Synthesis and characterization of some novel 1,2,4-triazoles, 1,3,4-thiadiazoles and Schiff bases incorporating imidazole moiety as potential antimicrobial agents

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(1,4,5-Triphenylimidazol-2-yl-thio)butyric acid hydrazide (3) was obtained via alkylation of 1,4,5-triphenylimidazol-2thiol (1) with ethylbromobutyrate, followed by addition of hydrazine hydrate. Treatment of acid hydrazide 3 with carbon disulfide in an ethanolic potassium hydroxide solution gave the intermediate potassium dithiocarbazinate salt, which was cyclized to 4-amino-5-[(1,4,5-triphenylimidazol--2-yl)thiopropyl]-2H-1,2,4-triazole-3-thione (4) in the presence of hydrazine hydrate. Condensation of compound 3 with alkyl/arylisothiocyanate afforded the corresponding 1-[4-(1,4,5-triphenylimidazol-2-ylthio)butanoyl]-4-alkyl/arvlthiosemicarbazides (5-7), which upon refluxing with sodium hydroxide, yielded the corresponding 1,2,4-triazole--3-thiols 8-10. Under acidic conditions, compounds 4-6 were converted to aminothiadiazoles 11-13. Moreover, the series of Schiff bases 14-18 were synthesized from the condensation of compound 3 with different aromatic aldehydes. The newly synthesized compounds were characterized by IR, ¹H NMR, ¹³C NMR and mass spectral analyses. They were also preliminarily screened for their antimicrobial activity.

Keywords: imidazole, 1,2,4-triazole, 1,3,4-thiadiazole, Schiff bases, antimicrobial activity

Imidazole nucleus has emerged as an important class of azoles used in various applications. In addition, thioimidazole nucleus constitutes the active part of several biologically active compounds with antifungal (1) and antiasthmatic (2) activity.

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Literature survey reveals that several 1,2,4-triazole derivatives have been investigated as therapeutically interesting compounds due to the large diversity of their effects, such as anticonvulsant (3), antimicrobial (4), analgesic (5), antioxidant (6), anti-inflammatory (7), pesticidal (8), insecticidal (9), herbicidal (10) and fungicidal ones (11).

Compounds bearing 1,3,4-thiadiazole moieties have displayed a number of chemotherapeutic properties including antibacterial (12), antifungal (13), antitubercular (14) and analgesic ones (15). On the other hand, Schiff bases are considered privileged structures in drug discovery, with significant antimicrobial, antiviral, antifungal, antibacterial and anti-inflammatory (16–20) activities as well as chelating properties towards various metal ions (21).

Prompted by these observations and in continuation of our search on the synthesis of heterocyclic compounds containing nitrogen, sulfur and bicyclic systems with potential biological activity (22–24), we have designed the synthesis of a novel series of imidazole derivatives. In particular, we emphasized the strategy of combining chemically different but pharmacologically compatible molecules (the imidazole nucleus and the 1,2,4-triazole, 1,3,4-thiadiazole and/or Schiff base moiety) in one frame in order to study their antifungal and antibacterial activities.

EXPERIMENTAL

General

Melting points were determined on a variable heater (Stuart, UK) Melt-temp apparatus and are uncorrected. Percoated silica gel (Kieselgel, 0.25 mm, 60 F254, Merck, Germany) were used for thin layer chromatography. A developing solvent system of ethyl acetate/ hexane (1/2) was used and the spots were detected by ultraviolet light. NMR spectra were measured with a Bruker 400 MHz (¹³C: 100 MHz) spectrometer (Bruker, Switzerland). Chemical shifts are expressed in δ -values (ppm) relative to TMS as an internal standard, using DMSO- d_6 as a solvent. The IR spectra were measured in potassium bromide pellets using a Perkin-Elmer 1430 series FTIR spectrometer (Perkin-Elmer, USA). Full scan accurate mass spectra (mass range from 200 to 1500 Da) were obtained at high resolution [100,000, full-width half-maximum (FWHM) at 400 m/z] on a LTQ-FT Orbitrap Velos mass spectrometer (Thermo Electron GmbH, Germany). The hybrid mass spectrometer was equipped with a heated electrospray ionization (ESI) probe for the ion max source and operated in the positive mode. The electrospray source conditions were: capillary voltage 3.75 kV, heated capillary temperature 275 °C, source temperature 450 °C and S-Lens RF level 60 %. Nitrogen gas flows were set at 40 for the sheath gas and at 20 for the auxiliary gas, in arbitrary units. Elemental analyses were performed using an elementar Analysensysteme GmbH-Vario EL III Element Analyzer (Germany).

Syntheses

Ethyl(1,4,5-*triphenylimidazol-2-yl)thiobutyrate* (2). – To a solution of compound 1 (3.281 g, 10 mmol) in DMF (15 mL), NaH (10 mmol) was added, followed by the addition of ethylbromobutyrate (1.506 g, 10 mmol). The mixture was heated under reflux for 8 h, then cooled and poured onto crushed ice. The product was collected by filtration, washed with water, dried and recrystallized from ethanol.

(1,4,5-Triphenylimidazol-2-ylthio)butyric acid hydrazide (3). – A mixture of compound 2 (4.425 g, 10 mmol) and hydrazine hydrate (10 mmol) in ethanol (25 mL) was refluxed for 10 h. After cooling to room temperature, ethanol was removed under reduced pressure, and the product was recrystallized from ethanol.

