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# EFFECT OF SODIUM TRIPOLYPHOSPHATE, GLYCEROL, VITAMIN B<sub>2</sub>, MANNITOL AND SOLID MALT EXTRACT ON SURVIVAL OF *BIFIDOBACTERIUM BIFIDUM* DURING FREEZE-DRYING

Short communication –

Guowei SHU1\*, Chun YIN\*, Haipeng QIN\*\*, Haiyan KOU\*\*, Shuai HE\*, Dan HUANG\*\*\*

\*School of Food and Biological Engineering, Shaanxi University of Science and Technology, Xi'an, 710021, China

\*\*Xi'an Marsh Animal Husbandry Co., Xi'an, 710021, China \*\*\*Shaanxi Pucheng Shiyang Feed Co., Ltd., Weinan, 715501, China

**Abstract**: The effect of sodium tripolyphosphate, glycerol, vitamin B<sub>2</sub>, mannitol and solid malt extract on the freeze-dried powder survival rate and the viable cells of the *B. bifidum* BB01 were studied by single-factor test in this study. The optimal concentration of protectant for *B. bifidum* during freeze-drying were 1.5% (W/V) sodium tripolyphosphate, 12% (W/V) glycerol, 6% (W/V) vitamin B<sub>2</sub>, 6% (W/V) mannitol and 6% (W/V) solid malt extract, and the survival rate of bacteria was 18.63%, 22.98%, 24.13%, 24.19% and 39.77%, respectively.

Keywords: Bifidobacterium bifidum, freeze-drying, protective agents, probiotics.

#### INTRODUCTION

With the introduction of the concept of probiotics, probiotic food has potential market of functional food in the world, and its output value has exceeded 65% of the world's GDP (Holzapfel et al., 2006). As one of the most important probiotics, *B. bifidum* has been widely used in functional foods. Many physiological functions of *B. bifidum* have been reported: decrease serum cholesterol (Xiao et al., 2003; Beena et al., 1997); relieve lactose intolerance (Parracho et al., 2007; Shu et al., 2016); improve the digestive system (Guyonnet et al., 2009) and delay aging (Yamazaki et al.,

## **MATERIALS AND METHODS**

Microorganism and media. B. bifidum BB01 was from School of Food and Biological Engineering, Shaanxi University of Science & Technology. MRS broth was used to activate and ferment the strain; MRS agar medium was used to determine the viable cells.

Certrifugal collection of B. bifidum broth. Activated B. bifidum was cultured in MRS

2002). At present, the main way obtained by vacuum freeze-drying technology for the *B. bifidum* (Zhang et al., 2005). However, how to increase the survival rate of the bacteria in the freeze-drying process and improve the storage stability became a technical problem.

This research investigates five protective agents to reduce the mortality rate of the bacteria during the freeze-drying process. Single-factor tests are used to obtain the best concentration of sodium tripolyphosphate, glycerol, vitamin B<sub>2</sub>, mannitol and solid malt extract, which provides certain reference for the optimization of the multiple cryoprotectant.

broth at 37°C for 18h, and finally centrifuged at 6000rpm for 15min to obtain *B. bifidum* cells.

**Freeze-drying process.** Protectant of B. bifidum cells were firstly frozen at -45°C for 12-24h, and finally frozen at -55°C, 4-6pa for 24h using vacuum lyophilizer.

**Viable counts.** The dried powder was rehydrated with sterile saline and then measured viable bacteria counts when pre and post freeze-drying by standard plate count.

<sup>&</sup>lt;sup>1</sup> Corresponding author. E-Mail address: shuguowei@gmail.com

## Calculation of survival.

Survival (%) =  $\frac{\text{viable counts after freeze - drying}}{\text{viable counts before freeze - drying}} \times 100\%$ 

## The viable counts of power

 $= \frac{\text{total number of viable cells lyophilized}}{\text{power weight}} \times \text{bacterial suspension volume}$ 

## **RESULTS AND DISCUSSIONS**

Effect of sodium tripolyphosphate on viability of B. bifidum during freeze drying. Effects of various concentrations of sodium tripolyphosphate on viability of B. bifidum during freeze-drying were shown in Figure 1. concentrations the of tripolyphosphate increased from 0.5 to 1.5%, the viable counts and the survival rate of the bacteria rose rapidly, but between 1.5 to 2.5%, those both reduced. When add of sodium tripolyphosphate was 1.5%, the viable counts and the survival rate reached maximum which were 18.63% (the control group was 0.03%),  $7.75 \times 10^{10}$  cfu/g (the control group was 0.07×10<sup>10</sup>cfu/g), respectively.

