

*Short communication***INHIBITION OF *IN VITRO* GROWTH OF PORCINE ENTEROTOXIGENIC AND SHIGA TOXIN-PRODUCING *ESCHERICHIA COLI* BY *LACTOBACILLUS PLANTARUM* STRAINS**

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The objective of this study was to assess cell-free fermented culture broth of 9 *Lactobacillus plantarum* strains as antibiotic alternatives for the inhibition of *in vitro* growth of enterotoxigenic *Escherichia coli* (ETEC) and Shiga toxin-producing *E. coli* (STEC) isolated from postweaning pigs with colibacillosis and edema disease in 2014. A total of 10 ETEC and 5 STEC strains isolated from postweaning pigs were tested in antimicrobial susceptibility tests. ETEC and STEC strains used in this study possessed at least one of fimbrial, enterotoxin, and Shiga-toxin genes when tested by polymerase chain reaction. Among 9 *L. plantarum* strains tested, 3 strains (Lp 2-05, 2-06, and 1-03) showed inhibitory activity of *in vitro* growth against 10 ETEC strains (100%) and 7 strains (Lp 6-13, 3-06, 3-05, 7-01, 2-06, 1-03, and 6-05) showed inhibitory activity of *in vitro* growth against 5 STEC strains (100%). Three strains (Lp 3-05, 2-06, and 1-03) showed inhibitory activity of *in vitro* growth against 10 ETEC (100%) and 5 STEC (100%) strains. The results of this study show the inhibitory activity of cell-free fermented culture broth of *L. plantarum* against ETEC and STEC isolated from postweaning pigs with colibacillosis and edema disease.

**Key words:** *Escherichia coli*, *in vitro* growth inhibition, *Lactobacillus plantarum*

## INTRODUCTION

Enterotoxigenic *Escherichia coli* (ETEC) and Shiga toxin-producing *E. coli* (STEC) are a major cause of diarrhea and edema disease and cause serious losses in postweaning pigs. ETEC has 2 types of virulence factors, fimbriae and enterotoxins. The fimbriae enable the ETEC to colonize the small intestine of piglets by mediating adhesion to the microvilli of epithelial cells. Enterotoxins (heat-labile, LT and heat-stable, ST) stimulate secretion of electrolytes and fluids by intestinal epithelial cells, resulting in

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diarrhea and dehydration [1]. Shiga toxin (Stx) produced by STEC causes vascular necrosis in small blood vessels, resulting in edema at specific locations [2-4]. *E. coli* carrying F4 (K88), F18, and Stx genes are widely prevalent in pigs with colibacillosis and edema disease [5].

Antibiotics have been commonly used to treat colibacillosis and edema disease in pigs. However, overuse of antimicrobials has resulted in an increase of antimicrobial resistance and failure of treatment of these diseases in pigs farms [6,7]. Therefore, special attention has been paid to the use of antibiotics in animals because the development of antibiotic resistant bacteria would produce a real threat to human health [8-10].

In an effort to reduce antibiotic use in animals, the Korean government has attempted to limit antibiotics use in animal husbandry, especially antimicrobials used in human medicine. This program was extended to phase out all antibiotic feed additives beginning in July 2011 (Ministry for Food, Agriculture, Forestry and Fisheries, Subparagraph No. 2010-142). After the ban of antibiotic feed additives was in effect, 857 postweaning pigs were examined at the Department of Veterinary Pathology at the Seoul National University. Out of 857 pigs that were examined during the 2-year period 2012-2013 immediately following the ban, 274 (32%) showed symptoms of colibacillosis and 32 (3.7%) exhibited edema disease. This represents a 4-fold increase in colibacillosis and an 8-fold increase in edema disease compared to the 2-year period 2010-2011, preceding the ban. This suggests the ban of the antibiotic feed additives may have contributed to the increase in the prevalence of the disease which was observed following the ban. Therefore, there is a need to improve the control and treatment of ETEC and STEC infections without the use of antibiotic feed additives. The objective of this study was to determine the ability of cell-free fermented culture broth from several *Lactobacillus plantarum* strains, as antibiotic alternatives, to inhibit in vitro growth of ETECs and STECs isolated from postweaning pigs with colibacillosis and edema disease.