4-Amino-5-[(1,4,5-triphenylimidazol-2-yl)thiopropyl]-2H-1,2,4-triazole-3-thione (4). – Carbon disulfide (1.14 g, 15 mmol) was added dropwise to a solution of compound **3** (4.281 g, 10 mmol) in absolute ethanol (30 mL) containing potassium hydroxide (0.84 g, 15 mmol) at 0 °C. The reaction mixture was stirred at room temperature for 16 h, and then cooled and diluted with diethyl ether. The precipitate was filtered, washed with diethyl ether and dried. The potassium dithiocarbazinate salt was obtained in nearly quantitative yield and was used without further purification since it was moisture sensitive.

Hydrazine hydrate (20 mmol) was added to a suspension of the potassium salt of dithiocarbazinate (10 mmol) in water (10 mL) and the mixture was refluxed with stirring for 4 h. After cooling, it was diluted with water, then acidified with aqueous hydrochloric acid. The precipitate was filtered, washed with water and recrystallized from ethanol to give yellow needles.

1-[4-(1,4,5-Triphenylimidazol-2-ylthio)butanoyl]-4-phenylthiosemicarbazides (5–7). General method. – A mixture of compound **3** (4.281 g, 10 mmol) and the appropriate isothiocyanate derivative (10 mmol) was refluxed in ethanol for 6 h. The solution was cooled and a white solid appeared. The obtained precipitate was filtered and recrystallized from ethanol to afford the desired product.

4-Alkyl/aryl-5-[(1,4,5-triphenylimidazol-2-yl)thiopropyl]-2H-1,2,4-triazole-3-thione (8–10). General method. – A solution of the corresponding thiosemicarbazide 5–7 (10 mmol) in 100 mL of 4 mol L⁻¹ NaOH was refluxed for 8 h. The resulting solution was cooled to room temperature and acidified with 37 % HCl. The precipitate formed was filtered, washed with water and recrystallized from ethanol to afford the desired product.

5-[(1,4,5-Triphenylimidazol-2-yl)thiopropyl]-2-(N-alkyl/arylamino)-1,3,4-thiadiazoles (11–13). General method. – A mixture of the corresponding thiosemicarbazide 5–7 (10 mmol) in cold concentrated sulfuric acid (20 mL) was stirred for 30 min. The mixture was then allowed to cool to room temperature. After stirring for additional 3 h, the resulting solution was poured into ice-cold water and made alkaline to pH 8 with ammonia. The precipitated product was filtered and recrystallized from ethanol to afford the desired product.

4-[(1,4,5-Triphenylimidazol-2-yl)thio]-N'-(arylidene)-butanehydrazides (**14–18**). General method. – A mixture of compound **3** (0.428 g, 1 mmol) and the appropriate aromatic aldehyde (1 mmol) in ethanol (20 mL) and (0.5 mL) of HCl (37 %) was refluxed for 4–6 h. After cooling, the obtained product was collected and recrystallized from ethanol.

Physical, analytical data and spectral data (MS, IR, ¹H- and ¹³C-) for the newly synthesized products **2–18** are collected in Tables I and II. Elemental analyses of all compounds were within \pm 0.4 % of the theoretical values.

Antimicrobial susceptibility testing

The clinical isolate tested in this study were obtained from the culture collection maintained at the RCMB (Regional Center for Mycology and Biotechnology/Antimicrobial unit test organisms), Al Azhar University, Cairo-Egypt. All compounds were tested *in vitro*

	M.p. (°C)/	Theor. (M+H)	(Calcd./found (%)
Compd.	yield (%)	Exp. MS : [M+H] ⁺	С	Н	Ν
2	115–116 (85)	443.18 443.18	73.27/73.03	5.92/5.69	6.33/6.23
3	169–170 (82)	429.18 429.18	70.07/69.84	5.64/5.35	13.07/13.19
4	263–264 (77)	485.18 485.18	64.43/64.25	4.99/5.16	17.34/17.13
5	238–239 (90)	564.19 564.19	68.18/67.99	5.19/5.01	12.42/12.30
6	190–191 (83)	516.19 516.19	65.21/65.47	5.67/5.39	13.58/13.34
7	222–223 (80)	502.17 502.17	64.64/64.97	5.42/5.14	13.96/13.71
8	261–262 (84)	546.18 546.18	70.43/70.31	4.99/5.12	12.83/12.56
9	233–234 (79)	498.18 498.18	67.57/67.42	5.47/5.26	14.07/13.76
10	250–251 (82)	484.16 484.16	67.05/66.79	5.21/5.49	14.48/14.76
11	279–280 (74)	546.18 546.18	70.43/70.17	4.99/5.29	12.83/12.99
12	247–248 (70)	498.18 498.18	67.57/67.91	5.47/5.70	14.07/14.38
13	262–263 (73)	484.16 484.16	67.05/67.29	5.21/5.44	14.48/14.16
14	225–226 (87)	533.20 533.20	72.16/72.45	5.30/5.19	10.52/10.21
15	196–197 (85)	551.17 551.17	69.74 /69.39	4.94/5.24	10.17/10.26
16	179–180 (90)	551.17 551.16	69.74/69.95	4.94/5.09	10.17/10.38
17	220–221 (82)	595.11 595.11	64.54/64.87	4.57/4.74	9.41/9.65
18	245–246 (86)	595.12 595.11	64.54/64.78	4.57/4.67	9.41/9.76

