

## Review article

# The genetics behind osteoarthritis: Asian focus

Rachaneekorn Tammachote

*Human Genetics Research, Department of Botany, Faculty of Science, Chulalongkorn University, Bangkok, 10330, Thailand*

---

Osteoarthritis (OA), a degenerative joint disease, is the most common form of arthritis in the elderly. Problems arising from the condition are not only health, physical, and psychological, but also economical and social. OA is a complex disease caused by environmental factors (obesity, female gender, advancing age) and genetics. A strong genetic contribution to primary OA has been observed in several studies. Asian population is undoubtedly genetically different from European, whom most studies have been performed. This review systematically compares results of molecules involved with OA from studies performed on Asian and Caucasian populations. The clear differences between Asian and Caucasian populations may be from the sex- and ethnic-specific nature of the disease, as well as from the inclusion criteria of the studies. Global collaboration is highly significant in understanding genetic underlying OA. Knowledge gained from genetic study is important to the development of prevention and therapeutic intervention, and to identify individuals at risk of developing severe or progressive OA.

**Keywords:** Asian, genetics, osteoarthritis

---

Osteoarthritis (OA), a degenerative disease of the joints, is the most common form of arthritis in the elderly. It is one of the major causes of disability in developed countries and causes physical, mental, social, and economical problems [1]. OA is characterized by degradation of articular cartilage, accompanied with sclerosis of the subchondral bone, joint space narrowing, and osteophytes (bony outgrowths at the joint margins). The affected joints include the hand, spine, knee, and hip. It can be local, which is restricted to one joint, or generalized, which involves more than three joints or a group of joints.

Known risk factors of OA are obesity, female gender, advancing age, injury to the joints, and genetics. Several epidemiological studies have demonstrated a strong genetic contribution to primary OA, with hand OA at the highest estimated heritability of 65%, while that of the knee and hip OA are 40% and 60% respectively [2]. The finding of genetics involvement with OA pathogenesis has significant

clinical importance. Firstly, the associated genes can improve knowledge on the molecular pathway underlying the disease incidence and progression. This information is necessary for development of prevention and therapeutic intervention. Secondly, physicians can identify individuals at risk of developing severe or progressive OA and allow them to monitor better the disease progression and to target the interventions.

## Approaches taken in genetic studies of osteoarthritis

Researches in the past decades have made a great step toward the understanding of genetic influences on the disease. The approaches include sib recurrent risk and familial aggregation studies, twin studies, linkage analysis, candidate gene association studies, and differential expression studies.

For the twin studies, sib recurrence risk and familial aggregation, the common phenotypes are not exclusively resulted from genetics but may be the consequence of shared environmental factors that the family members are exposed to. Therefore, large studies that include several families are very important. These studies include the Genetics, Arthrosis, and Progression study (GARP) in the

---

**Correspondence to:** Rachaneekorn Tammachote, PhD, Human Genetics Research, Department of Botany, Faculty of Science, Chulalongkorn University, Bangkok 10330, Thailand. E-mail: Rachpunyas@gmail.com

Netherlands, the Multicenter Osteoarthritis Study (MOST, of Boston University Medical Campus) and the Osteoarthritis Initiative (OAI, of University of California, San Francisco, USA).

Linkage analysis is employed to identify chromosomal regions that contain OA genes by investigating polymorphic loci that are inherited along with the trait of interest. If a polymorphic locus (marker) is close to the disease locus on the same chromosome, it is inherited jointly with the disease. With the advancement of technology, it is possible to analyze polymorphic markers of the entire genome. Studies in the United Kingdom, Finland, Iceland, and the United States (reviewed in Valdes and Spector, 2009 [3]), along with a meta-analysis [4] provided delineated regions of harboring OA susceptibility genes on chromosomes 7q34-7q36.3, 11p12-11q13.4, 6p21.1-6q15, 2q31.1-2q34, and 15q21.3-15q26.1. These regions include some candidate genes for having roles in OA incidence and progression, such as IL-1 cluster, *FRZB*, *BMP5* and *MATN3*. The roles of these genes' protein products will be discussed.

