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HLA typing in autoimmune diseases

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ABSTRACT

Genome scan, linkage and association studies have identified CMH genes in the genetic determinism of most autoimmune diseases, affecting approximately 5% of the population.

The susceptibility conferred by HLA alleles does not influence the development of autoimmunity in general, but rather the probability of some disorders.

From extremely high number of known HLA alleles, less than 30 are associated with diseases, and fewer than 10 are involved in the strongest associations.

The aim of this study was to determine HLA genes in autoimmune diseases.

The method used for the assignment of HLA alleles was molecular genotyping, primarily by the sequence specific oligonucleotide hybridization method (SSO), and when required, by the sequence specific primers method (SSP).

Molecular genotyping was performed in 282 cases of diabetes mellitus type 1, Hashimoto thyroiditis, Graves' disease and ankylosing spondylitis.

HLA typing in patients with autoimmune diseases is important for determining prognosis and genetic counseling. In ankylosing spondylitis HLA typing is used for differential diagnosis. In such studies it's important to be aware that there is a particular HLA gene expression depending on the geographic area.

Keywords: autoimmune diseases, major histocompatibility complex, risk alleles

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Introduction

Hundreds of diseases were tested for association with HLA. More than 50 statistical significant associations of HLA genes with different diseases are found so far [1].

Genome scan, linkage and association studies have identified CMH genes in the genetic determinism of most autoimmune diseases, affecting approximately 5% of the population [1].

CMH contribute at least 30% of familial aggregation in insulin-dependent diabetes mellitus, systemic lupus erythematosus and rheumatoid arthritis [1,2].

The association between a particular HLA allele and a disease is expressed as relative risk for the disease given the presence of particular HLA alleles (Table I).

It should be noted, however, that the same HLA allele may provide protection for a disease and at the same time, to confer susceptibility to another - for example alleles DQA1 * 0102, DQB1 * 0602 associated to HLA-DR2 are rare in patients with diabetes type 1, but is associated with multiple sclerosis [1,2].

It seems, therefore, that the susceptibility conferred by HLA alleles does not influence the development of autoimmunity in general, but rather the probability of some disorders.

From extremely high number of known HLA alleles, less than 30 are associated with diseases, and fewer than 10 are involved in the strongest

associations.

Some researchers believe that these associations are conserved haplotypes that existed millions of years - known as ancestral haplotypes (for example haplotype A1, B8, DR3) [1]. From recombinant ancestral haplotypes results 75% of the haplotypes in the Caucasian population. Identifying the type of histocompatibility antigens present on cells of an individual, it is called HLA typing.

Table I. Several significant associations of increased risk HLA alleles for various diseases are. Adapted from Goldsby et al., Immunology 5th ed. 2003

Disease	HLA associated	The relative risk*
Retinopathy "birdshot"	A29	150
Ankylosing spondylitis	B27	90
Reiter syndrome	B27	37
Reactive arthritis (Yersinia, Salmonella, Gonococcus)	B27	18
Narcolepsy	DR2	
Sensitivity to gluten enteropathy	DR3/5/7,DQ2	130
		30
Insulin-dependent diabetes mellitus	DR4/DR3, DQ2/DQ8	20
Goodpasture syndrome	DR2	16
Myasthenia gravis	DR3	10
Rheumatoid arthritis	DR4	10
Multiple sclerosis	DR2, DR6	5
Systemic lupus erythematosus	DR3	5
Hashimoto's thyroiditis	DR5	3

* Relative risk is calculated by dividing the frequency of HLA in the patient population at its frequency in the general population.

Aim of study

The aim of this study was to determine HLA genes in autoimmune diseases.

Material and method

During October 2008 - September 2011 HLA genotyping was performed in 282 cases registered at Molecular Laboratory of Medical Genetics Department for following diseases: 62 for diabetes mellitus type 1, 98 for Hashimoto thyroiditis, 112 for Graves' disease and 10 for ankylosing spondylitis.

The method used for the assignment of HLA alleles was molecular genotyping, primarily by the sequence specific oligonucleotide hybridization method (SSO), and when required, by the sequence specific primers method (SSP).

In the SSO method, polymorphic regions of genes of class I and II are amplified from genomic DNA by PCR reaction. Then, resulting amplicons are used in hybridization reaction with specific oligonucleotide probes, selected to detect regions with polymorphism. (See Figure 1 a, b).