Table I. Physical and analytical data for the newly synthesized compounds 2-18

Compd.	IR (KBr, $v_{max'}$ cm ⁻¹)	¹ H NMR (δ , ppm)	¹³ C NMR (<i>δ</i> , ppm)
2	1740 (C=O), 1637 (C=N), 1205 (C-O)	1.95–1.98 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.41 (t, 2H, CH ₂ CO), 3.16 (t, 2H, SCH ₂), 4.02–4.05 (q, 2H, OCH ₂), 7.15–7.45 (m, 15H, H-Ar)	14.09 (CH ₂ CH ₃), 24.61 (CH ₂ CH ₂ CH ₂), 31.72 (CH ₂ CO), 32.22 (SCH ₂), 59.88 (OCH ₂), 126.19, 126.5, 128.17, 128.28, 128.35, 128.92, 129.13, 130.12, 130.72, 131.04, 134.17, 135.36, 137.35, 142.35 (C-Ar, C=N)
3	3256–3380 (NH, NH ₂), 1692 (C=O), 1630 (C=N)	1.91–1.96 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.14 (t, 2H, CH ₂ CO), 3.14 (t, 2H, SCH ₂), 4.16 (s, 2H, NH ₂), 7.12–7.44 (m, 15H, <i>H</i> -Ar), 9.00 (s, 1H, NH)	25.33 (CH ₂ CH ₂ CH ₂), 32.01 (CH ₂ CO), 32.25 (SCH ₂), 126.22, 126.47, 128.16, 128.28, 128.32, 128.51, 128.90, 129.13, 130.13, 130.73, 130.98, 134.18, 135.37, 137.33, 142.51 (C-Ar, C=N), 170.89 (C=O)
4	3240–3367 (NH, NH ₂), 1643 (C=N), 1290 (C=S)	2.04–2.11 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.82 (t, 2H, CH ₂ C=N), 3.27 (t, 2H, SCH ₂), 5.76 (s, 2H, NH ₂), 7.22–7.46 (m, 15H, H-Ar), 14.40 (s, 1H, NH)	23.58 (CH ₂ CH ₂ CH ₂), 25.10 (CH ₂ C=N), 32.11 (SCH ₂), 126.80, 126.91, 127.54, 128.22, 128.42, 128.59, 128.91, 129.24, 129.53, 130.76, 131.57, 131.57, 134.57, 141.90, 142.08, 163.38 (C-Ar, C=N), 177.64 (C=S)
5	3210–3352 (NH), 1670 (C=O), 1642 (C=N), 1302 (C=S)	1.96–2.03 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.32 (t, 2H, CH ₂ CO), 3.21 (t, 2H, SCH ₂), 7.12–7.47 (m, 20H, <i>H</i> -Ar), 9.57 (d, 1H, NH), 9.62 (s, 1H, NH), 9.90 (d, 1H, NH)	24.64 (CH ₂ CH ₂ CH ₂), 31.95 (CH ₂ CO), 32.12 (SCH ₂), 126.20, 126.45, 128.13, 128.24, 128.30, 128.49, 128.89, 129.12, 130.06, 130.68, 130.95, 134.13, 135.31, 137.31, 142.49 (C-Ar, C=N), 173.66 (C=O)
6	3240–3367 (NH), 1689 (C=O), 1634 (C=N), 1290 (C=S)	1.03 (t, 3H, CH ₂ CH ₃), 1.91–1.98 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.27 (t, 2H, CH ₂ CO), 3.18 (t, 2H, SCH ₂), 3.36–3.42 (m, 2H, NCH ₂), 7.16–7.45 (m, 15H, H-Ar), 7.91 (t, 1H, NH), 9.06 (d, 1H, NH), 9.65 (s, 1H, NH)	14.45 (CH ₂ CH ₃), 24.60 (CH ₂ CH ₂ CH ₂), 31.92 (CH ₂ CO), 32.03 (SCH ₂), 38.35 (NCH ₂), 126.19, 126.45, 128.13, 128.23, 128.31, 128.49, 128.90, 129.12, 130.05, 130.68, 130.95, 134.11, 135.31, 137.30, 142.48 (C-Ar, C=N), 171.31 (C=O)
7	3234–3380 (NH), 1663 (C=O), 1650 (C=N), 1303 (C=S)	1.92–1.99 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.27 (t, 2H, CH ₂ CO), 2.84 (d, 3H, NCH ₃), 3.18 (t, 2H, SCH ₂), 7.14–7.45 (m, 15H, H-Ar), 7.88 (d, 1H, NH), 9.12 (d, 1H, NH), 9.67 (s, 1H, NH)	24.55 (CH ₂ CH ₂ CH ₂), 30.81 (NCH ₃), 31.91 (CH ₂ CO), 32.01 (SCH ₂), 126.20, 126.46, 128.14, 128.24, 128.30, 128.50, 128.90, 129.13, 130.04, 130.67, 130.95, 134.11, 135.30, 137.30, 142.49 (C -Ar, C=N), 171.37 (C=O)
8	3224–3290 (NH), 1697 (C=N), 1282 (C=S)	1.92–1.99 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.53 (t, 2H, CH ₂ C=N), 3.12 (t, 2H, SCH ₂), 7.14–7.52 (m, 20H, <i>H</i> -Ar), 13.74 (s, 1H, NH)	24.03 (CH ₂ CH ₂ CH ₂), 25.19 (CH ₂ C=N), 31.34 (SCH ₂), 126.15, 126.43, 128.15, 128.22, 128.30, 128.50, 128.85, 129.07, 129.34, 130.04, 130.94, 133.64, 134.07, 135.25, 137.29, 142.13, 151.54, 167.65 (C-Ar, C=N)

Table II. IR, ¹H-, and ¹³C NMR spectral data for the newly synthesized compounds 2–18