Another approach to identify OA genes is genetic associations. This strategy analyzes the effect of a gene variant on disease occurrence. If the incidence of a variant occurring in the affected group more than in the control, higher than it can be found by chance, that gene is highly possible to associate with the disease. Some genes encoding for cartilage components, extracellular matrix, and bone density, such as collagen and aggrecan, were chosen as candidate genes to be analyzed. By this approach, polymorphisms in genes encoding estrogen receptor alpha (*ESR1*) and the vitamin D receptor (*VDR*) were found to be associated with OA [5].

Another approach employs a differential expression of genes taken from cartilage and synovium of OA patients and normal control [5]. Single nucleotide polymorphisms (SNPs) in genes with significantly different expression profiles between the two groups are analyzed for the association with OA in the population. Several SNPs were identified by this method.

### Pathways and molecules involved in osteoarthritis

Several genes involving developmental processes or maintenance of cartilage and bone were found to associate with OA susceptibility and progression. Understanding the gene functions has improved

understanding towards the disease pathogenesis. Valdes and Spector [3] categorized molecules involved with OA into five broad classes. Interplay between molecules in these classes further complicates the analysis of OA pathogenesis as shown in **Fig. 1**.

### Inflammation

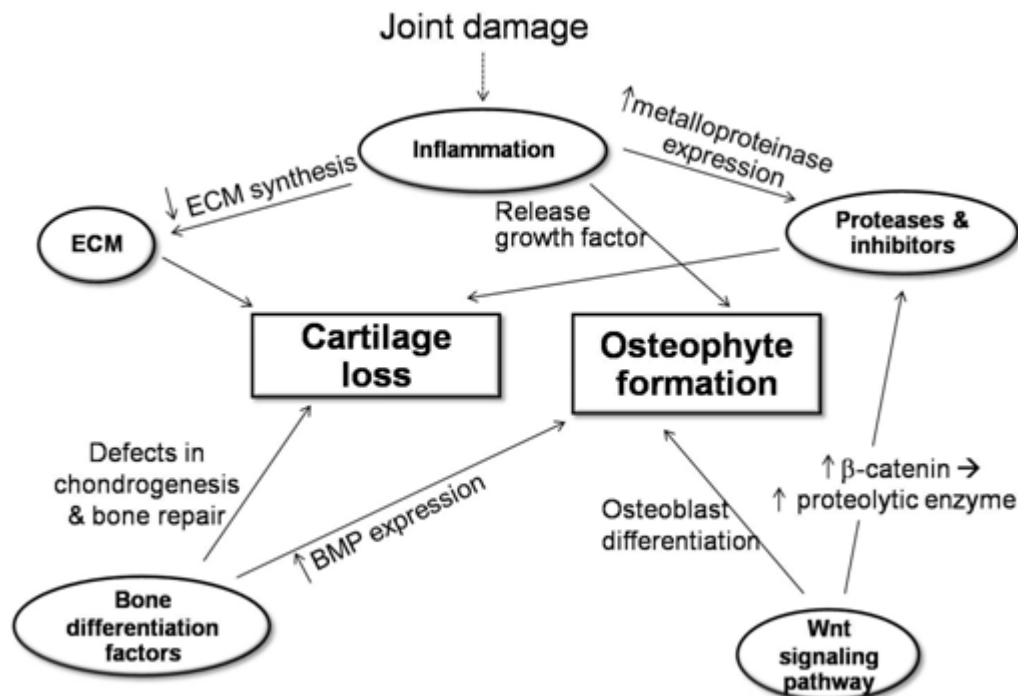
Synovial inflammation is seen in early OA [6]. Some cytokines, such as interleukin 6 (IL-6) was found to involve in the cartilage degradation. The chromosome region of 2q12-2q21 where the IL-1 gene cluster resides was also found to be involved with distal inter-phalangeal joint OA by linkage analysis [7]. Sclerotic osteoblasts express IL-6, IL-8, and TGF- $\beta$ 1 at a significantly higher level than non-sclerotic counterpart expresses, and suggested that the subchondral bone had a pro-inflammatory environment.

### Extracellular matrix (ECM) molecules

As one of OA hallmarks involves the degradation of ECM, genes encoding for these molecules, particularly collagen and cartilage oligomeric matrix protein (COMP), were excellent candidates of genetic association studies. Mutations in *COMP* cause pseudoachondroplasia and multiple epiphyseal dysplasias. Recently, COMP serum level was found to be promising as a biomarker for OA [8], but so far no polymorphism in *COMP* was found to be associated with OA [9, 10].