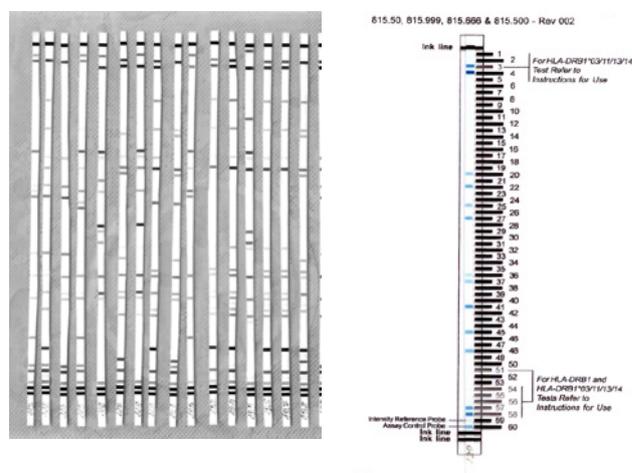


Figure 1. a. Image of a batch of strips after the completion of the RELI SSO HLA-DRB1 assay. 17 amplifications followed by colorimetric detection were run simultaneously. b. Interpretation of the result – the strip from the image shows the following specificities: 3, 4, 20, 22, 25, 27, 36, 37, 41, 45, 48, 59, 60 and the genotype assigned by the Pattern Matching Program were DRB1*0401 & 1101.

The ambiguities were solved with two

sequence-specific primers systems (SSP). DNA amplification occurs only if in genomic DNA it is a sequence perfectly complementary with primers. The presence or absence of PCR products of correct size is than analyzed by electrophoresis. (see Figure 3 and 4).

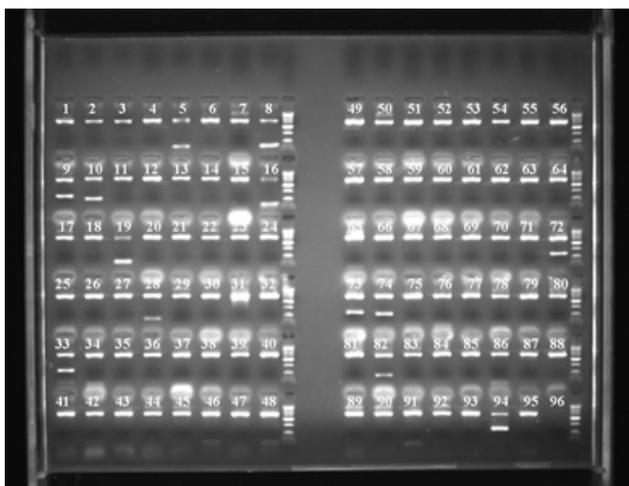


Figure 3. Gel image photographed by a digital camera of the gel documentation system. Control lines are observed (black arrow) in all tracks except number 96 which is negative amplification control. That shows the presence of all amplification reactions (1-95). Lack of amplification products in position 96 shows no contamination. Specific lines (5, 8, 9, 10, 16, 19, etc. open black arrow) indicate the presence of specific polymorphism in the HLA-DRB1 gene structure. Weak nonspecific lines can be observed (white arrow) that are usually unused primers or primer dimmers.

Then gel photo is print and apply on gel documentation sheet. After a carefully examination of the gel are determined the positive tracks. They are marked on the form (Figure 4).

After establishing positive lines, alleles designation is made by using a computer program called UniMatch™ (Figure 5). In the dialog is selected the type of determination used (e.g. High Resolution HLA-DRB1), patient data recorded and specificities found (positive lines on electrophoresis).

invitrogen Gel Documentation Form
 A/ISet™ Gold SSP HLA-DRB1 High Res Kit
 Invitrogen Corporation
 Tel: 1-800-955-6288
 Fax: 1-800-331-2286
 www.invitrogen.com

Name: PATRICE LAURENTI Sample I.D.: 101 DR Date: 2.08.2009
 Institution: FAC. MEDICINA Lot: 014 Batch: 637819 Exp. Date: 2010-11
 Tested By: WROBAL IRINA Tray Identification Number: 540408

Gel Picture

Well location	H1	G1	F1	E1	D1	C1	B1	A1	H2	G2	F2	E2	D2	C2	B2	A2	H3	G3	F3	E3	D3	C3	B3	A3
Lane number	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24
Size (bp)	210	85	230	210	185	148	230	105	208	230	176	176	180	200	220	180	210	180	100	200	200	200	148	130
	200	78	210	200	85	110	150		170	180	145	115		210	85	200	145		130	105				
									130	130		75	110			200	195							
																180								

