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THE HONEY AS A BIOINDICATOR OF THE ENVIRONMENT

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Abstract: Monofloral rape, sunflower, linden and acacia honeys as well as honeys sampled within and around the Austrian towns Vienna and Linz, as well as in rural areas in Lower Austria, were analysed for main and trace elements. The results were tested to be used as indication of urban dust exposure. Main elements of the ash are K, P, and B, which are specific for plant origin. Ash based data correct for dilutions by the sugar matrix. They showed enrichments of Cu, Zn and Mo, whereas the contaminant elements Cd, Pb and Co were found at about the same levels as expectable in dust. Ash-based Al, Fe, Cr and Li were much lower than soil levels obtainable from aqua regia. Among the monofloral honey samples, the effect of adjacent soil was largest for Mn. Most differences between rural areas and urban areas in Vienna and Linz were within experimental errors, both per sample weight and per ash weight. In cases honey samples are considered to trace contaminations, the additional use of ash-based data is recommended.

Keywords: honey, bioindicator, trace elements, boron, ash-based data, environmental contamination

Introduction

Honey as a biomonitor

Modern life style in connection with an enormous increase in the use of vehicles, the massive increase in power generation and the industrialization of agriculture have increased dramatically the pollution of the environment. Biological monitoring has the advantage of measuring integrated exposure from all sources.

Within a bee hive, about 10,000 individuals are continuously working daily upon about 1,000 blossoms, growing at an area of usually more than 7 km² [1]. Contrary to the data from a single individual honeybee, effects of short-time variations in location and time get balanced by the bees' metabolism. Spatial differences will be masked by inherent differences in the elemental content of honey at any one location. Therefore it has been obvious to use honey as a bioindicator-substrate to estimate environmental pollution.

Modern multi-element determination methods enable to determine several elements of satisfactory recovery during the digestion procedure. Data from as many elements as

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possible permit the discrimination between randomly occurring elements and those, which are indicative for effects whatsoever

Composition of honey in general

Honey consists essentially of different sugars, predominantly fructose and glucose as well as other substances such as organic acids, enzymes and solid particles derived from honey collection. The colour of honey varies from nearly colourless to dark brown. The consistency can be fluid, viscous or partly to entirely crystallised. Flavour and aroma vary, and are due to the plant origin. Honey is a characteristically quite acidic item, its pH being between 3.2 and 4.5, which is low enough to be inhibitory to many animal pathogens. Gluconic acid was found to be the principal acid in honey. The acid-producing enzyme in honey is a glucose oxidase producing gluconic acid and hydrogen peroxide from glucose.

When the composition of various food items are compared, honey contains all elements at lowest levels, except C (from its sugar matrix), K and B [2].

Plant sources

When honey bees (*Apis mellifera* L.) feed mainly on nectar and pollen, they produce various kinds of floral honey, which can be characterized by flavours and pollen detected therein. But in forests, beneath various blossoms, honeybees collect also honeydew produced by aphids sucking the sap from the leaves of the trees. The inorganic matrix, which is termed as the ash, is received after combustion or volatilization of the sample. It can be calculated as the sum of the respective metal and non-metal oxides. Calculated ash and electrical conductivity are strongly correlated with the K-contents, which makes the main inorganic cation. Whereas K and most of the trace elements and thus the ash contents, reach higher levels in honeydew honeys, boron and Ca behave opposite (see e.g. Table 1) [2, 3].

Table 1
Calculated ash content and electrical conductivity in honey samples from East of Austria (A) and Greece (GR)
and correlation between ash and conductivity (from [3])

Honey	1	Ash [mg/kg]	Electr. co	Corr.	
	Median	Mean ±SD*	Median	Mean ±SD	
Floral (A)	1715	1653 ±423	478	489 ±118	0.8773
Honeydew (A)	4521	4752 ±928	1226	1461 ±498	0.7315
Floral (GR)	770	1016 ±705	261	303 ±176	0.9646
Honeydew (GR)	3091	3071 ±1363	746	783 ±302	0.9872

^{*} SD - Standard Deviation

Environmental impacts on honey composition

The composition of honey depends on green plant origin, the bee race, soil composition, distance to the sea, inputs from beekeeping and honey processing, as well as environmental contaminations by dust pollution covering the entire area (industries, cities; but not streets and railways) [2-5]. Many papers do not give any indication of these parameters, however.

The choice of the nutritional substrate is determined by availability and genetics of the bee race. Beneath plant selection, input of dust from the bees' hair and the metabolism in the honey-vesicle have to be considered. Less collection efficiency might lead to less

dilution of adhering dust by sugars. In Europe, bee-keeping is restricted to *apis mellifera* only and all data from this work and from references cited, refer to honey from these [5].

As a highly viscous liquid, honey is a good adhesive for dust particles. Dust particles might emanate from the abrasion of rocks, buildings and roads, from combustion processes, from sea-salt aerosol and from organic debris [3]. The local influences of highways and roads seems marginal due to the sampling strategy of the bees [1] (see also below).

In order to exclude variations due to plant composition and environmental impacts, effects of soil composition get only revealed, when monofloral honeys from different obviously non-contaminated locations are compared. Rape was selected because it is easy to find fields of it at different sites. Among monofloral rape honey samples, manganese (r = 1.00), but also nickel, iron and molybdenum correlated with the respective aqua regia soluble contents in the soil, the most significant slope was observed for manganese. As manganese can be measured rather easily, it could be considered as indicative for the site of the origin of honey. The levels of most other elements were presumably subject to the metabolism and transport pathways of the rape plant, including the non-metabolically driven contents of lithium and strontium [4].