Similar to *COMP*, *COL2A1* had been a focus of many association studies. *COL2A1* encodes for the alpha 1 polypeptide chain of type II collagen, the major component of articular cartilage. It has increased expression in early and late-stage OA [11], while chondroprogenitor cells are normally expressed during development. Polymorphisms in the gene were found to be associated with knee and hip OA in Caucasian and with hip OA in Japanese [10]. However, no such association was found in cohorts from the US [12] and Finland (with primary early-onset hip and/or knee OA) [13].

Along with the type II collagen, aggrecan is also the major component of the cartilage ECM. To add another layer of complexity into the effect of genetics on OA, polymorphisms in genes controlling the expression of *COL2A1* and the aggrecan gene *AGC1* are also linked to the disease. A Japanese group demonstrated that a variant of *ASPN* gene, encoding for asporin, with a polymorphic stretch of 14 aspartic



ECM = Extra Cellular Matrix  
BMP = Bone Morphogenic Protein  
Wnt = Wingless-type

**Fig. 1** Interaction between molecular pathways associated with OA pathophysiology. Gene/proteins associate with OA phenotypes (in rectangular boxes) are categorized into five classes (in oval boxes). Interconnection between these pathways further complicates studies in OA pathogenesis. Inflammation that is caused by imbalance between the pro- and anti-inflammatory factors increases metalloproteinase expression. The imbalance between the proteolytic enzymes (e.g. matrix metalloproteinase, a disintegrin and metalloproteinase domains) and their inhibitors, in turn, causes cartilage degradation. Inflammation also causes cartilage loss by inhibiting ECM synthesis and chondrocyte proliferation. Genes encoding for ECM molecules and bone differentiation factors directly regulate cartilage homeostasis. Uncontrolled cell proliferation and maturation lead to another OA hallmark, osteophytes. Inflammation, the villain in OA pathogenesis, activates releases of growth factors such as Vascular Endothelial Growth Factor (VEGF) and causes angiogenesis that contributes to osteophyte formation.  $\beta$ -catenin level, controlled by Wnt pathway, also effects VEGF expression, matrix mineralization, and matrix degrading enzyme, and is involved in OA phenotype development.

acid residues (D14) within exon 2 is more common in patients with the knee and hip OA than the unaffected [14]. The investigators also reported that the variant with 13 repeats of aspartic acid residues (D13) was under-represented in the knee and hip OA patients. It has been shown that transforming growth factor (TGF)- $\beta$  induces transcription of *AGC1* and *COL2A1*. D14 asporin inhibits TGF- $\beta$ , and subsequently lowers the synthesis of aggrecan and type II collagen, whereas the D13 asporin has a weaker inhibitory effect to TGF- $\beta$  [14].

Another gene controlling *AGC1* and *COL2A1* expression is also found to be associated with OA susceptibility. *CALM1*, encoding for calmodulin 1, was identified as a hip OA susceptibility loci in Japanese cohort by a genome-wide association analysis [15]. The T allele at the core promoter SNP of *CALM1* lowers the synthesis of aggrecan and type II collagen. A combinatorial effect of polymorphisms of calmodulin1 and asporin in regulating the expression of *AGC1* and *COL2A1* was also demonstrated. Taken together, it is highly possible that genes encoding for ECMs are associated to OA.

### ***Wnt signaling proteins***

Wnt signaling pathway is involved in cartilage and bone development and degeneration (reviewed in detail in Corr, 2008 [16]. It functions in developmental processes and organogenesis. Wnt signaling molecule inhibits the transcription activity of  $\beta$ -catenin, which was suggested to contribute to cartilage loss, as areas of degenerative cartilage in OA was found to have increased levels of  $\beta$ -catenin [17]. This pathway also plays role in endochondral ossification that causes osteophytes [18]. As bone growth in OA is characterized by osteophyte and subchondral sclerosis, re-activation of Wnt pathway fits with the theory that unbalanced growth processes is a cause of OA.