Sodium tripolyphosphate and sodium glutamate are of the same class of low molecular weight compounds. Wang et al (2013) found that when add of sodium glutamate was 7g/100mL as a protective agent, the survival rate of *C. tropical* was 27.92%. The same result was confirmed from our study. However, Zhao et al (2009) found that when add of sodium glutamate was 2.5% (W/V), the survival rate of *Oenococcus oeni* reach high at 72.4%. The reason may be due to individual differences of strains.

Effect of glycerol on viability of *B. bifidum* during freeze drying. Effects of various concentrations of glycerol on viability of *B. bifidum* during freeze-drying were shown in Figure 2. When add of glycerol was 12%, the survival rate and the viable counts of *B. bifidum* reached maximum 22.98%,  $8.15 \times 10^{10}$  cfu/g, respectively.

Zeng et al (2013) found that when the concentration of glycerol was 20g/L, the survival rate of *Lactobacillus casei* reached 60%, which was 53% higher than the control group. The results showed that the glycerol as protectant had an obvious effect on viability of bacteria during freeze-drying.

Effect of vitamin  $B_2$  on viability of B. bifidum during freeze drying. Effects of

various concentrations of vitamin  $B_2$  on viability of *B. bifidum* during freeze-drying were shown in Figure 3.The viable counts and the survival rate of the bacteria increased slowly during 4%-6%, but when 6 to 10%, those both decreased rapidly. When the concentration of vitamin  $B_2$  was 6.0%, the survival rate and the viable counts of bacteria were the highest (24.13%),  $6.59 \times 10^{10} \text{cfu/g}$ , respectively.

Zhang et al (2013) found that when add of vitamin  $B_2$  was 1% (W/V), the survival rate and the viable counts of *Lactobacillus bulgaricus* were 16.3%,  $1.26 \times 10^{11}$  cfu/mL, respectively. This result showed that vitamin  $B_2$  as a protectant, the influence was not very obvious compared with other protective agents.

Effect of mannitol on viability of *B. bifidum* during freeze drying. Effects of various concentrations of mannitol on viability of *B. bifidum* during freeze-drying were shown in Figure 4. During 2 to 6%, the survival rate and the viable cells of *B. bifidum* increased rapidly and reached maximum 24.19%, 8.99×10<sup>10</sup>cfu/g, respectively.

However, Zhang et al (2010) found that when the concentration of mannitol was 50g/L, the survival rate of *Bifidobacterium bifidum* reached 88.34%, which was 85.72% higher than the control group.

Effect of solid malt extract on viability of B. bifidum during freeze drying. Effects of different concentrations of solid malt extract on survival of B. bifidum during freeze-drying shown in Figure 5. With concentrations of solid malt extract increased from 3 to 6%, the survival rate and the viable counts of bacteria increased progressively, while both of them decreased with the solid extract was 6%-15%. When concentration of solid malt extract was 6.0%, the survival rate and the viable counts of bacteria reach their maximum value which were 39.77%, 1.75×10<sup>11</sup>cfu/g, respectively.

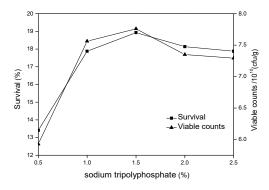


Figure 1. Effect of sodium tripolyphosphate on viability and survival of *B. bifidum* 

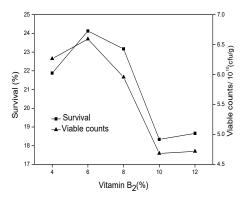
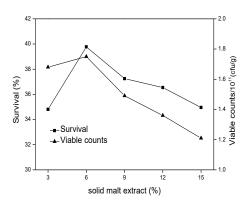


Figure 3. Effect of vitamin B<sub>2</sub> on viability and survival of *B. bifidum* 



The research indicated that it was difficult to obtain a satisfactory protective effect for a single protective agent (Dong et al., 2007; Yuan et al., 2003). Meanwhile, it was well known that malt extract was rich in maltose, glucose,

# **CONCLUSIONS**

Adding of sodium tripolyphosphate, vitamin B<sub>2</sub>, glycerol, solid malt extract and mannitol as protective agents had an obvious effect on viability of *B. bifidum* BB01 during freezedrying. In addition, adding of sodium

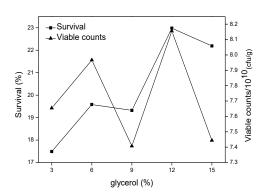


Figure 2. Effect of glycerol on viability and survival of *B. bifidum* 

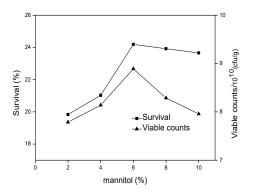


Figure 4. Effect of maniol on viability and survival of *B. bifidum* 

Figure 5. Effect of solid malt extract on viability and survival of *B.bifidum* 

laevulose, peptide, vitamin and so on (Huang, 2012). Therefore, solid malt extract that was used as protective agent of *B. bifidum* was better than other kinds of single protective agent during freeze-drying.

tripolyphosphate had minimal effects while solid malt extract had imperative influence on survival rate of *B. bifidum*. When the concentration of solid malt extract was 6.0% (W/V), the survival rate and the viable cells of *B. bifidum* both reached maximum 39.77%, 1.75×10<sup>11</sup>cfu/g.