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			13 C N M (S)
Compd.	IR (KBr, $\nu_{max'}$ cm ⁻¹)	¹ H NMR (δ , ppm)	¹³ C NMR (δ , ppm)
9	3265–3305 (NH), 1694 (C=N), 1305 (C=S)	1.15 (t, 3H, CH ₂ CH ₃), 2.08–2.15 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.80 (t, 2H, CH ₂ C=N), 3.27 (t, 2H, SCH ₂), 3.89–3.94 (q, 2H, NCH ₂), 7.17–7.43 (m, 15H, <i>H</i> -Ar), 13.56 (s, 1H, NH)	13.37 (CH ₂ CH ₃), 23.27 (CH ₂ CH ₂ CH ₂), 25.56 (CH ₂ C=N), 31.90 (SCH ₂), 48.56 (NCH ₂), 126.36, 126.82, 128.23, 128.53, 129.13, 129.56, 130.60, 130.71, 131.23, 131.76, 134.08, 135.66, 138.12, 142.09, 151.90, 166.58 (C- Ar, C=N)
10	3220–3279 (NH), 1642 (C=N), 1295 (C=S)	2.07–2.14 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.50 (s, 3H, NCH ₃), 2.78 (t, 2H, CH ₂ C=N), 3.21 (t, 2H, SCH ₂), 7.14–7.45 (m, 15H, H-Ar), 13.52 (s, 1H, NH)	23.60 (CH ₂ CH ₂ CH ₂), 25.16 (NCH ₃), 29.62 (CH ₂ C=N), 31.53 (SCH ₂), 126.10, 126.44, 128.18, 128.33, 128.50, 128.67, 129.09, 130.04, 130.69, 131.03, 134.05, 135.29, 137.28, 142.20, 151.90, 166.58 (C-Ar, C=N)
11	3254–3298 (NH), 1634 (C=N)	2.03–2.11 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.99 (t, 2H, CH ₂ C=N), 3.24 (t, 2H, SCH ₂), 7.21–7.59 (m, 20H, <i>H</i> -Ar), 10.48 (s, 1H, NH)	27.81 (CH ₂ CH ₂ CH ₂), 28.70 (CH ₂ C=N), 33.11 (SCH ₂), 116.22, 125.86, 126.56, 126.85, 127.23, 128.23, 128.69, 129.36, 130.04, 130.73, 132.30, 134.03, 140.68, 141.50, 141.96, 147.98, 158.75, 164.02 (C-Ar, C=N)
12	3213–3296 (NH), 1637 (C=N)	1.20 (t, 3H, CH ₂ CH ₃), 2.04–2.11 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.97 (t, 2H, CH ₂ C=N), 3.24 (t, 2H, SCH ₂), 3.33–3.42 (q, 2H, NCH ₂), 7.19–7.54 (m, 15H, H-Ar), 9.96 (s, 1H, NH)	13.34 (CH ₂ CH ₃), 27.78 (CH ₂ CH ₂ CH ₂), 28.16 (CH ₂ C=N), 32.17 (SCH ₂), 40.83 (NCH ₂), 125.96, 126.33, 128.23, 128.64, 129.01, 129.30, 129.65, 130.65, 131.03, 131.93, 134.33, 142.11, 147.28, 157.74, 167.74 (C-Ar, C=N)
13	3207–3231 (NH), 1641 (C=N)	2.05–2.12 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.96–2.99 (m, 5H, CH ₂ C=N, NCH ₃), 3.24 (t, 2H, SCH ₂), 7.19–7.53 (m, 15H, <i>H</i> -Ar), 9.90 (s, 1H, NH)	27.79 (CH ₂ CH ₂ CH ₂), 28.17 (CH ₂ C=N), 32.07 (SCH ₂), 38.83 (NCH ₃), 125.67, 126.25, 128.23, 128.64, 128.97, 129.29, 129.62, 130.64, 131.22, 131.88, 134.52, 142.23, 146.77, 157.85, 168.75 (C- Ar, C=N)
14	3254–3342 (NH), 1704 (C=O), 1672 (C=N)	$\begin{array}{l} 1.95-2.03 \; (quin, 2H, \\ CH_2CH_2CH_2), 2.36 \; (t, 0.8H, \\ CH_2CO), 2.76 \; (t, 1.2H, CH_2CO), \\ 3.17-3.23 \; (m, 2H, SCH_2), \\ 7.14-8.07 \; (m, 20H, H-Ar, OH), \\ 8.10 \; (s, 0.25H, H-C=N), 8.31 \; (s, \\ 0.25H, H-C=N), 8.49 \; (s, 0.15H, \\ H-C=N), 8.70 \; (s, 0.4H, H-C=N), \\ 11.37 \; (s, 0.35H, CONH), 11.49 \; (s, 0.25H, CONH), 11.54 \; (s, \\ 0.25H, CONH), 11.67 \; (s, 0.15H, \\ CONH) \end{array}$	24.24, 24.89 (CH ₂ CH ₂ CH ₂), 30.56, 31.76 (CH ₂ CO), 31.88, 32.09 (SCH ₂), 122.17, 123.11, 125.70, 126.13, 126.17, 126.41, 127.26, 128.10, 128.46, 128.84, 129.07, 130.65, 130.92, 131.13, 132.94, 134.11, 135.35, 135.96, 137.29, 140.88, 144.04, 160.56 (C- Ar, C=N), 168.18, 173.82 (C=O)