Polymorphisms associated with OA susceptibility were found in some Wnt-signaling genes: Two non-synonymous SNPs in *FRZB*, encoding for secreted frizzled-related protein 3 (sFRP3), were found to associate with hip and knee OA in Caucasian, predominantly in female [19]. The association was confirmed by subsequent replication studies [10]. sFRP3 is expressed in adult articular chondrocytes. Functional analysis of the two SNPs in *FRZB* suggested that the Arg324Gly substitution and the Arg200Trp/Arg324Gly double substitution reduced the ability of sFRP3 to antagonize Wnt signaling [19].

Moreover, a SNP in the 3' untranslated region of Wnt-1-induced secreted protein 1 (*WISP-1*) gene was associated with spinal OA in postmenopausal Japanese women [20], suggesting site-specific and sex-specific effects of the pathway.

### ***Proteins related to modulation of osteocyte or chondrocyte differentiation***

Proteins in this class, such as estrogen receptor alpha (ESR1), the vitamin D receptor (VDR) and osteoprotegerin (OPG), are related to bone density and osteoporosis. Therefore, they are good candidates for genetic association studies. In fact, several variants in genes encoding for these proteins have been reported to be associated with OA in several populations [5, 21-26]. While some negative findings have also been reported [12, 27-29].

Bone morphogenetic proteins (BMPs) are a group of growth factor and cytokine in the TGF- $\beta$  superfamily. Microsatellite in intron 1 of *BMP5* and SNPs in *BMP2* were suggested to be associated with OA [5, 26, 30]. Protein encoded from the *growth and differentiation factor 5* (*GDF5*) gene belongs to the BMP family and functions in chondrogenesis and bone

repair. A SNP in the 5' untranslated region of *GDF5* has been reported to associate with OA in several populations [31, 32] and to cause significant decrease in transcriptional activity [31].

It was found that the BMP-7 level was elevated in plasma and synovium fluid of Thai OA patients [33]. BMP-7 is under investigation for being a potential inhibitor of OA progression [34].

The thyroid pathway was also implicated in OA at multiple joints. *DIO2* gene encoding for an enzyme converting T4 (inactive thyroid) into T3 (active thyroid) was found to associate with the disease in UK, Dutch, and Japanese [35]. T3 in the growth plate functions in chondrocyte differentiation and bone formation.

### ***Proteases and their inhibitors***

Proteins in this class include A disintegrin and metalloproteinase domain (ADAM) and ADAM with thrombospondin motif (ADAMTS). ADAM and ADAMTS are metalloproteinase involved in osteoclast formation, chondrocytes maturation, and proliferation. Polymorphisms in *ADAM12* were reported to associate with prevalence and progression of knee OA [5, 26, 36]. While Rodriguez-Lopez (2009) found no association of these polymorphisms with OA, they found a SNP in *ADAMTS14* associated with the knee OA in women [37].

Moreover, levels of some matrix metalloproteinases (MMPs) are elevated in synovial fluid of OA patients and a polymorphism in MMP-1 was found to associate with knee OA in Turkish [38]. MMPs and aggrecanases involve in chondrocyte maturation by degrading the cartilage matrix, to be filled with bone-specific collagen. MMP-3 interacts with sFRP3 of the wnt signaling pathway and MMP-3 level is increased in *Frzb* knockout mice [39].

In addition to genes belonging to these five categories, epigenetic control, such as DNA methylation, may be important in determining complex gene expression pattern found in arthritic chondrocytes [40]. Methylation of *leptin*, a gene involved in obesity, directly effects the expression of MMP-13 [41]. Some microRNAs, whose function is to silence their target genes, were identified to be differentially expressed in osteoarthritic chondrocytes compared with normal chondrocytes [42].

### ***Site-specific, sex-specific, and ethnic-specific nature of osteoarthritis genetic effects***

It is obvious that some genes and variants are



controversial on being related to OA. The reason may be that effects of genetic variations on OA are mostly site-specific and some are ethnic- and sex-specific (**Table 1**). For example, haplotypes in genes encoding for the cartilage intermediate layer protein (CILP) and ADAM12 were found to be associated with knee OA, especially in men [5]. Moreover, while a variation in the promoter of the calmodulin gene, *CALM1*, was reported to be associated with hip OA in Japanese [15], studies in Han Chinese and Caucasian were not able to replicate the results [10, 43, 44]. Another variation in *GDF5* was found to be associated with susceptibility to OA in Japanese and Chinese [31], and Europeans [32], but not in Greek [45]. Similarly, genetic predisposition of rheumatoid arthritis is difference between population of European and Asian ancestries [46]. The ethnic-specific nature of the findings may not exclusively due to genetic factors, as shared environmental factors are important in the disease incidence and progression.