Composition of commercially available honeys and historic samples

Usually, data from about 1850 are taken as the zero for industrial contamination and metal inputs to the environment have grown after this time. For honey, however, this is not the case. Frequently, data about concentrations met in honey found in older papers are much higher than from recent ones, due to laboratory blanks and determinations close to the detection limits. It can be assumed, however, that in former times there was some input of dirt from centrifugation and further processing of the honey samples, as well as dust from unpaved roads and coal burning, which could be the case particularly for 3rd World countries still today.

Within a government report [6] (detailed data have not been published yet), much lower levels of any ion in 122 commercial honey samples from Europe in 2009 (but samples from China, Tunisia and Australia were within this range) were found, than in 28 historic samples from 1927, found in the Tyrol and analysed by the same analytical methods, however [7]. Medians and ranges are given in Table 2.

Table 2 Median and ranges of commercially available honeys (2009) and historic samples

Elements in honeys	Honeys (2009)	Honeys (1927)
[mg/kg]	(122 samples) [6]	(28 samples) [7]
K	746 // 178-2930	1709 // 396-3966
Na	11.3 // 4.1-43	109 // 61-559
Fe	0.60 // 0.14-3.07	2.47 // 0.99-17.5
Zn	0.57 // 0.19-2.80	13.0 // 2.0-49.7
Mn	0.66 // 0.09-6.11	1.44 // 0.22-4.43
Cu	0.144 // 0.037-0.833	0.34 // 0.04-1.34
Ni	0.030 // < 0.006-0.296	0.150 // 0.002-0.77
Pb	0.011 // 0.003-0.040	0.148 // 0.048-0.42
Cr	0.0060 //0.0018-0.0351	0.0096 // <0.0003-0.0326
Co	0.0045 // 0.0002-0.0465	0.015 // 0.0055-0.056
Mo	0.0020 // 0.0001-0.0092	0.0067 // 0.0009-0.0114
Cd	0.0011// 0.0002- 0.011	0.0041// 0.0011-0.0116

References about honey as an environmental indicator

In the North of England, emissions of smelters around Liverpool and Birmingham markedly increased Ag, Cd and Pb in honeys sampled within the entire area, whereas proximity to roads had no effect, in spite of intense use of alkylated Pb-compounds in the fuel at this time. Similarly, no significant correlations between the concentrations met in honey and soils adjacent to the bee hives, were found [8].

In Poland, effects of a copper smelter were clearly visible in As, Cd, Zn and Pb honey contents, whereas seasonal effects were marginal (except for Zn) [9]. But trace elements in honeys from rural areas of Poland were close to samples from Sweden and Finland [10].

In the city of Rome, honeybees and, to a lesser extent, some of their products (pollen, propolis, wax), but not honey, were considered as representative bioindicators of environmental pollution for Cd, Cr and Pb [11].

Other authors have investigated honey samples from Spain [12], Portugal [13], Chile [14], Egypt [15], or Turkey [16].

Material and methods

Most of the honey samples were obtained directly from beekeepers. Samples obtained at the roof of the biocenter university building Vienna, at the University of Natural Resources and Life Sciences Vienna, as well as the monofloral honeys were obtained from the working group of bee-research, established at the University of Natural Resources and Life Sciences Vienna.

The monofloral honeys were isolated directly from the honeycombs, which were cut with a plastic knife and the covers removed with a plastic comb. These samples were placed in plastic beakers and put into a drying oven (40°C) in order to melt and to collect the outflowing honey from wax and pollen [4].

For the determination of total element contents, 4 g of honey samples were weighed into 50 cm³ Erlenmeyer flasks (precleaned with hydrochloric acid), 30 cm³ of suprapure nitric acid (65%) were added and gently heated in a special fume cupboard equipped with alkaline washing of exhaust gases to absorb the nitrous oxides evolved and finally temperature was gradually increased. When the residue had come to almost dryness at 140°C, 25 cm³ of ultrapure water were added, mixed and filtered from wax into disposable plastic cups. Contrary to green plants containing cellulose, or feed and food samples containing substantial amounts of fat, addition of perchloric acid or hydrogen peroxide was not necessary. Each batch was accompanied by 2 blanks [2]. Water purified by reverse osmosis had to be used exclusively. Checks revealed that the wax contained even less minerals than the honey.

A simultaneously operating optical ICP (Perkin Elmer Optima 3000 XL) was used as a multi-element analytical tool, due to low analyte levels the sample solutions were used undiluted. K was determined by flame emission on a Perkin Elmer 3030 AAS after appropriate dilution (20-100 fold). Pb, Cd, Cr and Mo were analysed by graphite furnace AAS on a Perkin Elmer 3030 Z, equipped with Zeeman background correction.

The recovery of all elements investigated including boron was > 90%, except for sulphur (recovery of S from methionine was just 40-64%). Sulphur data given below are thus just minimum values; no sulphur data were found in the references, however. Barium traces seem to interact with the glass and respective data were not reliable.

Detection limits were calculated from the standard deviations of the average of differences of the two blanks analysed within each batch. Considering 4 g sample digested and made up to 25 cm³, in the graphite furnace detection limits of 0.0001 mg/kg for Cd, 0.004 mg/kg for Pb, 0.001 mg/kg for Mo and 0.0015 mg/kg for Cr had been achieved, which permits reliable data in most cases, whereas many data for Co and V were smaller than the detection limits achieved at the ICP-OES (0.006 mg/kg for Co and 0.007 mg/kg for V, resp.). In case an appreciable part of data is below detection limits, in the subsequent data tables, data ranges are given instead of mean and standard deviation.