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### **REFERENCES**

- 1. Beena, A. & Prasad, V. (1997). Effect of yogurt and bifidus yogurt fortified with skim milk powder, condensed whey and lactose-hydrolysed condensed whey on serum cholesterol and triacylglycerol levels in rats. *Journal of Dairy Research*, 64(3), 453-457.
- 2. Chen, H., Zhang, Q.H., Luo, Q. & Shu, G.W. (2013). Effect of five materials including sucrose, lactose, skim milk, yeast, vitamin B<sub>2</sub> on survival of *Lactobacillus bulgaricus* during freezedrying. *Advanced Materials Research*, 700(12), 255-258.
- 3. Guyonnet, D., Woodcock, A., Stefani, B., Trevisan, C. & Hall, C. (2009). Fermented milk containing *Bifidobacterium* lactis DN-173 010 improved self-reported digestive comfort amongst a general population of adults. A randomized, open-label, controlled, pilot study. *Journal of Digestive Diseases*, 10(1), 61-70.
- 4. Huang, G.X. (2012). Malt extract used in solid beverage. *China Food Additives*, 221-223.
- 5. Holzapfel, W.H., Goktepe, I., Juneja, V. K. & Ahmedna, M. (2006). Introduction to prebiotics and probiotics. *Probiotics in Food Safety and Human Health*. 35 (2):109-116
- 6. Helena, P., McCartney, A.L. & Gibson, G.R. (2007). Probiotics and prebiotics in infant nutrition. *Proceedings of the Nutrition Society*, 66(3): 405-411
- 7. Jinmei, H.E. & Hua, W. (2013). Optimization of cryoprotectant formulation for candida tropical by central composite design. *Food Science*, *53*(6), 1150-1.
- 8. Shu, G.W., Lei, N., Chen, H., Hu, M. & Yang, H. (2016). Application of central composite design to optimize the amount of carbon source and prebiotics for *Bifidobacterium bifidum* BB01. *Acta Universitatis Cibiniensis*. *Series E: Food Technology*, 20(1), 41-52. DOI: 10.1515/aucft-2016-0003.
- 9. Xiao, J.Z., Kondo, S., Takahashi, N., Miyaji, K., Oshida, K. & Hiramatsu, A., et al. (2003). Effects of milk products fermented by bifidobacterium longum on blood lipids in rats and healthy adult male volunteers. *Journal of Dairy Science*, 86(7), 2452-2461.
- 10. Yamazaki, S., Kaneko, T., Taketomo, N., Kano, K. & Ikeda, T. (2002). 2-amino-3-carboxy-1,4-naphthoquinone affects the end-product profile of *Bifidobacteria* through the mediated oxidation of NAD(p)H. *Applied Microbiology and Biotechnology*, 59(1), 72-78.
- 11. Ying, D., Ze-Ying, L., Guo-Lei, Z., Zhao-Guan, L. & University, J. (2007). Study on Cryoprotector for *Lactic Acid* Bacteria Starter in pickles. *Food Industry*, 14 (1):10-12
- 12. Ya-Hong, Y., Tian-Li, Y., Zhen-Peng, G. & Duo-Yong, Z. (2003). Research on the precess of high active lactobacillaceae powder. *Journal of Northwest Sci-Tech University of Agriculture and Forestry*, 31(S1), 82-84
- 13. Zhang Y.H., Huo G.C. & Guo L. (2005). Research advance of the freeze-drying technology of lactic acid bacteria. *Journal of Northeast Agricultural University*, 36 (6): 799-803
- 14. Zeng X.Q., Pan D.D., Bao H.Y., Liu T.W. & Xu H.N. (2013). Research on cryoprotectant of *Lactobacillus Casei*. *Journal of China Institute of Food Science and Technology*, 13(1): 44-50
- 15. Zhang F., Zhao M., Li L.Y., Lu X.F., Wang Y. & Zhang R. (2010). Preparation of lyophilized *Bifidobacterium bifidum* Powder. *Food Science*, 31(15): 221-224
- 16. Zhao, G.Q. & Gui, Z. (2009). Influences of protectants, rehydration media and storage on the viability of freeze-dried oenococcus oeni for malolactic fermentation. *World Journal of Microbiology & Biotechnology*, 25(10), 1801-1806.