#### **Challenges in genetic studies of osteoarthritis**

There are a few challenges in studies of genetic effects on OA, in addition to the disease being multifactorial, which is the interplay between several genes and environmental factors.

#### **Lack of good animal models**

Conducting experiment in animal model is the gold standard in functional study of disease-causing genes. However, thus far, there is no “ideal” model for OA genetic research. For example, articular cartilages of mouse and human have considerable differences, both in cartilage metabolism and histology. The reason impedes the functional analysis of *ADAMTS5*, a gene encoding for major aggrecanase in articular cartilage, was proved involved in OA as deletion of this gene prevents cartilage degradation in a mouse model of OA [47]. Moreover, *cystatin 10*, a gene implicated in OA in a mouse model, has no human counterpart. This challenge makes it difficult to analyze functionally the role of susceptibility genes.

#### **Low rate of replication**

This problem is of particular for studies that include different ethnic groups. A standard inclusion criterion is crucial for a global collaboration. Some countries, like Thailand, have population from mixed ethnic background. Thus, it is one of the major causes of the low rate of replication.

#### **Small sample size in specific studies**

This problem arises from OA being site- (e.g., hip, hand, and knee), sex-, and ethnic-specific. Therefore, it is difficult to recruit enough cases and controls for each subgroup to conduct reliably the studies.

#### **Inclusion criteria**

While some studies used radiographic OA (ROA) as an inclusion criterion, some used symptomatic OA with radiographic evaluation, or used total joint replacement as a condition for terminal stage of the disease. Both have advantages and disadvantages. ROA, usually accompanied with evaluation scores, such as Kellgren-Lawrence (K-L) score, allows detection of early changes in the anatomy. However, ROA is very common and most ROA are not symptomatic [48]. Universal agreement of disease phenotypes of OA must be generated in order to conduct global collaboration and cross-reference.

#### **Clinical implications**

Several studies have focused on developing reliable biomarkers that can identify individual at risk of developing OA or at early stage of the disease. Protein levels in serum and synovial fluid were tested; some were successful and some were not. Among the promising candidates are collagen type II, glycoprotein 39, COMP, BMP-7, osteopontin, and endoglin [33, 49-53]. Markers to identify OA in its early stages are very important as they enable early interventions. Further knowledge on molecular pathways underlying the disease will allow direct and effective therapy.

Thus far, clinical implications of OA had been on obesity. This is because of the biomechanical overloading and the biomolecular effects. Leptin, an obesity gene, regulate *MMP-13* expression in chondrocytes [41] and a microRNA differentially expressed in OA is also associated with high body mass index [42]. Weight control may modulate the adipokine profile and delay the disease incidence or progression [54].

#### **Conclusion**

While the Human Genome Project and the HapMap Project help uncovering many of the genetic determinants of complex disorders, to fully understand the genetics underlying OA pathogenesis, a global, or at least, a continental collaboration is crucial to overcome these challenges. From **Table 1**, it is clear

that many candidate genes have not been studied in Asian cohort, whose genetic compositions are undoubtedly different from that of European. While most studies of Asian cohort were performed in Japanese and Han Chinese, very few were on Southeast Asian. It is estimated that more than one million senior citizens in Thailand are suffering from

OA [55], though not many studies have been done on genetics of OA in Thais. Moreover, replication associations for a complex disease like OA are crucial in understanding the disease etiology. It is also important to distinguish between the true and false positives.