Results and discussion

The composition of honey depends on the plant origin (floral honey - honeydew honey), soil composition, bee race, and possible contaminations from dust immissions and honey processing [2]. Within Europe, no other bees except *apis mellifera* have been admitted. Processing in the lab was done as clean as possible (see above). In spite of different climatic conditions and plant cover ([3], Table 2), similar concentration levels and differences between floral honey and honeydew honey appeared between samples from Austria and Greece. Correlations with soil composition obtained by aqua regia, however, are less than expected [4]. With respect to more recent data from Turkey [16], the data presented in this work tend to be at the same level in Cu, Mn, Cr and Pb, somewhat lower in Cd, Zn and Fe, but slightly higher in Al and Ni.

Table 3 Monofloral honeys and soil composition (aqua regia) per sample weight (after [4])

Element	Wieselburg		Wr. Neustadt		Hollabrunn	
[mg/kg]	rape	soil	rape	soil	rape	soil
	N = 23		N = 25		N = 19	
Al	0.458 ± 0.191	17887	0.486 ± 0.115	23225	0.297 ±0.089	20859
В	11.71 ±1.82		7.78 ± 1.51		8.07 ±1.07	
Ca	50.01 ±4.50	7732	38.45 ±6.38	72774	47.15 ±5.46	76466
Cd	0.00026 ±0.00015	0.25	0.00019 ±0.00014	0.43	0.00027 ±0.00011	0.18
Co	< 0.006-0.014	11.38	< 0.006-0.015	10.26	0.018 ± 0.009	9.46
Cr	0.00316 ± 0.0023	31.6	0.0056 ±0.0051	29.3	0.0021 ±0.0019	28.6
Cu	0.096 ± 0.035	11.8	0.266 ± 0.066	30.6	0.096 ± 0.024	13.4
Fe	0.65 ±0.17	23340	0.34 ±0.08	25806	0.53 ±0.09	23953
K	322 ±69	3080	492 ±168	6000	218 ±41	3219
Li	0.0013 ±0.0014	17.9	0.0016 ±0.0007	31.0	0.0006 ±0.0007	24.6
Mg	17.30 ±2.47	4797	12.41 ±2.33	20866	12.40 ±1.20	12926
Mn	0.549 ± 0.134	1400	0.252 ± 0.078	691	0.132 ± 0.022	435
Mo	0.0041 ±0.0013	0.705	0.0040 ± 0.0014	0.582	0.0040 ±0.0019	0.417
Na	6.66 ± 1.23	112.4	4.59 ±1.43	191.7	3.08 ±0.69	150.7
Ni	0.057 ±0.019	19.4	0.064 ± 0.020	19.9	0.104 ±0.043	21.1
P	37.9 ± 7.7	929	31.4 ± 5.2	1198	38.9 ±6.1	586
Pb	0.0056 ± 0.0035	18.2	0.0057 ±0.0032	56.7	< 0.004	13.0
S	13.01 ±2.46		11.06 ±2.00		10.60 ±1.26	
Sr	0.032 ± 0.009	19.8	0.030 ± 0.010	64.6	0.031 ±0.008	91.9
V	< 0.007-0.025	36.8	< 0.007	34.6	-	33.3
Zn	0.355 ± 0.132	87.3	0.341 ±0.089	118.0	0.879 ± 0.258	81.1

Element	Hollabrunn		Hollabrunn		Hirschstetten	
[mg/kg]	sunflower	soil	acacia	soil	linden	soil
	N = 25		N = 24		N = 20	
Al	0.171 ±0.101	25071	0.083 ± 0.050	11961	0.467 ±0.129	17926
В	8.84 ±1.00		2.646 ±0.608		11.06 ±2.30	5.8
Ca	74.49 ± 7.52	81007	6.86 ± 2.94	47125	62.68 ± 7.08	78537
Cd	0.00035 ±0.00012	0.15	0.00008 ± 0.00004	0.18	0.00048 ±0.00007	0.127
Co	< 0.006-0.013	10.14		5.68	< 0.006-0.014	9.97
Cr	0.0038 ±0.0019	33.6	0.0054 ± 0.0041	17.8	0.0036±0.0014	37.4
Cu	0.222 ±0.031	35.4	0.077 ± 0.034	10.4	0.264 ±0.049	23.8
Fe	0.15 ±0.04	24934	0.17 ± 0.07	14394	1.40 ±0.19	17682
K	353 ±56	3944	174 ±8	1456	678 ±25	
Li	0.0005 ±0.0003	25.6	0.0004 ±0.0003	10.7	0.0128 ±0.0021	22.0
Mg	16.45 ±2.12	7517	2.43 ±0.56	7683	22.93 ±4.81	23324
Mn	0.103 ±0.010	484	0.023 ± 0.003	320	0.285 ±0.032	486
Mo	0.0039 ±0.0026	0.364	0.0026 ± 0.0016	0.283	0.0038 ±0.0013	0.02
Na	2.30 ±1.45	137.5	1.96 ±0.57	71.4	17.99 ±7.51	148
Ni	0.047 ±0.022	25.8	0.040 ± 0.016	13.9	0.032 ±0.031	31.5
P	36.2 ±3.4	606	21.6 ± 1.5	308	69.6 ±10.8	887
Pb	0.0178 ±0.0106	11.9	0.0049 ± 0.0058	13.8	0.0327 ± 0.0102	18.7
\mathbf{S}^*	7.96 ±0.95		2.84 ± 0.56		20.37 ±4.45	350
Sr	0.025 ±0.004	64.3	0.007 ± 0.003	34.2	0.111 ±0.012	140
V	< 0.007	38.9	< 0.007	19.3	< 0.007	34.2
Zn	1.254 ±0.419	72.3	0.647 ±0.310	59.1	1.734 ±0.457	71.0

^{*} incomplete recovery of methionine; N - number of samples

If element contents is taken per sample weight, data presented in Tables 3, 5 and 7 show very low levels of inorganic matrix elements, except for K and B. Indeed, levels met in honey are usually lowest among various food items, maybe except for sugar (not investigated). High boron is probably also a tool to detect forgeries of honey by coloured sugar solutions. Antibacterial activities of honey has been known for more than a century (first report in 1892, reviewed in [17]). Beneath osmotic effects because of low water activity, acidic pH and enzymes like glucose oxidase, high boron contents might be a reason of antibacterial activities also.