## MATERIALS AND METHODS

### ***Escherichia coli* isolates**

All of the 10 ETEC and 5 STEC isolated from postweaning pigs with colibacillosis or edema disease in 2014 were used for antimicrobial susceptibility tests. ETEC isolates used in this study carry at least one of fimbrial or enterotoxin genes (Table 1) [5,11] while STEC isolates carry a gene expressing Shiga-toxin as determined by polymerase chain reaction (PCR) as previously described [5].

### ***Agar spot test***

A total of 9 *L. plantarum* strains were isolated from the feces of a healthy commercial pig to determine inhibitory activity of in vitro growth against ETEC and STEC strains.

A modification of the agar spot test was used for the determination of antimicrobial activity against *E. coli* as previously described [12,13]. Briefly, a mixture of cell-free fermented culture broth of *L. plantarum* (Genebiotech Co. Ltd., Seoul, Korea) was filtered with a 0.2 µm membrane and 2 µl of mixture of cell-free fermented culture broth of *L. plantarum* was spotted on 1.5% de Man-Rogosa-Sharpe (MRS) agar plates. Plates were dried for 30 minutes at room temperature and overlaid with 10 ml of Tryptic Soy agar (0.85%) at 45°C seeded with 1% (v/v) of an overnight culture of *E. coli* to obtain a final concentration of 10<sup>6</sup> colony forming units/ml. Plates were then incubated at 37°C and measured for zone of inhibition (clear agar) after 12 hours. Inhibition of *in vitro* growth test was performed in triplicate on each *L. plantarum* strain against ETEC and STEC. Preparation of agars, dilutions, and inoculations were performed by the same technician for consistency. All plates were set up and read by same technician.

**Table 1.** Enterotoxigenic *Escherichia coli* (ETEC) and Shiga toxin-producing *E. coli* (STEC) strains used in this study

<i>E. coli</i> strains	Virulence profile				Shiga toxin
	Fimbriae		Enterotoxin		
	F4(K88)	F18	Heat-labile	Heat-stable	
ETEC-01	O	X	O	O	X
ETEC-02	O	X	X	O	X
ETEC-03	O	X	O	X	X
ETEC-04	O	X	O	X	X
ETEC-05	O	X	O	X	X
ETEC-06	X	O	X	O	X
ETEC-07	O	X	X	O	X
ETEC-08	O	X	O	X	X
ETEC-09	O	X	O	O	X
ETEC-10	O	O	O	O	X
STEC-01	X	X	X	O	O
STEC-02	X	O	X	O	O
STEC-03	X	O	X	X	O
STEC-04	X	O	X	X	O
STEC-05	X	O	X	X	O

### Statistical analysis

Growth data were analyzed using a one-way analysis of variance (ANOVA). If the ANOVA showed a significant effect, Tukey's test for multiple comparisons was performed. A value of  $P < 0.05$  was considered to be significant.

## RESULTS AND DISCUSSION

Cell-free fermented culture broth of *L. plantarum* inhibited *in vitro* growth of ETEC and STEC. Basal medium that was not incubated with *L. plantarum* was used as the negative control and had no inhibitory effect on ETEC and STEC growth. The zone of inhibition by *L. plantarum* against ETEC is presented in Table 2. Among 9 *L. plantarum* strains, 3 strains (Lp 3-05, 2-06, and 1-03) showed inhibitory activity of *in vitro* growth against all 10 ETEC strains (100%). One strain (Lp 7-01) showed inhibitory activity of *in vitro* growth against 9 ETEC strains (90%). One strain (Lp 6-01) showed inhibitory activity of *in vitro* growth against 8 ETEC strains (80%). Strain Lp 3-05 showed a significantly larger ( $P < 0.05$ ) inhibitory zone against all 10 ETEC strains compared to strain Lp 6-05. The zone of inhibition by *L. plantarum* against STEC is summarized in Table 3. Among 9 *L. plantarum* strains, 7 strains (Lp 6-13, 3-06, 3-05, 7-01, 2-06, 1-03, and 6-05) showed inhibitory activity against all 5 STEC strains (100%). Three strains (Lp 3-05, 2-06, and 1-03) showed inhibitory activity all 10 ETEC (100%) and all 5 STEC (100%) strains.