**Table 1.** Selected genes associated with osteoarthritis.

Gene	Protein	Function	OA association	Association in Asian subjects*
<b>Inflammation</b>				
<i>CCL2</i>	Chemokine (C-C motif) ligand 2	Immunoregulatory and inflammation. Binds to chemokine receptors cCR2 and CCR4	Differentially expressed in OA bone [56]	A polymorphism in the promoter is associated with primary knee OA in Korean [57]
<i>COX2</i> ( <i>PTGS2</i> )	Cyclooxygenase 2 (Prostaglandin G/H SYNTHASE 2)	Osteogenesis and bone repair	Associated with hip and knee OA [58, 59]	-
<i>IL-1</i> gene cluster	Interleukin 1- $\alpha$ , $\beta$ , interleukin 1 receptor antagonist (IL1RN)	Stimulate osteoclast activity in vitro, increase production of metalloproteinases and aggrecanases, in turn stimulate cartilage degradation	Polymorphisms in IL1RN are associated with disease severity in knee OA [60] and implicated in a meta-analysis [61]	Negative finding in Turkish population [62]
<i>IL-6</i>	Interleukin-6 Interferon $\beta$ -2	Stimulates osteoclasts, a polymorphism involved in BMD	Increased expression in radiographic knee OA in British women [63] and in osteophytes [64]. A polymorphism is associated with osteolysis [65] and distal interphalangeal OA [66]	-
<i>IL-10</i>	Interleukin -10	Anti-inflammatory, prevents cartilage destruction by reducing IL-1 $\beta$ and TNF- $\beta$ expression in articular chondrocytes in a mouse model [67]	Associated with knee OA in Greek [68] and distal interphalangeal OA [69]	-
<i>HLA</i>	Human leukocyte antigen system	Antigen presentation, associated with rheumatoid arthritis	HLA-DRB1 alleles are associated with OA in Italian [70] and German [71] and distal interphalangeal OA in Dutch [72]	HLA class I is associated with generalized OA in Japanese [73]

**Table 1.** Selected genes associated with osteoarthritis (Continued).

Gene	Protein	Function	OA association	Association in Asian subjects*
<b>Extracellular matrix molecules</b>				
<i>ASPN</i>	Asporin, Periodontal ligament-associated protein 1	Associated with cartilage matrix. Suppresses TGF- $\beta$ -mediated effects and reduces proteoglycan accumulation [14]	Knee OA and hip OA [14, 74-76] Found associated by meta-analyses [10, 77] Negative finding in Spanish Caucasians [78]	Associated with knee and hip OA in Japanese [14], knee OA [74] and age at onset [79] in Chinese, female-specific knee OA in Korean [76]
<i>COL2A1</i>	Collagen, Type II $\alpha$ -1	Cartilage collagen	Increased expression in OA [11]. Associated with hip and knee OA in the Rotterdam Study [80], Dutch [25] and Caucasian men [10]. Negative finding in US [12] and Finnish with primary early-onset hip and/or knee OA cohort [13]	Associated with hip OA in Japanese [81]
<i>COMP</i>	Cartilage oligomeric matrix protein	Chondrocytes territorial matrix, a candidate gene for pseudoachondroplasia and hereditary osteochondral dysplasias with early-onset OA	Lower serum levels in hand and knee OA [82]. Negative finding in Caucasian [10]	Negative finding in Japanese [9]
<i>CILP</i>	Cartilage intermediate layer protein	Inhibits TGF $\beta$ 1-mediated induction of cartilage matrix genes. Increase in synthesis in early OA cartilage	Associated with knee OA prevalence and progression [26] and knee OA in men [5]	-
<i>MATN3</i>	Matrilin 3	Cartilage extracellular matrix protein. Involved in development and homeostasis of cartilage and bone	Increased expression in OA [83] Associated with hand OA, but not knee OA [84, 85]. Activates the expression of OA-associated genes [86]	-
<i>SPP1</i>	osteopontin (secreted phosphoprotein 1)	A bone matrix protein involving bone maturation and fracture repair.	Up-regulated in human OA chondrocytes [87]	High level in plasma and synovial fluid relative to knee OA severity in Thais [49] and Chinese [88]