Ash-based data

Determinations of all main elements permit to calculate a presumable ash contents from the sum of the elements after transformation to the respective oxides (as it is usual in geology). Tables 4, 6 and 8 contain respective data referring to ash. These ash-based data correct for dilution by various sugars and else organics.

Beneath pollen composition for floral honeys, and transported loads in the phloem sap in case of honeydew honeys, some input via dust immission might occur. Dust immission measurements were not available on site, but it turned out that among the ash-based data, Cd, Pb, Mn and Sr were found to range within the levels met in arable soils, whereas Cu, Mo, Ni, Zn and above all B were enriched, and on the other hand, Al, Fe, Cr, Li and V were depleted (Tables 4, 6, 8), presumably due to green plant physiology.

For some of the samples, pollen analyses and electrical conductivity of 10% sample solutions were available. Ash contents, calculated as the sum of the respective metal and non-metal oxides, as well as electrical conductivity, are clearly different for floral and honeydew samples. Because K is undoubtedly the main inorganic component of honey,

Soil

some smaller labs may limit their investigations to determine K and electrical conductivity. A look at Tables 4, 6 and 8 shows that calculations of total ash from K only are not reliable, however.

Ash-based data might reflect soil influences even more (Table 4), particularly if the elements get sorted in the order of descending concentration per ash.

Soil

Wieselburg

Substance

 $\label{thm:monofloral} Table~4~\underline{Mo} no floral~honeys~and~soil~composition~(aqua~regia)~per~ash~weight~(B~and~S~in~soils~from~KClO_3-digests)$