Well location: H4 G4 F4 E4 D4 C4 B4 A4 H5 G5 F5 E5 D5 C5 B5 A5 H6 G6 F6 E6 D6 C6 B6 A6
 Lane number: 25 26 27 28 29 30 31 32 M 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48
 Size (bp): 225 140 265 180 180 180 200 220 220 170 195 90 185 230 165 180 255 120 115 180 235 245 170 100 190 200 210 255 210 180 110 200 185 185 185 170 180 95 70 180 95 110 130 215 235 185 70 185 115 120 200 225 75 170

Well location: H7 G7 F7 E7 D7 C7 B7 A7 H8 G8 F8 E8 D8 C8 B8 A8 H9 G9 F9 E9 D9 C9 B9 A9
 Lane number: 49 50 51 52 53 54 55 56 M 57 58 59 60 61 62 63 64 65 66 67 68 69 70 71 72
 Size (bp): 105 225 225 210 268 220 155 210 185 230 250 230 210 155 165 185 85 130 175 170 225 190 240 215 90 119 200 210 255 210 180 110 200 185 185 185 170 180 95 128 70 185 205 115 125 150 140 95 95

Well location: H10 G10 F10 E10 D10 C10 B10 A10 H11 G11 F11 E11 D11 C11 B11 A11 H12 G12 F12 E12 D12 C12 B12 A12
 Lane number: 81 82 83 84 85 86 M 87 88 89 90 91 92 93 94 95 96
 Size (bp): 200 115 175 175 118 225 148 100 215 180 225 160 255 130 45 250 200 205 165 215 245 245 215 135 105 115 100 185 70 90 210 130 100 185 105 175 115 110 155 130 105 220 210 105 145 105 145 140 80 85

Positive Lanes: 5, 8, 9, 10, 16, 19, 28, 33, 72, 73, 74, 82, 94
 Failed Lanes:
 Contamination (lane 96): Yes No

Size of internal control PCR fragment: app. 796 bp
 Base pair sizes are approximate.
 For detailed information, please see the lot specific worksheet or primer mix specificity table.

Figure 4. Gel documentation form. Gel photograph is attached by the form. Patient data and data about the kit used are note too. Tracks marked with specific amplification, those were amplification failed and presence or absence of contamination.

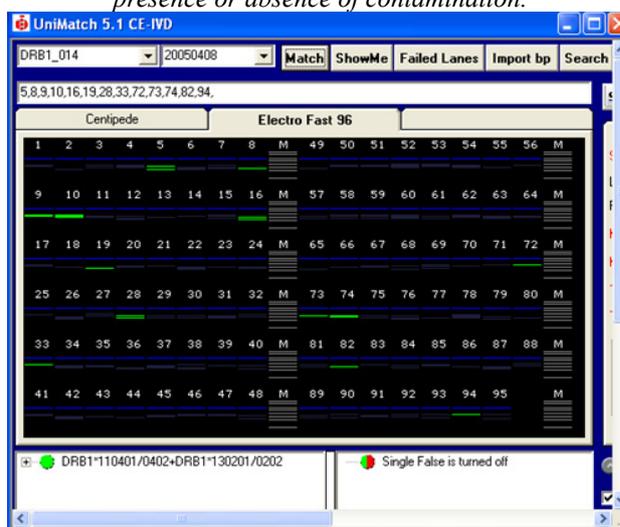


Figure 5. UniMatch™ program analysis window: determination of HLA-DRB1 High Resolution. The specificities marked as positive are: 5, 8, 9, 10, 16, 19, 28, 33, 72, 73, 74, 82, 94. HLA-DRB1 alleles are designated: * 1104 and * 1302

Results and discussions

In our laboratory were 282 HLA typing for the following diseases: diabetes mellitus type 1, Hashimoto thyroiditis, Graves' disease, celiac disease and ankylosing spondylitis.

From 62 patients with type 1 diabetes 95.1% have one or both risk alleles DR3 or DR4, similar to those of other studies that have analyzed the HLA genes in 14168 children with type 1 diabetes [1].

