and DJ-1 is considering a promising option on the era of molecular medicine.

11. Limitations

Besides the small sample size, the retrospective studies are always criticized due to incomplete data, probability of selection or information bias, and for depending totally on medical documentation. The data on risk factors such as smoking or cancer history either family or past history were not available almost in all medical files. Also, the survival analysis was difficult to be evaluated.

References

- [1] Siegel RL, Miller KD, Jemal A. CA Cancer J Clin. 2017; 67:7–30.
- [2] Casimiro MC, Crosariol M, Loro E, Li Z, Pestell RG; Cyclins and Cell Cycle Control in Cancer and Disease Genes Cancer. 2012; 3:649–57.
- [3] Zhu XL, Wang Z, Lei WB, Zhuang H, Hou W, Wen Y et al. Tumorigenesis role and clinical significance of DJ-1, a negative regulator of PTEN, in supraglottic squamous cell carcinoma. Journal of Experimental & Clinical Cancer Research 2012; 31:94.
- [4] Hameed O and Humphrey PA. Immunohistochemistry in the diagnosis of minimal prostate cancer Current Diagnostic Pathology 2006; 12: 279–291.
- [5] Omar H, Jack S and Peter H. Immunohistochemical Stains for p63 and α -Methylacyl-CoA Racemase, Versus a Cocktail Comprising Both, in the Diagnosis of Prostatic Carcinoma: A Comparison of the Immunohistochemical Staining of 430 Foci in Radical Prostatectomy and Needle Biopsy Tissues. American Journal of Surgical Pathology 2005; 29 (5): 579–587.
- [6] Atta IS, Eid AG, El-Hag MA, AlQahtani FF. Can cyclin D1 be utilized as a second step “after basal cell marker” for both diagnosis and prognosis of prostatic adenocarcinoma? International Journal of Medical Science and Public Health 2016; 5(09).
- [7] Gupta V, Garg M, Chaudhry M, Singh S, Sen R, Meenu Gill M et al. Role of cyclin D1 immunoreactivity and AgNOR staining in the evaluation of benign and malignant lesions of the prostate Prostate International 2014; 2; 21–30
- [8] Khoo ML, Beasley NJ, Ezzat S J, Freeman L and Asa SL. Overexpression of cyclin D1 and underexpression of p27 predict lymph node metastases in papillary thyroid carcinoma. J. Clin. Endocrinol Metab 2002; 87: 1814–1818.

12. Conclusions & Recommendations

Previous studies were focused mainly on the prognostic value of cyclin D1 and DJ-1 in PC. In our study, besides the prognostic value, there was statistically significant staining and overexpression of cyclin D1 and DJ-1 in PC in relation to BPH. Cyclin D1 had 70% sensitivity while DJ-1 had 80%, which made them markers for diagnosis in difficult cases such as limited tissue sample, small foci of carcinoma, or benign mimics of prostate cancer. We need to think out of **the** box to define nontraditional diagnostic and prognostic markers rather than the current parameters; PSA level, GS, and tumor stage for better precise care protocols.

13. Conflict of interest

The authors certify that there is no potential or actual conflict of interest related to this research.