Wr. Neustadt

Soil

Hollabrunn

[%]	rape		rape		rape	
	N = 23		N = 25		N = 19	
K ₂ O	58.85 ±4.39	0.37	72.98 ±5.48	0.72	52.54 ±4.61	0.39
P ₂ O ₅	13.18 ±2.31	0.21	9.36 ±1.97	0.27	17.94± 1.92	0.13
CaO	10.75 ±1.50	1.08	7.06 ±1.71	10.19	13.49± 2.51	10.71
MgO	4.38 ±0.40	0.80	2.65 ±0.48	3.46	4.18 ±0.57	2.14
SO ₃ *	4.92 ±0.68	-	3.61 ±0.51	-	5.37 ±0.69	
Na ₂ O	1.39 ±0.30	0.015	0.77 ±0.16	0.026	0.85 ±0.17	0.020
B_2O_3	5.74 ±1.51	-	3.35 ±1.05	-	5.37 ±0.88	
[mg/kg]						
Al	674 ±270	17887	613 ±155	23225	607 ±172	20859
Cd	0.406 ±0.213	0.25	0.264 ±0.222	0.43	0.560 ±0.246	0.18
Со	< 6-21	11.38	8-20	10.26	36.3 ±16.6	9.46
Cr	4.28 ±4.00	31.6	7.86 ±6.78	29.3	4.43 ±4.19	28.6
Cu	150 ±51	11.8	338 ±60	30.6	196 ±45	13.4
Fe	962 ±198	23340	452 ±137	25806	1082 ±183	23953
Li	1.9 ±1.9	17.9	2.1 ±1.0	31.0	1.2 ±1.2	24.6
Mn	834 ±75	1400	318 ±81	691	270 ±61	435
Mo	6.04 ±1.85	0.705	5.49 ±2.07	0.582	8.15 ±3.84	0.417
Ni	86 ±27	19.4	80 ±39	19.9	206 ±74	21.1
Pb	9.1 ±5.1	18.2	8.2 ±5.3	56.7	< 10	13.0
Sr	47 ±12	19.8	39 ±12	64.6	64 ±20	91.9
V	< 3-43	36.8	< 6	34.6	_	33.3
•					_	
Zn	545 ±182	87.3	442 ±75	118.0	1799 ±537	81.1
Zn Substance	545 ±182 Hollabrunn				1799 ±537 Hirschstetten	
Zn	545 ±182	87.3	442 ±75	118.0	1	81.1
Zn Substance	545 ±182 Hollabrunn	87.3	442 ±75 Hollabrunn	118.0	Hirschstetten linden N = 20	81.1
Zn Substance	545 ±182 Hollabrunn sunflower	87.3	442 ±75 Hollabrunn acacia	118.0	Hirschstetten linden $N = 20$ 69.44 ± 1.59	81.1
Zn Substance [%]	545 ±182 Hollabrunn sunflower N = 25 63.29 ±6.04 12.16 ±1.10	87.3 Soil 0.48 0.14	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35	0.18 0.07	Hirschstetten linden N = 20 69.44 ±1.59 12.27 ±0.88	81.1 Soil 1.90 0.17
Zn Substance [%] K ₂ O	545 ±182 Hollabrunn sunflower N = 25 63.29 ±6.04	87.3 Soil 0.48 0.14 11.34	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64	118.0 Soil	Hirschstetten linden N = 20 69.44 ±1.59 12.27 ±0.88 7.29 ±0.29	81.1 Soil
Substance [%] K ₂ O P ₂ O ₅ CaO MgO	545 ±182 Hollabrunn sunflower N = 25 63.29 ±6.04 12.16 ±1.10 15.87 ±3.36 3.95 ±0.36	87.3 Soil 0.48 0.14	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30	0.18 0.07	Hirschstetten linden N = 20 69.44 ±1.59 12.27 ±0.88 7.29 ±0.29 2.79 ±0.12	81.1 Soil 1.90 0.17
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ *	545 ±182 Hollabrunn sunflower N = 25 63.29 ±6.04 12.16 ±1.10 15.87 ±3.36	87.3 Soil 0.48 0.14 11.34	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48	0.18 0.07 6.60	Hirschstetten linden N = 20 69.44 ±1.59 12.27 ±0.88 7.29 ±0.29	81.1 Soil 1.90 0.17 11.54
Zn Substance [%]	545 ± 182 Hollabrunn sunflower $N = 25$ 63.29 ± 6.04 12.16 ± 1.10 15.87 ± 3.36 3.95 ± 0.36 2.92 ± 0.46 0.47 ± 0.32	87.3 Soil 0.48 0.14 11.34	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31	0.18 0.07 6.60	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23	1.90 0.17 11.54 4.43 0.088 0.78
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ *	545 ± 182 Hollabrunn sunflower $N = 25$ 63.29 ± 6.04 12.16 ± 1.10 15.87 ± 3.36 3.95 ± 0.36 2.92 ± 0.46	87.3 Soil 0.48 0.14 11.34 1.252	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46	0.18 0.07 6.60 1.27	Hirschstetten linden N = 20 69.44 ±1.59 12.27 ±0.88 7.29 ±0.29 2.79 ±0.12 3.70 ±0.19	1.90 0.17 11.54 4.43 0.088
Zn Substance [%]	545 ± 182 Hollabrunn sunflower $N = 25$ 63.29 ± 6.04 12.16 ± 1.10 15.87 ± 3.36 3.95 ± 0.36 2.92 ± 0.46 0.47 ± 0.32	87.3 Soil 0.48 0.14 11.34 1.252	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31	0.18 0.07 6.60 1.27	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23	1.90 0.17 11.54 4.43 0.088 0.78
Zn Substance [%]		87.3 Soil 0.48 0.14 11.34 1.252 0.019	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67	0.18 0.07 6.60 1.27 0.010	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ Na ₂ O B ₂ O ₃ [mg/kg] Al Cd		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67	0.18 0.07 6.60 1.27 0.010	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ Na ₂ O B ₂ O ₃ [mg/kg] Al Cd Co		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15 10.14	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67 288 ±175 0.274 ±0.126	0.18 0.07 6.60 1.27 0.010 11961 0.18 5.68	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062 $< 5-12$	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127 9.97
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ Na ₂ O B ₂ O ₃ [mg/kg] Al Cd Co Cr		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15 10.14 33.6	$\begin{array}{c} 442 \pm 75 \\ \hline \textbf{Hollabrunn} \\ \textbf{acacia} \\ N = \textbf{24} \\ 71.91 \pm 1.64 \\ 17.05 \pm 1.35 \\ 3.40 \pm 1.48 \\ 1.36 \pm 0.30 \\ 2.40 \pm 0.46 \\ 0.94 \pm 0.31 \\ 2.89 \pm 0.67 \\ \hline \\ 288 \pm 175 \\ 0.274 \pm 0.126 \\ \hline \\ 21.2 \pm 13.3 \\ \hline \end{array}$	0.18 0.07 6.60 1.27 0.010 11961 0.18 5.68 17.8	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062 $< 5-12$ 2.95 ± 1.20	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127 9.97 37.4
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ Na ₂ O B ₂ O ₃ [mg/kg] Al Cd Co Cr Cu		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15 10.14 33.6 35.4	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67 288 ±175 0.274 ±0.126 21.2 ±13.3 258 ±120	0.18 0.07 6.60 1.27 0.010 11961 0.18 5.68 17.8 10.4	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062 $< 5-12$ 2.95 ± 1.20 216 ± 32	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127 9.97 37.4 23.8
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ Na ₂ O B ₂ O ₃ [mg/kg] Al Cd Co Cr Cu Fe		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15 10.14 33.6 35.4 24934	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67 288 ±175 0.274 ±0.126 21.2 ±13.3 258 ±120 577 ±237	0.18 0.07 6.60 1.27 0.010 11961 0.18 5.68 17.8 10.4 14394	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062 $< 5-12$ 2.95 ± 1.20 216 ± 32 1144 ± 124	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127 9.97 37.4 23.8 17682
Zn Substance [%]		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15 10.14 33.6 35.4 24934 25.6	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67 288 ±175 0.274 ±0.126 21.2 ±13.3 258 ±120 577 ±237 1.5 ±1.1	0.18 0.07 6.60 1.27 0.010 11961 0.18 5.68 17.8 10.4 14394 10.7	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062 $< 5-12$ 2.95 ± 1.20 216 ± 32 1144 ± 124 10.5 ± 1.7	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127 9.97 37.4 23.8 17682 22.0
Zn Substance [%] K ₂ O P ₂ O ₅ CaO MgO SO ₃ * Na ₂ O B ₂ O ₃ [mg/kg] Al Cd Co Cr Cu Fe Li Mn		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15 10.14 33.6 35.4 24934 25.6 484	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67 288 ±175 0.274 ±0.126 21.2 ±13.3 258 ±120 577 ±237 1.5 ±1.1 78 ±9	0.18 0.07 6.60 1.27 0.010 11961 0.18 5.68 17.8 10.4 14394 10.7 320	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062 $< 5-12$ 2.95 ± 1.20 216 ± 32 1144 ± 124 10.5 ± 1.7 234 ± 19	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127 9.97 37.4 23.8 17682 22.0 486
Zn Substance [%]		87.3 Soil 0.48 0.14 11.34 1.252 0.019 25071 0.15 10.14 33.6 35.4 24934 25.6	442 ±75 Hollabrunn acacia N = 24 71.91 ±1.64 17.05 ±1.35 3.40 ±1.48 1.36 ±0.30 2.40 ±0.46 0.94 ±0.31 2.89 ±0.67 288 ±175 0.274 ±0.126 21.2 ±13.3 258 ±120 577 ±237 1.5 ±1.1	0.18 0.07 6.60 1.27 0.010 11961 0.18 5.68 17.8 10.4 14394 10.7	Hirschstetten linden $N = 20$ 69.44 ± 1.59 12.27 ± 0.88 7.29 ± 0.29 2.79 ± 0.12 3.70 ± 0.19 1.51 ± 0.23 2.71 ± 0.14 381 ± 97 0.392 ± 0.062 $< 5-12$ 2.95 ± 1.20 216 ± 32 1144 ± 124 10.5 ± 1.7	81.1 Soil 1.90 0.17 11.54 4.43 0.088 0.78 0.0015 17926 0.127 9.97 37.4 23.8 17682 22.0