**Table 1.** Selected genes associated with osteoarthritis (Continued).

Gene	Protein	Function	OA association	Association in Asian subjects*
<b>Genes/proteins in Wnt signaling pathway</b>				
<i>CALM1</i>	Calmodulin 1	A member of calcium-modulated protein family. Functions in growth, cell cycle and signal transduction. Regulates <i>COL2A1</i> expression	Negative findings in Caucasian [10] and UK Caucasian women with hip OA [43]	Associated with hip OA in Japanese [15]. Negative finding in Han Chinese [44]
<i>ENG</i>	Endoglin, CD105	Receptor for members of TGF- $\beta$ superfamily. Involves in angiogenesis, inflammation and wound healing	Up-regulated in RA and OA synovial tissues [89]	Elevated level in plasma and synovial fluid associated with knee OA severity in Thais [50]
<i>FRZB</i>	Secreted frizzled-related protein 3 (sFRP)	Antagonist in wnt-signaling pathway, modulates chondrocyte maturation	Associated with hip OA [19, 90], generalized OA, multiple joints OA [10, 91, 92]. These SNPs may leads to elevated serum sFRP level, which can be used as a biomarker.	-
<i>WISP-1</i>	Wnt-1-induced secreted protein 1	A target of the wnt pathway and is directly regulated by beta-catenin	Increased in expression in synovium and cartilage of mice with experimental OA and human OA, and induces expression of MMPs and aggrecanase [93]	Associated with spinal OA in post-menopausal Japanese women [20]
<b>Proteins related to modulation of osteocyte or chondrocyte differentiation</b>				
<i>BMP2</i>	Bone morphogenetic protein 2	A member of TGF- $\beta$ superfamily. Involved in chondrogenesis and osteogenesis, induces the formation of ectopic cartilage	Associated with prevalence of knee OA [5, 26]. mRNA localized in OA tissues [94]	-
<i>BMP5</i>	Bone morphogenetic protein 5	A member of TGF- $\beta$ superfamily. Induced endochondral osteogenesis in vivo	Mapped for a primary OA susceptibility locus with a functional microsatellite [30, 95]	-
<i>BMP7</i>	Bone morphogenetic protein 7 (osteogenic protein-1)	A regulator of cartilage and bone induction activity with roles in stimulation of proteoglycan synthesis and cartilage repair	Detected in synovial fluid of RA and OA patients [96]	High level in plasma and synovial fluid relative to knee OA severity in Thais [33]



**Table 1.** Selected genes associated with osteoarthritis (Continued).

Gene	Protein	Function	OA association	Association in Asian subjects*
<i>ESR1</i>	Estrogen receptor alpha	Associates with BMD. Modulates proteoglycan degradation and matrix metalloproteinase expression in chondrocytes	Radiographic knee and hip OA [5, 21, 24]	Associated with primary knee OA in Korean [22]
<i>GDF5</i>	Growth differentiation factor 5	A member of TGF- $\beta$ superfamily. Required for normal formation of bones and joints in the limbs, skull, and axial skeleton [97]	A functional SNP in the 5' UTR, which reduces the gene expression, is associated with OA in European, Spanish, and Asian population [32], but not in Greek population [45]	Associated with OA in Japanese and Chinese [31]
<i>OPG</i>	Osteoprotegerin (osteoclastogenesis inhibitory factor)	A secreted glycoprotein that regulates bone resorption, inhibits osteoclast differentiation	Associated with knee OA progression [5, 26]	-
<i>VDR</i>	Vitamin D receptor	Mediates effects of VitD, regulates osteoclastogenesis, involved in BMD and osteoporosis	Associated with hand OA (Finnish women) [98], osteophytes in knee and lumbar spine OA [23, 25] Some negative studies [12, 29]	Japanese: no association to hand, hip, knee OA [28]
<b>Protease/ protease inhibitors</b>				
<i>AACT</i>	Alpha-1-antichymotrypsin	a plasma protease inhibitor involved in cartilage proteoglycan degradation	Associate with knee OA progression [5, 26]	-
<i>ADAM12</i>	A disintegrin and metalloproteinase domain 12	A metalloproteinase involved in osteoclast formation, chondrocytes maturation and proliferation	Associated with prevalence and progression of knee OA [5, 26, 36]	-
<i>ADAMTS14</i>	ADAM with Thrombospondin motif (ADAMTS) 14	A metalloproteinase	Associated with knee OA in women [37]	-
<i>MMPs</i>	Matrix metalloproteinase	Major enzyme responsible for collagen and aggrecan degradation in articular cartilage	Elevated expression in serum, cartilage and synovial tissues of OA patients [99]	A polymorphism in <i>MMP-1</i> is associated with knee OA in Turkish [38].
<i>TNA</i>	Tetranectin	Extracellular matrix degradation. Induced during the mineralization phase of osteogenesis	Associate with knee OA progression [5, 26]	-

\* indicates that, to the author's knowledge, no association study of the gene and OA in Asian subjects has been reported at the time of this manuscript submission. Genes associated to OA are divided into five categories, as suggested by Valdes and Spector, 2009 [3].