- [9] Tillman JE, Yuan J, Gu G, Fazli L, Ghosh R, Flynt AS, et al. DJ-1 binds androgen receptor directly and mediates its activity in hormonally treated prostate cancer cells. *Cancer Res* 2007;67(10):4630–7.
- [10] Mohammed AA. Biomarkers in prostate cancer: new era and prospective. *Med Oncol*. 2014; 31:140.
- [11] Rubicz R, Zhao S, April C, Wright JL, Kolb S, Coleman I, et al. Expression of cell cycle regulated genes and prostate cancer prognosis in a population-based cohort. *Prostate*. 2015 Sep; 75:1354–62.
- [12] Knudsen K. E., Scher H. I. (2009) *Clin. Cancer Res*. 2009; 15, 4792–4798.
- [13] Drobnjak M, Osman I, Scher HI, Fazzari M, Cordon-Cardo C. Overexpression of cyclin D1 is associated with metastatic prostate cancer to bone. *Clin Cancer Res*. 2000; 6:1891–1895.
- [14] Fleischmann A, Rocha C, Saxer-Sekulic N, Zlobec I, Sauter G, Thalmann GN. High-level cytoplasmic cyclin D1 expression in lymph node metastases from prostate cancer independently predicts early biochemical failure and death in surgically treated patients. *Histopathology*. 2011; 58:781–789.
- [15] Kolar Z, Murray PG, Scott K, Harrison A, Vojtesek B, Dusek J. Relation of Bcl-2 expression to androgen receptor, p21WAF1/CIP1, and cyclin D1 status in prostate cancer. *Mol Pathol*. 2000; 53:15–18.
- [16] Ueda N, Yamashita M, Kuroda I, Takenaka I. Immunohistological evaluation of the expression of P27 and cyclin D1 in prostatic specimens. *Nishinohon J Urol* 2001; 63:246–9.
- [17] Qahtani FF, Atta IS, Mady EA. P63 and Cyclin D 1 Expression in Benign Prostatic Hyperplasia versus Prostatic Adenocarcinoma: A Clinicopathologic, Radiologic, and Immunohistochemical Study. *International Journal of Healthcare Sciences* 2015; 2: 320–305.
- [18] Drobnjak M, Osman I, Scher HI, Fazzari M, Cordon-Cardo C. Overexpression of cyclin D1 is associated with metastatic prostate cancer to bone. *Clin Cancer Res* 2000; 6:1891–1895.
- [19] Kallakury B.V., Sheehan C.E., Ambros R.A., Kaufman H.A and Ross JS. The prognostic significance of p34 and cyclin D1 protein expression in prostate adenocarcinoma. *Cancer* 1997; 80: 753-763.
- [20] Lev N, Roncevic D, Ickowicz D, Melamed E, Offen D. Role of DJ-1 in Parkinson's disease. *J Mol Neurosci*. 2006; 29:215-25. Review. Erratum in: *J Mol Neurosci*. 2007; 31:307.
- [21] Ismail IA, Kang HS, Lee HJ, Kim JK, Hong SH. DJ-1 upregulates breast cancer cell invasion by repressing KLF17 expression. *Br J Cancer*. 2014 Mar 4; 110:1298–306.
- [22] Lisitskaia KV, Eremina LS, Ivanov AV, Kovaleva MA, Okhrits V, Toropygin I, et al. Study of DJ-1 protein in tissuespecimens cultured cells and serum of prostate cancer patients. *Biomed Khim* 2011;57:392–401.
- [23] Osman W, Attia R and Abou Gabal H. DJ-1 and androgen receptor immunohistochemical expression in prostatic carcinoma: A possible role in carcinogenesis. *Egypt Natio Can Inst J* 2013; 25:223–230.
- [24] Wang Q, Li F, Shi W, Zhang Q, Wang J, Yan X, et al. Overexpression of DJ-1 correlates with aggressive clinicopathological characteristics and poor prognosis in malignant tumors: a meta-analysis. *Onco Targets Ther*. 2018; 11:3931–3942.
- [25] Lee JT, Lehmann BD, Terrian DM, Chappell WH, Stivala F, Libra M, et al. Targeting prostate cancer based on signal transduction and cell cycle pathways. *Cell Cycle*. 2008; 7:1745–1762.
- [26] Comstock CE, Revelo MR, Bunche CR, Knudsen KE. Impact of differential cyclin D1 expression and localisation in prostate cancer. *Br J Cancer*. 2007; 96: 970–979.
- [27] Ozbek E, Mizrak B, Ozbek M, Buyukberber S, Davarci M. Cyclin-D1 protooncogen expression in prostate cancer. *Turk J Cancer* 2000; 30: 15–21
- [28] Shiraishi T, Watanabe M, Muneyuki T, Nakayama, Morita J, Ito H, et al. clinicopathological study of p53, p21 (WAF1/CIP1) and cyclin D1 expression in human prostate cancers. *Urol Int* 1998; 61 p: 90–94
- [29] Prahlad V. Raninga, Trapani G, Kathryn F. Cross talk between two antioxidant systems, Thioredoxin and DJ-1. *Oncoscience* 2014; 1 (1).
- [30] Imrali A, Mao X, Yester-Velasco M. Shamash J, Lu Y. Rapamycin inhibits prostate cancer cell growth through cyclin D1 and enhances the cytotoxic efficacy of cisplatin. *Am J Cancer Res* 2016; 6:1772–1784.