Substance	Hollabrunn	Soil	Hollabrunn	Soil	Hirschstetten	Soil
[mg/kg]	sunflower		acacia		linden	
	N = 25		N = 24		N = 20	
Pb	28.6 ±16.9	11.9	26.4 ±19.6	13.8	27.0 ±8.5	18.7
Sr	38 ±9	64.3	24 ±10	34.2	91 ±8	140
V	< 3	38.9	< 3	19.3	< 3	34.2
Zn	1853 ±727	72.3	2081 ±1111	59.1	1417 ±335	71.0

^{*} incomplete recovery of methionine; N - number of samples

Monofloral honeys

Analysis of monofloral honeys from presumably non-contaminated sites (Table 3) shows that both the kind of plant and the soil composition influence honey composition. Monofloral honeys were collected in fields of rapeseeds and sunflowers and compared with aqua regia data from soils sampled at the site of the bee hive (Table 3; [4]). Monofloral honey of the similar plant origin differ at various sites, and vice versa. From this, estimations of the composition of non-contaminated honey from pollen analysis resp. plant origin will fail.

Monofloral honeys are not easily to obtain, and data are hardly found in the literature. The beekeeper has to move the hives to the field or the orchard just in time, collect the honey immediately after flowering and move the hives elsewhere. Because flowering lasts only a few weeks, bees have to get their nutrition from other sites during the rest of the season. Collection of monofloral honey was only possible to be done by colleagues from the beekeeping department in our house.

The selected sites were all situated in Lower Austria and differed in soil type and climate, but fertilization rate was about equal. At Hollabrunn, we meet a calcic chernozem on loess (pH 7.4), at Holzing (near Wieselburg) a haplic gleysol (pH 6.8) and at Wiener Neustadt a gleyic phaeozem (pH 7.5). Whereas the original data from rape honey and sunflower honey correlated with Mn, Ni, Fe and Mo aqua regia soluble contents of the soil, the ash-based data correlated with Mn and Cu [4].

With respect to floral and honeydew honeys sampled in gardens and close to woods, monofloral honeys from rape, sunflowers, acacia and lindens contain higher proportions of P and Ca over K, which can be more clearly seen in the composition of (calculated) total ash contents in Tables 4, 6 and 8.

Differences between urban and rural areas

Within Table 5, data from honeys sampled within a rural area partially covered with woods and no close industrial emission (Scheibbs district), are compared with honey data from western suburbs of Vienna and Vienna Woods, from gardens in the northern part of Vienna (around the University of Natural Ressources and Life Sciences Vienna), as well as from a university building close to a highway and railway station (Biocenter Althanstrasse). Differences in plant cover result in differences in K, Cu, and Zn contents. Elements indicating pollution, like Cd, Pb, Cr, and Ni, were found at about the same levels.

Similarly, corresponding calculated ash based data (Table 6) did not show enrichments of pollutant elements at the urban site.

Table 5 Honey samples from Lower Austria and Vienna, per sample weight

Element	Scheibbs district (Lower Austria)	West of Vienna - Vienna Woods	Univ. of Nat. Res. Life Sci.	Univ. Biocenter
[mg/kg]	N = 16	N = 11	N = 7	N = 5
Al	1.86 ± 1.10	2.68 ±2.86	0.99 ±0.61	0.94 ± 0.33
As		0.0098 ±0.0033		
В	5.29 ±1.51	7.41 ±3.62	4.51 ±1.45	6.34 ± 3.82
Ca	33.69 ±8.12	40.8 ±13.8	41.12 ±9.34	33.35 ± 6.50
Cd	0.0083 ± 0.0060	0.0038 ± 0.0054	0.0019 ±0.0017	0.0008 ± 0.0004
Co	< 0.006-0.046	< 0.006-0.028	< 0.006-0.023	< 0.006
Cr	0.0047 ± 0.0024		0.0065 ±0.0047	0.0079 ± 0.0049
Cu	0.773 ± 0.303	0.340 ±0.339	0.276 ±0.053	0.211 ±0.123
Fe	1.49 ± 0.92	1.59 ±1.18	1.53 ±0.98	0.84 ± 0.52
K	2268 ±847	1161 ±732	1434 ±295	921 ±309
Li	0.0087 ± 0.0036	0.0054 ±0.0030	0.0098 ±0.0037	0.0047 ± 0.0017
Mg	65.5 ±47.8	25.1 ±18.3	18.35 ±2.59	11.68 ±2.27
Mn	2.54 ± 1.77	1.32 ±1.28	0.432 ±0.249	0.189 ± 0.037
Mo	0.0178 ± 0.0144	0.0106 ±0.0018	0.0050 ±0.0061	0.0098 ± 0.0026
Na	14.75 ±5.34	6.99 ±2.58	7.10 ±2.03	5.47 ± 1.72
Ni	0.208 ± 0.129	0.218 ± 0.193	0.152 ±0.134	0.072 ± 0.017
P	78.8 ± 27.2	53.6 ±19.0	52.1 ±10.9	38.2 ± 3.9
Pb	0.019 ± 0.010	0.019 ±0.017	0.049 ±0.050	0.032 ±0.018
\mathbf{S}^*	27.4 ±12.2	15.51 ±8.64	18.05 ±5.28	14.30 ± 3.30
Sr	0.112 ± 0.054	0.092 ±0.049	0.094 ±0.036	0.059 ± 0.014
V	< 0.007	< 0.007	< 0.007-0.015	< 0.007
Zn	2.32 ±1.21	1.06 ±0.99	0.63 ±0.21	0.91 ±0.78