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Compd.	IR (KBr, $v_{max'}$ cm ⁻¹)	¹ H NMR (δ , ppm)	¹³ C NMR (δ , ppm)
15	3260–3322 (NH), 1698 (C=O), 1645 (C=N)	1.99–2.07 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.39 (t, 0.8H, CH ₂ CO), 2.72 (t, 1.2H, CH ₂ CO), 3.17–3.26 (m, 2H, SCH ₂), 6.80–7.46 (m, 19H, <i>H</i> -Ar), 8.26 (s, 0.40H, <i>H</i> -C=N), 8.34 (s, 0.60H, <i>H</i> -C=N), 11.30 (s, 0.60H, CONH), 11.67 (s, 0.40H, CONH)	24.21, 24.55 (CH ₂ CH ₂ CH ₂), 30.58, 31.91 (CH ₂ CO), 32.35, 32.51 (SCH ₂), 116.06, 116.29, 118.15, 118.56, 119.20, 119.59, 126.41, 126.58, 126.88, 128.50, 129.13, 129.55, 130.80, 131.14, 133.23, 135.05, 136.49, 140.77, 142.38, 156.31, 158.60 (C-Ar, C=N), 167.74, 172.75 (C=O)
16	3275–3330 (NH), 1690 (C=O), 1640 (C=N)	1.99–2.02 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.38 (t, 0.8H, CH ₂ CO), 2.73 (t, 1.2H, CH ₂ CO), 3.19, 3.21 (2t, 2H, SCH ₂), 7.13–7.93 (m, 19H, <i>H</i> -Ar), 8.11 (s, 0.40H, <i>H</i> -C=N), 8.72 (s, 0.60H, <i>H</i> -C=N), 11.38 (s, 0.60H, CONH), 11.56 (s, 0.40H, CONH)	24.26 (CH ₂ CH ₂ CH ₂), 30.86, 31.90 (CH ₂ CO), 31.88, 32.10 (SCH ₂), 126.13, 126.90, 127.83, 128.09, 128.21, 128.46, 129.05, 130.65, 130.89, 131.16, 134.10, 137.29, 142.34, 160.56 (C-Ar, C=N), 160.61, 173.84 (C=O)
17	3206-3287 (NH), 1700 (C=O), 1692 (C=N)	1.99–2.04 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.37 (t, 0.8H, CH ₂ CO), 2.76 (t, 1.2H, CH ₂ CO), 3.18–3.25 (m, 2H, SCH ₂), 7.16–8.50 (m, 19H, H-Ar), 8.71 (s, 0.40H, H-C=N), 8.90 (s, 0.60H, H-C=N), 11.39 (s, 0.60H, CONH), 11.77 (s, 0.40H, CONH)	24.28, 24.89 (CH ₂ CH ₂ CH ₂), 30.55, 30.65 (CH ₂ CO), 32.12, 32.22 (SCH ₂), 125.51, 126.03, 126.63, 128.12, 128.21, 128.74, 130.40, 130.66, 131.23, 132.27, 132.58, 133.13, 133.63, 133.93, 134.16, 140.71, 143.89, 159.68, 160.53 (C -Ar, C=N), 161.87, 167.34 (C=O)
18	3206–3287 (NH), 1700 (C=O), 1692 (C=N)	2.00–2.04 (quin, 2H, CH ₂ CH ₂ CH ₂), 2.30 (t, 0.8H, CH ₂ CO), 2.78 (t, 1.2H, CH ₂ CO), 3.11–3.23 (m, 2H, SCH ₂), 7.14–7.89 (m, 19H, <i>H</i> -Ar), 8.31 (s, 0.60H, <i>H</i> -C=N), 8.49 (s, 0.40H, <i>H</i> -C=N), 11.50 (s, 0.60H, CONH), 11.62 (s, 0.40H, CONH)	24.29, 24.85 (CH ₂ CH ₂ CH ₂), 30.68, 31.90 (CH ₂ CO), 32.06, 32.86 (SCH ₂), 123.12, 123.36, 126.14, 126.18, 126.90, 127.96, 128.10, 128.24, 128.82, 129.05, 130.08, 130.91, 131.27, 131.55, 132.96, 133.08, 134.12, 135.35, 140.92, 142.41, 142.46, 144.06 (C -Ar, C=N), 168.13, 173.83 (C=O)

for their antibacterial activity against the following clinical bacterial strains: Gram negative bacteria, *Escherichia coli* (RCMB 0100520, *Pseudomonas aeruginosa* (RCMB 010043), and Gram positive bacteria, *Streptococcus pneumonia* (RCMB 010010), *Bacillus subtilis* (RCMB 010067) by the agar diffusion method (25) using Müller-Hinton agar medium for bacteria.

Antimicrobial susceptibility testing was performed by the agar disc-diffusion method. Sterile filter paper discs (8 mm diameter) were moistened with the compound solution in dimethylsulphoxide of specific concentration. Forty μ L of the solution containing 200 μ g of test compound or reference drug (gentamicin or ampicillin trihydrate) was loaded onto filter paper discs. Solvent was evaporated and discs were transferred to the inocu-

lated plates. Loaded discs were carefully placed on Müller-Hinton for bacteria and Sabouraud's agar for fungi, previously inoculated separately with the test microorganisms. The plates were incubated at 37 °C for bacteria and 28 °C for fungi, diameters of the growth inhibition zones were measured after 24 hours in case of bacteria and after 48–96 hours in case of fungi. The antimicrobial activity was evaluated by measuring the zone of inhibition against the tested organisms and compared with that of the standards. Antimicrobial activities were expressed as the inhibition diameter zones in millimeters (mm) and are presented in Tables III and IV. Each experiment was carried out in triplicate and the average zone of inhibition was calculated.