The results of this study demonstrate that *L. plantarum* strains are promising probiotics to control colibacillosis and edema disease in postweaning pigs. These observations are clinically meaningful because outbreaks of colibacillosis and edema disease are rapidly increasing since the restriction of antibiotic feed additives was implemented by the government. An increase of antimicrobial resistance by excessive and improper use of antibiotics is the main cause of failure to effectively treat colibacillosis and edema disease in pig farms [6,7]. The current trend worldwide emphasizes reduction of antibiotic use in animal production in favor of good husbandry practices, and use of antibiotic alternatives and vaccination [14]. Therefore, a cell-free fermented culture broth of *L. plantarum* should be considered as an alternative treatment for the prevention of porcine ETEC and STEC infection. Although antimicrobial activity of *L. plantarum* has not been fully understood, the antimicrobial activity has been attributed to the production of antimicrobial substances, like oxygen peroxide, organic acids, biosurfactants, and bacteriocins [15].

The present data show the inhibitory activity of cell-free fermented culture broth of *L. plantarum* against ETEC and STEC isolated from postweaning pigs with colibacillosis and edema disease. Interestingly, different *L. plantarum* strains have different inhibitory activities against ETEC and STEC strains. Despite the fact that *in vitro* results may not reflect *in vivo* results, it is necessary to screen and select potential candidate *L. plantarum* strains for further *in vivo* antimicrobial studies.

**Table 2.** Inhibition zone of cell-free fermented culture broth of 9 *Lactobacillus plantarum* strains against 10 enterotoxigenic *Escherichia coli* (ETEC) isolated from pigs with colibacillosis by agar spot test

Inhibition zone diameter (mm) in ETEC isolates											
<i>L. plantarum</i> strains	01	02	03	04	05	06	07	08	09	10	Mean
Lp 6-01	8.3±0.7	4.7±0.6	4.3±0.5	5.5±0.6	8.5±0.9	—	9.5±0.7	5.6±0.8	—	5.5±0.8	5.1±3.2 <sup>a,b</sup>
Lp 6-13	5.4±0.9	8.2±1.1	4.5±0.5	9.1±0.8	5.9±0.5	7.2±0.6	4.8±0.7	8.4±0.7	—	—	5.3±3.2 <sup>a,b</sup>
Lp 3-06	8.2±0.7	4.8±0.6	9.2±0.6	5.8±0.9	6.2±0.5	7.0±0.7	5.9±0.5	—	4.3±0.8	—	5.1±3 <sup>a,b</sup>
Lp 3-05	9.5±0.8	9.4±0.7	8.2±0.8	8.0±0.8	8.4±0.8	7.7±0.7	7.6±0.9	7.5±0.8	8.1±0.8	7.4±0.6	8.1±0.7 <sup>a</sup>
Lp 7-25	8.3±0.8	4.8±0.7	8.1±0.8	8.4±0.8	5.6±0.5	—	4.8±0.6	—	5.6±0.6	5.2±0.9	5±3 <sup>a,b</sup>
Lp 7-01	5.7±0.9	8.4±0.6	7.0±0.5	4.2±0.5	3.9±0.7	6.5±0.7	6.5±0.9	4.6±0.9	4.4±0.8	—	5.1±2.3 <sup>a,b</sup>
Lp 2-06	8.0±0.8	7.5±0.5	4.8±0.7	6.4±0.7	6.5±0.9	7.4±0.6	6.5±0.8	5.4±0.7	4.7±0.5	7.5±0.6	6.4±1.1 <sup>a,b</sup>
Lp 1-03	6.4±1.1	7.1±0.5	7.2±0.7	5.5±0.6	6.4±0.6	8.2±0.5	4.4±0.6	7.4±0.7	7.5±0.7	8.5±0.8	6.8±1.2 <sup>a,b</sup>
Lp 6-05	8.2±0.8	5.5±1.1	7.5±0.5	5.9±0.7	4.2±0.6	4.3±0.7	—	5.0±0.5	—	4.9±0.5	4.5±2.7 <sup>b</sup>

<sup>a,b</sup>Different letters (a and b) indicate significant ( $P < 0.05$ ) difference among *L. plantarum* strains.