^{*} incomplete recovery of methionine; N - number of samples

Table 6 Honey samples from Lower Austria and Vienna, main element composition and trace elements per ash weight

Substance	Scheibbs district (Lower Austria)	West of Vienna - Vienna Woods	Univ. of Nat. Res. Life Sci.	Univ. Biocenter
[%]	N = 16	N = 11	N = 7	N = 5
K ₂ O	85.51 ±1.81	77.5 ±11.9	85.78 ±2.16	80.47 ±8.32
P_2O_5	5.97 ±1.71	8.56 ±3.29	6.04 ±1.21	7.02 ±2.48
CaO	1.77 ±0.95	4.57 ±2.85	2.91 ±0.69	4.00 ±2.39
MgO	3.03 ±1.38	2.46 ±0.75	1.55 ±0.29	1.58 ±0.75
$\mathbf{SO_3}^*$	2.12 ±0.39	3.44 ± 3.35	2.24 ±0.45	2.73 ±0.50
Na ₂ O	0.76 ± 0.47	0.65 ± 0.25	0.49 ±0.18	0.66 ± 0.52
B_2O_3	0.65 ± 0.41	2.33 ±2.33	0.77 ±0.39	0.620 ± 0.705
[mg/kg]				
Al	556 ±230	1457 ±987	536 ±392	827 ±491
As		8.65 ± 5.86		
Cd	2.42 ±1.32	1.63 ±1.71	0.976 ±0.975	0.609 ± 0.244
Co	< 2-29	< 3-16	< 3-13	< 3
Cr	1.57 ±0.89		3.12 ±2.22	4.49 ±3.96
Cu	243 ±43	179 ±92	142 ±40	149 ±68
Fe	431 ±131	1078 ±994	789 ±587	682 ±366
Li	3.2 ±2.1	3.8 ± 3.2	5.2 ±2.5	3.4 ± 1.0
Mn	707 ±351	651 ±344	234 ±161	164 ±110
Mo	6.70 ±7.05	8.80 ±5.70	2.37 ±2.53	8.65 ±6.41
Ni	64 ±29	130 ±72	83 ±81	59 ±27
Pb	5.84 ±2.44	20.2 ±19.2	24.2 ±26.6	23.3 ±13.0

Substance	Scheibbs district (Lower Austria)	West of Vienna - Vienna Woods	Univ. of Nat. Res. Life Sci.	Univ. Biocenter
[mg/kg]	N = 16	<i>N</i> = 11	N = 7	N = 5
Sr	44 ±32	60 ±23	48 ±22	45 ±9
V	< 2	< 3	< 3-6	< 2
Zn	700 ±251	654 ±480	328 ±140	626 ±447

^{*} incomplete recovery of methionine: N - number of samples

This can be explained by the collection strategy of the bees, which moved to a park close to the riverside (Donaukanal) nearby and did not cover their surrounding homogeneously. Bees flying to the highway or railway will never come back and others go to the site where something has been found. Similarly, bee hives put close to a highway did hardly contain contaminated honey [1]. High contribution of "less mineralized" floral honey counteracts effects of the city, but even among ash-based data, the honeys from the rural area contained more Cd, Cu and Mn.

The city of Linz is a nucleus of steel and chemical industry in Austria, and had been a pollution hotspot from heavy industries in the past. Tables 7 and 8 contain honey data arranged within an approximate profile starting from rural areas north of the city, moving to northern suburbs, to a central area, and finally to southern suburbs (Molasse area). Whereas in the northern part, acid soils upon igneous rocks prevail, the city and its southern suburbs are situated in a Molasse zone of tertiary and quaternary origin.