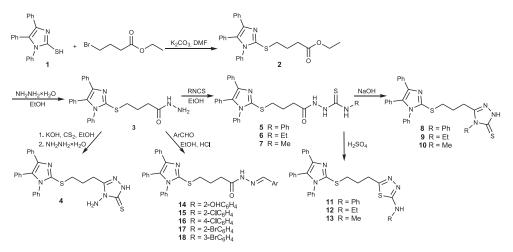
Moreover, compounds **2–18** were also subjected to *in vitro* testing of antifungal activity against the following four clinical fungal strains: *Aspergillus fumigatus* (RCMB 02568), *Syncephalastrum racemosum* (RCMB 05922), *Geotrichum candidum* (RCMB 05097), and *Candida albicans* (RCMB 05031) by agar diffusion method (26) using Sabouraud's agar medium for fungi.

	Mean value of inhibition zone diameter (mm)			
Compound	Gram positive bacteria		Gram negative bacteria	
-	S. pneumoniae	B. subtilis	P. aeruginosa	E. coli
2	13.0	15.3	NA	12.3
3	13.9	16.0	NA	13.2
4	19.4	24.6	NA	16.3
5	19.8	22.4	NA	14.2
6	17.2	18.3	NA	13.7
7	12.3	14.2	NA	NA
8	19.4	21.3	NA	12.7
9	18.1	19.6	NA	11.9
10	10.3	12.4	NA	NA
11	19.1	20.8	NA	14.9
12	17.3	18.7	NA	11.0
13	17.9	19.0	NA	12.7
14	20.6	25.4	NA	17.2
15	14.4	18.2	NA	13.9
16	13.8	16.5	NA	10.2
17	18.2	19.4	NA	15.6
18	13.7	15.0	NA	10.4
Ampicillin	20.8	26.7	-	-
Gentamicin	-	_	16.1	18.3

Table III. Antibacterial activity of compounds 2-18 against Gram negative and Gram positive bacteria

C 1	Mean value of inhibition zone diameter (mm)				
Compound -	A. fumigatus	S. racemosum	G. candidum	C. albicans	
2	14.3	NA	16.1	13.6	
3	14.9	NA	16.8	14.2	
4	19.3	NA	23.3	18.3	
5	18.6	NA	20.6	18.7	
6	16.7	NA	17.8	15.1	
7	NA	NA	NA	NA	
8	17.4	NA	18.3	16.4	
9	16.8	NA	18.3	15.9	
10	NA	NA	NA	NA	
11	19.4	NA	20.6	18.2	
12	15.2	NA	16.4	15.8	
13	15.6	NA	17.8	16.3	
14	20.1	NA	25.8	21.6	
15	19.3	NA	20.3	18.3	
16	13.2	NA	14.3	11.1	
17	17.8	NA	15.8	14.7	
18	13.9	NA	16.2	13.4	
Amphotericin B	20.4	17.3	26.3	22.0	

Table IV. Antifungal activity of compounds 1–7 against some fungi





RESULTS AND DISCUSSION

Chemistry

Synthesis of the title azole compounds required stepwise reactions starting from 1,4,5-triphenylimidazole-2-thiol (1), as illustrated in Schemes 1–3. The starting compound 1 was prepared according to the literature (27).

Alkylation of compound **1** with 4-ethylbromobutyrate in dimethylformamide in the presence of sodium hydride as a base gave ethyl(1,4,5-triphenylimidazol-2-yl)thiobutyrate (**2**) in 85 % yield. Hydrazinolysis of ester **2** with an equimolar amount of hydrazine hydrate furnished the corresponding acid hydrazide **3** (Scheme 1).

All the newly synthesized compounds were characterized by MS, IR, ¹H and ¹³C NMR spectra. The structure of compound **2** was in accord with its spectroscopic data. The IR spectrum of compound **2** showed the principal absorption band at 1740 cm⁻¹, indicating the presence of a carbonyl group (C=O) in the molecule, which confirms the success of the alkylation reaction. In the ¹H NMR spectrum, the characteristic ester protons appeared as a triplet at δ 1.15 ppm and a quartet at δ 4.02–4.05 ppm. The other methylene groups were observed as quintets ranging from δ 1.95–1.98 ppm, triplets at δ 2.41 ppm and triplets at δ 3.16 ppm, according to CH₂CH₂CH₂, CH₂CO and SCH₂, respectively. All aromatic protons were observed at their usual chemical shifts with δ 7.15–7.45 ppm. In the ¹³C NMR spectrum, one methyl carbon, was observed at δ 14.09 ppm and the carbons of the four methylene groups at δ 24.61, 31.72, 32.22 and 59.88 ppm, respectively (see Table II). All aromatic carbons of the three phenyl groups and C=N gave signals between δ 126.19–142.35 ppm. In addition, the carbon of the carbonyl group appeared at δ 172.38 ppm.

The IR spectrum of acid hydrazide **3** showed absorption bands at 3256–3380 cm⁻¹ corresponding to the NH and NH₂ groups of the acid hydrazide, which confirms the success of the hydrazinolysis reaction. The ¹H NMR spectrum of **3** showed a quintet around δ 1.91–1.96 ppm due to the methylene group CH₂*CH*₂CH₂. The remaining methylene groups of *CH*₂CO and S*CH*₂ showed triplets at δ 2.14 and 3.14 ppm, respectively. In addition, NH₂ and NH groups were assigned to two singlets appearing at δ 4.16 and 9.00 ppm, respectively. The structure of **3** was also illustrated from its ¹³C NMR spectrum, which showed three carbons of the methylene groups at δ 25.33, 32.01 and 32.25 ppm, while the remaining aromatic carbons and C=N appeared in the chemical shift range of δ 126.22–142.51 ppm. The carbon of the carbonyl group showed a signal at δ 170.89 ppm.