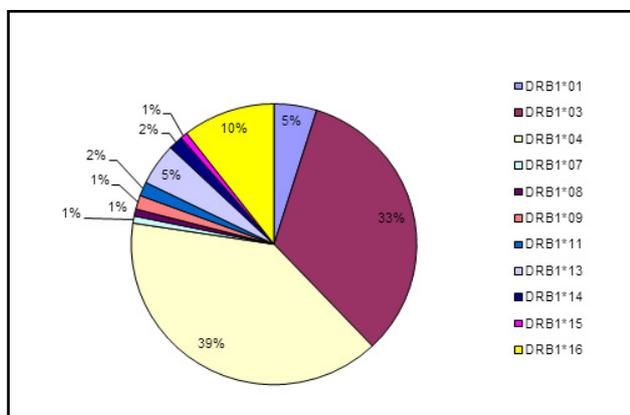


Figure 6. The frequency of HLA-DRB1 alleles in patients with DM1

Frequency of genotype DR3 / 4 (30.64%) was slightly lower than the frequencies of 36-38% in the studies mentioned, but was slightly higher than in other studies of type 1 diabetes from Romania [1,2], who reported a frequency of 20.75% for this genotype. Non DR3-nonDR4 genotype frequency was lower in our study (4.9%) than in other studies from Romania, in which had found a frequency of 10.8% for this genotype. Results are similar for DQ due to linkage disequilibrium between DR3 – DQ2 and DR4 – DQ8.

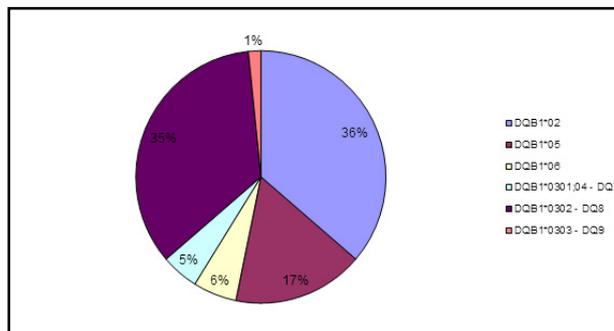


Figure 7. The frequency of HLA-DQB1 alleles in patients with DM1

4 patients with DM1 had also celiac disease. We found 3 patients with DR3-DQ2 and one with DR4 – DQ8. 95% of patients with celiac disease have one of the genes HLA DQ2 or DQ8. The association between the two diseases can be explain by common susceptibility genes conferred by HLA class II.

In Caucasians, the risk alleles of HLA class II genes are considered HLA-DRB1*04, *05 (divided into *11 and *12 allele groups) and inconstant HLA-DRB1*03 for Hashimoto thyroiditis [3].

In our study from Dobrogea, besides HLA-DRB1*11 (DR5) risk alleles, we found also HLA-DRB1*03 and HLA-DRB1*16 predisposing for Hashimoto thyroiditis.

From 98 patients with HT only 8 associated diabetes mellitus type 1 and 4 rheumatoid arthritis. These associations are expected because they share the same HLA-DRB1 risk alleles. However 74.5% of cases are not associated with other autoimmune diseases.

In a multicenter study from England, only 14% of patients with Hashimoto thyroiditis associate other autoimmune diseases, and the most frequent was rheumatoid arthritis [3].

In most studies, HLA-DRB1*03 alleles are predisposing to Graves' disease in Caucasians [1]. In our study both HLA-DRB1*03 and *04 are predisposing to Graves' disease. From 112 patients only 10 associated rheumatoid arthritis and all had HLA- DRB1*04. 9 patients with GD associated DM 1 and all had HLA-DRB1*03 or *04.

In a study of 2791 Caucasian with Graves' disease, only 9.67% had other autoimmune diseases

of which 15.3% had rheumatoid arthritis [1,2].

Only 10 patients with ankylosing spondylitis were typed for HLA. 3 of them were homozygous and 7 heterozygous for HLA B*27. There are described 40 allelic variants but only HLA B*27:05 is present in 95% English caucasians. Subtype HLA B*27:06/09 have a low penetrance.

Conclusions

HLA typing in patients with autoimmune diseases is important for determining prognosis and genetic counseling. In ankylosing spondylitis HLA typing is used for differential diagnosis.

If the predisposition for the diseases is determined by all the alleles of the group, than a low resolution HLA typing is performed. But if it is necessary to identify exactly the allele from a group, than a high resolution HLA typing is indicated.

In such studies it's important to be aware that there is a particular HLA gene expression depending on the geographic area.

References

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