**Table 3.** Inhibition zone of cell-free fermented culture broth of 9 *Lactobacillus plantarum* strains against 5 Shiga toxin-producing *Escherichia coli* (STEC) isolated from pigs with edema disease by agar spot test

<i>L. plantarum</i> strains	Inhibition zone diameter (mm) in STEC isolates					
	01	02	03	04	05	Mean
Lp 6-01	6.5±0.5	-	6.8±0.8	5.4±0.9	7.0±0.8	5.1±2.9
Lp 6-13	4.3±0.5	6.1±0.8	5.8±0.7	6.8±0.7	5.4±0.5	5.6±0.9
Lp 3-06	5.2±0.2	4.9±0.5	7.3±0.9	7.1±0.9	6.2±0.9	6.1±1
Lp 3-05	5.2±0.5	6.3±1.2	7.1±0.8	6.4±0.5	7.0±0.3	6.4±0.7
Lp 7-25	6.8±0.9	5.2±0.7	-	6.6±0.8	7.1±0.8	5.1±2.9
Lp 7-01	5.5±0.5	5.2±0.3	6.4±0.9	6.2±0.6	5.3±0.9	5.7±0.5
Lp 2-06	6.2±0.7	7.0±0.7	7.2±1.1	6.3±0.5	6.7±0.9	6.6±0.4
Lp 1-03	6.1±0.8	6.8±0.9	6.8±0.9	6.9±0.8	7.3±0.9	6.7±0.4
Lp 6-05	6.5±0.4	5.4±0.8	5.5±0.4	7.2±0.9	7.2±0.7	6.3±0.8

## CONCLUSION

Among 9 *L. plantarum* strains tested, 3 strains (Lp 3-05, 2-06, and 1-03) showed inhibitory activity of *in vitro* growth against 10 ETEC (100%) and 5 STEC (100%) strains tested. These observations are clinically meaningful because outbreaks of colibacillosis and edema disease are rapidly increasing since the restriction of antibiotic feed additives was implemented by the government. *L. plantarum* strains are promising probiotics to control colibacillosis and edema disease in postweaning pigs.

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## **INHIBICIJA *IN VITRO* RASTA ENTEREROTOKSIČNE I ŠIGA TOKSIN PRODUKUJUĆE *E. COLI* POMOĆU *LACTOBACILLUS PLANTARUM* SOJEVA KOD PRASADI**

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Cilj rada je bio procena fermentisanog rastvora 9 *Lactobacillus plantarum* sojeva kao alternativa antibioticima za inhibiciju *in vitro* rasta enterotoksične *E. coli* (ETEC) i sojeva *E. coli* koji proizvode šiga toksin (STEC) izolovane kod prasadi sa kolibacilozom ili edemskom bolešću nakon zalučenja tokom 2014. godine. Antimikrobna osetljivost je testirana na ukupno 10 ETEC i 5 STEC sojeva izolovanih kod prasadi nakon zalučenja. ETEC i STEC sojevi koji su korišćeni u ovoj studiji posedovali su barem jedan od fimbrijalnih, enterotoksičnih i šiga toksin gena ustanovljenih PCR metodom. Od 9 *L. plantarum* testiranih sojeva 3 soja (Lp 2-05, 2-06, and 1-03) su imala inhibitornu aktivnost na *in vitro* rast 10 ETEC sojeva (100%), dok je 7 sojeva (Lp 6-13, 3-06, 3-05, 7-01, 2-06, 1-03, and 6-05) pokazalo *in vitro* inhibitornu aktivnost rasta 5 STEC sojeva (100%). Tri soja (Lp 3-05, 2-06, and 1-03) je *in vitro* pokazalo inhibitornu aktivnost na rast 10 ETEC (100%) i 5 STEC (100%) soja. Rezultati ove studije ukazuju na inhibitornu aktivnost fermentisanog hranjivog medijum *L. plantarum* na rast ETEC i STEC izolovanih kod zalučene prasadi sa kolibacilozom i edemskom bolešću.