The treatment of acid hydrazide **3** with carbon disulphide in ethanolic potassium hydroxide solution led to the formation of potassium dithiocarbazinate salt. Its treatment with hydrazine hydrate under reflux furnished the desired 4-amino-5-[(1,4,5-triphenyl-imidazol-2-yl)thiopropyl]-2*H*-1,2,4-triazole-3-thione (**4**) in 88 % yield (Scheme 1). IR spectrum of compound **4** exhibited N–H bands in the region 3240–3367 cm⁻¹. The absorption bands at 1643 and 1290 cm⁻¹ are due to the presence of C=N and C=S stretchings of the triazole ring system, respectively. The appearance of signals approximately at 5.76 ppm (NH₂) and 14.40 ppm (NH), integrating two and one protons, respectively, in the ¹H NMR spectra of compound **4** confirmed the conversion of **3** into the corresponding aminotriazole **4**. Aromatic protons were observed at 7.22–7.46 ppm. These groups were observed at 126.80–163.38 ppm in the ¹³C NMR spectrum.

On the other hand, the reaction of the same hydrazide **3** with alkyl/aryl isothiocyanate furnished the corresponding aryl/alkylthiosemicarbazide derivatives **5**–7 (Scheme 1). Their structures were established on the basis of their spectral data, which indicated the

presence of the thiocarbonyl group (C=S) by the appearance of new absorption bands at 1290–1303 cm⁻¹ in IR spectra. Their ¹H NMR spectra showed the disappearance of NH₂ protons. The CONH and NHCSNH protons were observed in the range of δ 7.88–9.90 ppm, confirming the formation of thiosemicarbazides. Their carbon signals were observed in a chemical shift range of δ 171.31–173.66 ppm.

Cyclodehydration of compounds **5–7** was performed in alkaline medium to give the corresponding 4-alkyl/aryl-5-[(1,4,5-triphenylimidazol-2-yl)thiopropyl]-2*H*-1,2,4-triazole-3-thiones (**8–10**) in yields ranging 79–84 % (Scheme 1).

The structure of 1,2,4-triazoles **8–10** displayed IR, ¹H NMR and ¹³C NMR spectra consistent with the assigned structures. In their IR spectra, the absorption peak corresponding to the C=N group was observed around 1642–1697 cm⁻¹. In their ¹H NMR spectra, the appearance of signals arising from the thiol group (SH) at δ 13.52–13.74 ppm provided evidence for the formation of triazole ring. Another support for the structure of these compounds was the disappearance of resonances arising from –CONH and –NHCSNH-around δ 7.88–9.90 ppm. The remaining protons were also observed in the expected regions (see Table II).

Ring closure of acid thiosemicarbazides **5**–7 in acidic medium is a well known method for the synthesis of 1,3,4-thiadiazoles (28). Thus, 5-[(1,4,5-triphenylimidazol-2-yl) thiopropyl]-2-(*N*-alkyl/arylamino)-1,3,4-thiadiazoles **11–13** were obtained in 70–84 % yield *via* the treatment of the above acid thiosemicarbazides **5**–7 with H₂SO₄, concentration 98 %, at 0 °C. Their IR spectra showed the absence of C=O and C=S stretching bands around 1663–1670 and 1290–1302 cm⁻¹, respectively, observed in the IR spectra of the thiosemicarbazide derivatives **5**–7. The exhibited chemical shifts obtained from their ¹H NMR spectra were all supported by the proposed structures of these thiadiazoles. Thus, the disappearance of CONH and NHCSNH signals and appearance of a sharp singlet at δ 9,90–10.48 ppm of the NH group in the ¹H NMR spectra of compounds **11–13** accounted for the thiadiazole ring formation and was reconfirmed by the ¹³C NMR peak at δ 164.02–168.75 due to C2 of thiadiazole (see Table II).

The acid hydrazide **3** was also condensed with various aromatic aldehydes in the presence of a catalytic amount of concentrated hydrochloric acid in refluxing ethanol to afford a new series of Schiff bases **14–18** (Scheme 1).

It was reported in the literature (29, 30) that compounds having azomethine linkage exhibit E/Z geometrical isomerism around -C=N double bond and can exist as *cis/trans* amide conformers (30). Moreover, hydrazones were proven exist in higher percentage in DMSO- d_6 solution in the form of geometrical E isomer.

The structures of the Schiff bases were in accord with their spectroscopic data. The IR spectra of compounds **14–17** showed the disappearance of the absorption bands of NH_2 groups and the appearance of strong absorption bands at 3254–3342 cm⁻¹ attributed to the NH. The ¹H and ¹³C NMR spectra of hydrazones **14–18** clearly showed the disappearance of the NH_2 group of hydrazide structure and appearance of additional signals in the aromatic region belonging to the aldehydic protons.