Table 7 Honey samples around Linz/Upper Austria, per sample weight

Element	Igneous rock area; woods close	Igneous rock area; gardens	Close to the city; gardens	Molasse area; woods close
[mg/kg]	N = 8	N = 10	N = 6	N = 4
Al	0.503 ±0.305	0.596 ±0.355	0.409 ±0.141	0.517 ±0.239
В	7.61 ±2.01	6.80 ± 2.81	9.06 ±2.81	9.74 ±3.96
Ba	0.044 ± 0.154	0.084 ± 0.118	0.089 ± 0.089	-
Ca	63.6 ±17.3	65.1 ±18.5	66.0 ±13.7	48.89 ±6.52
Cd	0.0011 ±0.0007	0.0011 ± 0.0006	0.0017 ±0.0013	0.0004 ±0.0003
Co	0.018 ± 0.009	0.016 ± 0.009	0.012 ±0.007	< 0.005
Cr	0.0059 ±0.0035	0.0070 ± 0.0032	0.0049 ± 0.0028	0.0057 ±0.0028
Cu	0.352 ±0.081	0.424 ± 0.136	0.323 ± 0.059	0.140 ±0.036
Fe	1.23 ±0.82	1.07 ±0.41	1.25 ±0.40	0.72 ±0.10
K	1650 ±623	1614 ±417	1278 ±348	516 ±253
Li	0.0035 ±0.0022	0.0042 ± 0.0022	0.0042 ±0.0036	0.0097 ±0.0098
Mg	25.3 ±11.5	23.98 ±7.56	22.11 ±4.21	15.16 ±1.19
Mn	1.116 ±0.908	1.343 ±0.994	0.646 ±0.551	0.261 ±0.096
Mo	0.0140 ±0.0140	0.0124 ± 0.0057	0.0091 ±0.0034	0.0052 ±0.0015
Na	10.15 ±3.03	12.09 ±5.14	10.56 ±4.91	8.36 ±4.28
Ni	0.125 ±0.032	0.113 ± 0.030	0.121 ±0.047	0.064 ±0.027
P	81.9 ±37.2	66.8 ± 22.1	78.5 ±21.7	48.9 ±9.1
Pb	0.043 ±0.018	0.036 ±0.021	0.044 ±0.028	0.044 ±0.031
\mathbf{S}^*	16.64 ±4.91	18.17 ±5.14	19.87 ±4.41	13.36 ±2.45
Sr	0.096 ± 0.036	0.099 ± 0.040	0.068 ± 0.019	0.045 ±0.025
V	< 0.007	< 0.007-0.009	< 0.007	< 0.007
Zn	1.185 ±0.348	1.248 ±0.258	0.887 ±0.281	0.522 ±0.334

^{*} incomplete recovery of methionine; N - number of samples

Among the main constituents, decrease of K and P together with increase in B indicate an increase of floral components from north to south. The levels of contaminant elements were at the same levels in the city area and outside. Among the corresponding ash-based data only, higher Cd appeared for the city sites. Among ash-based data, Ca, Mg, Na, Al, B and S were enriched in the Molasse zone, whereas data obtained from the original samples suggest the opposite. This might reflect differences in geology, but also the effect of some woods within the action range of the bees, resulting in higher total ash and lower %K in ash contents. Possibly, the proximity of highway A1 to the sites in the Molasse zone is reflected by higher Pb and Cr just in the ash-based data, too.

Table 8 Honey samples around Linz/Upper Austria, main element composition and trace elements per ash weight

Substance	Igneous rock area;	Igneous rock area;	Close to the city;	Molasse area;
Substance	woods close	gardens	gardens	woods close
[%]	N = 8	N = 10	N = 6	N = 4
K ₂ O	82.86 ±1.99	83.37 ±2.56	78.85 ±1.86	66.50 ±8.39
P ₂ O ₅	7.73 ±1.19	6.81 ±1.15	9.20 ±1.17	12.90 ±2.07
CaO	3.86 ± 0.53	3.93 ±0.68	4.82 ±0.62	8.09 ±2.30
MgO	1.74 ±0.43	1.69 ±0.19	1.92 ±0.26	3.02 ±0.95
SO ₃ *	1.81 ±0.34	1.97 ±0.35	2.61 ±0.46	3.88 ± 0.81
Na ₂ O	0.63 ± 0.30	0.72 ±0.31	0.74 ± 0.35	1.21 ±0.27
B_2O_3	1.23 ±0.74	1.03 ±0.65	1.66 ±0.97	4.12 ±2.71
[mg/kg]				
Al	212 ±104	254 ±122	212 ±78	589 ±251
Ba	9 ±58	38 ±50	38 ±44	
Cd	0.49 ± 0.26	0.46 ± 0.21	0.92 ± 0.74	0.45 ± 0.32
Co	7.5 ± 2.4	6.5 ±2.8	6.2 ±3.0	< 6
Cr	2.57 ±1.45	3.15 ±1.71	2.53 ±1.60	7.63 ± 5.58
Cu	158 ±45	188 ±59	171 ±35	161 ±37
Fe	476 ±152	485 ±188	648 ±175	876 ±364
Li	1.4 ±0.5	1.8 ±1.0	2.2 ±2.0	9.5 ±5.7
Mn	495 ±365	568 ±382	353 ±311	333 ±199
Mo	5.01 ±3.44	5.33 ±1.84	4.62 ±0.62	5.86 ± 1.03
Ni	56 ±16	49 ±12	64 ±25	70 ±7
Pb	19.9 ±11.9	17.5 ±14.6	23.3 ±16.1	45.3 ±22.6
Sr	41 ±11	42 ±12	35 ±3	49 ±20
V	< 2	< 3-5	< 3	< 5
Zn	565 ±286	565 ±191	464 ±163	542 ±147

^{*} incomplete recovery of methionine

Conclusions

Because honey samples may cover a rather large area, their use as indicator substances to detect environmental pollution seems promising. However, data from monofloral honeys show that effects of plant origin and soil composition prevent tracing the exact composition of a non-contaminated baseline sample. Also, contrary to soils and sediments, honey samples from a pre-industrial era contained trace elements at higher levels than current.

Determination of the inorganic matrix permits the calculation of the presumable ash contents and composition, which is largely a K-phosphate borate. Interpretation of the inorganic matrix of honey samples in terms of environmental contamination is biased by effects of green plant origin, in particular differences in ash contents of floral and

honeydew honey. Because dilutions by the sugar matrix are eliminated, ash-based data reflect inputs from dust and pollen in a more reliable way. Ash-based data show Cd, Pb and Co at about the same levels than expectable in dust, whereas Cu, Mo and Zn get enriched by physiological reasons and thus should not be taken as proper indicators for environmental contaminations.

Due to the sampling strategy of the honeybees to collect at specific spots instead of homogenous sampling at the entire area, however, differences between urban, suburban and agricultural areas in Vienna and Linz were small and within experimental errors.

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