The ¹H NMR spectra of compounds **14–18** revealed the presence of two sets of signals belonging to the SCH₂ and CH₂CO groups of *cis* and *trans* conformers at 2.30–2.78 and 3.17–3.26 ppm, respectively, in a 0.4 to 0.6 ratio (Fig. 1). Moreover, the azomethine CH proton and the amidic NH proton for compounds **15–18** resonated as two sets of signals at 8.10–8.90 and 11.30–11.77 ppm, respectively, with the same ratio. On the other hand, the appearance of these protons in the ¹H NMR spectrum of compound **14** as two sets of four

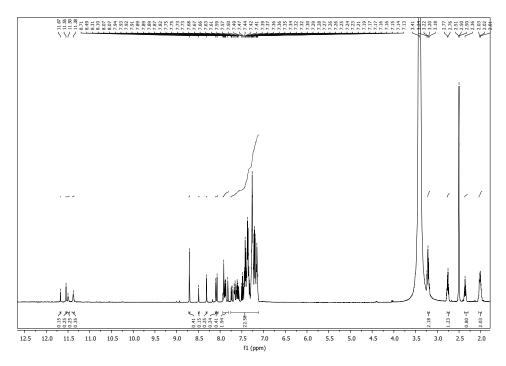


Fig. 1. ¹H NMR spectrum of Schiff base 14.

singlets can be considered as a proof of the formation of the geometrical *Z* form in both *cis* and *trans* conformation. This can be explained by an intramolecular hydrogen bonding connecting the azomethine nitrogen and the OH group of the aldehyde side chain, favoured by a stable six-member ring.

The formation of *E* and *Z* forms could be attributed to different steric rearrangements of hydrazone functionality in the geometric *syn* and *anti* isomers resulting from the hindered rotation of azomethine linkage.

Further evidence can be found by comparing the ¹H NMR spectra of hydrazide **3** with hydrazones **14–18**; no conformational equilibrium, under the same experimental conditions, was observed for hydrazide **3**.

Therefore, the NMR spectra of compounds **14–18** recorded in DMSO- d_6 solution revealed that all compounds existed as *E* geometrical isomers, except compound **14** which exhibited both *E*/*Z* geometrical isomerisms. Moreover, the *cis/trans* conformers of *E* and *Z* isomers were also observed.

Biological activity

Antibacterial activity. – Ethyl(1,4,5-triphenylimidazol-2-yl)thiobutyrate (2) and its hydrazide derivative 3 displayed good activity against *S. pneumonia* and *B. subtilis* compared

to the ampicillin reference and comparable activity against *E. coli* compared to gentamicin reference. Triazole **4**, bearing amino and thiol groups, exhibited approximately equal antimicrobial activity against all tested microorganisms except *P. aeruginosa*.

The antimicrobial activity of the series **5–7** divulged compound **5**, comprising phenyl substitution at *N*-4 exhibited the highest inhibition, whereas compounds **6** and **7** with alkyl substitution showed lower activity loss of inhibition against *E. coli* for compound **7**.

Evaluation of the antimicrobial activity of the synthesized 4-alkyl/aryl-5-[(1,4,5-triphenylimidazol-2-yl)thiopropyl]-1,2,4-triazole-3-thiols **8–10** revealed that compounds were more effective against Gram positive bacteria. In particular, *N*-4 phenyl substituted triazole **8** exhibited activity against tested Gram positive bacteria comparable to its ampicillin reference and lower inhibition against *E. coli* against compared to gentamicin. In contrast, compound **10**, comprising methyl substitution, displayed no activity against Gram negative organisms.

The mean values of inhibition zone diameters summarized in Table III show that compounds **11–13** possess congruent antibacterial activities against the growth of *S. pneumonia*, *B. subtilis* and *E. coli* compared to the standards ampicillin and gentamicin. Furthermore, *N*-phenyl thiadiazole **11** exhibited the highest antibacterial activity against all tested microorganisms, which can be attributed to the presence of the thiadiazole ring.

Among the synthesized Schiff bases **14–18**, compound **14**, having 2-OH substitution on the phenyl ring, displayed the best antibacterial activity against *S. pneumonia*, *B. subtilis* and *E. coli*. However, compounds possessing unsubstituted and halosubstituted phenyl rings exhibited good to moderate activity.

Antifungal activity

In general, most of the tested compounds were found to possess significant antifungal activity against all fungal strains except *S. racemosum*.

Compounds **2** and **3** exhibited lower antifungal activity against *A. fumigatus, G. candidum* and *C. albicans* compared to amphotericin B. The conversion of hydrazide structure 3 to 1,2,4-triazole functionalized with both thiol and amino groups caused congruent antifungal activity against *A. fumigatus, G. candidum* and *C. albicans* compared to the standard. On the other hand, 1-[4-(1,4,5-triphenylimidazol-2-ylthio)butanoyl]-4-phenyl/ethylthiosemicarbazides (**5**, **6**) displayed remarkable antifungal activity, whereas their methyl analogue 7 displayed no activity against any of the fungal strains.

Contrary to what was expected, the cyclization of compounds **5–7** to their corresponding 1,2,4-triazoles (**8–10**) caused no enhancement of the antifungal activity. On the other hand, the antifungal activity of the 1,3,4-thiadiazoles **11–13** revealed that all the tested compounds showed significant inhibition; compound **11** having phenyl substitution exhibited the highest activity. Thiadiazoles **12** and **13** showed comparatively lower activity against *A. fumigatus, G. candidum* and *C. albicans* compared to the standard.

In the studied Schiff bases series, 4-[(1,4,5-triphenylimidazol-2-yl)thio]-*N*'-(2-hydroxybenzylidene)-butanehydrazide (14) exhibited the highest antifungal activity against all fungal strains compared to amphotericin B.

CONCLUSIONS

In conclusion, a series of novel imidazole compounds bearing 1,2,4-triazole, 1,3,4-thiadiazole or Schiff base moieties were synthesized successfully *via* multistep synthesis. All the new compounds were fully spectroscopically characterized. The title compounds were synthesized as new compounds with antimicrobial activity *in vitro*.

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