Knowledge gained from studies in the last few years prompted researchers to look at OA as a systematic musculoskeletal disorder involving cell differentiation and metabolism, rather than a disease of articular cartilage as previously thought. Genetic component is exceedingly probable as an underlying cause of phenotypic variation, making some people more susceptible in developing OA or having the disease progress more rapidly than others. Reversion in gene expression in OA suggests that the cause of generalized OA may be unbalance bone metabolism. Some proteins are selected as candidate biomarkers for OA; some are therapeutic targets. Information gathered from clinical and basic research is highly significant in understanding and tackling this disabling and costly disease.

### Acknowledgement

This study was supported by Research Funds from the Faculty of Science, Chulalongkorn University, and the Thailand Research Fund. The author has no conflict of interest to report.

### References

1. Michaud CM, McKenna MT, Begg S, Tomijima N, Majmudar M, Bulzacchelli MT, et al. The burden of disease and injury in the United States 1996. *Popul Health Metr.* 2006; 4:11.
2. Spector TD, MacGregor AJ. Risk factors for osteoarthritis: genetics. *Osteoarthritis Cartilage.* 2004; 12:S39-44.
3. Valdes AM, Spector TD. The contribution of genes to osteoarthritis. *Med Clin North Am.* 2009 ; 93:45-66.
4. Lee YH, Rho YH, Choi SJ, Ji JD, Song GG. Osteoarthritis susceptibility loci defined by genome scan meta-analysis. *Rheumatol Int.* 2006; 26:996-1000.
5. Valdes AM, Van Oene M, Hart DJ, Surdulescu GL, Loughlin J, Doherty M, et al. Reproducible genetic associations between candidate genes and clinical knee osteoarthritis in men and women. *Arthritis Rheum.* 2006; 54:533-9.
6. Benito MJ, Veale DJ, FitzGerald O, van den Berg WB, Bresnihan B. Synovial tissue inflammation in early and late osteoarthritis. *Ann Rheum Dis.* 2005; 64:1263-7.
7. Leppavuori J, Kujala U, Kinnunen J, Kaprio J, Nissila M, Heliovaara M, et al. Genome scan for predisposing loci for distal interphalangeal joint osteoarthritis: evidence for a locus on 2q. *Am J Hum Genet.* 1999; 65: 1060-7.
8. Tseng S, Reddi AH, Di Cesare PE. Cartilage Oligomeric Matrix Protein (COMP): A Biomarker of Arthritis. *Biomark Insights.* 2009; 4:33-44.
9. Mabuchi A, Ikeda T, Fukuda A, Koshizuka Y, Hiraoka H, Miyoshi K, et al. Identification of sequence polymorphisms of the COMP (cartilage oligomeric matrix protein) gene and association study in osteoarthritis of the knee and hip joints. *J Hum Genet.* 2001; 46:456-62.
10. Valdes AM, Loughlin J, Oene MV, Chapman K, Surdulescu GL, Doherty M, et al. Sex and ethnic differences in the association of ASPN, CALM1, COL2A1, COMP, and FRZB with genetic susceptibility to osteoarthritis of the knee. *Arthritis Rheum.* 2007;56: 137-46.
11. Nelson F, Dahlberg L, Laverty S, Reiner A, Pidoux I, Ionescu M, et al. Evidence for altered synthesis of type II collagen in patients with osteoarthritis. *J Clin Invest.* 1998; 102:2115-25.
12. Baldwin CT, Cupples LA, Joost O, Demissie S, Chaisson C, McAlindon T, et al. Absence of linkage or association for osteoarthritis with the vitamin D receptor/type II collagen locus: the Framingham Osteoarthritis Study. *J Rheumatol.* 2002; 29:161-5.
13. Jakkula E, Melkonien M, Kiviranta I, Lohiniva J, Raina SS, Perala M, et al. The role of sequence variations within the genes encoding collagen II, IX and XI in non-syndromic, early-onset osteoarthritis. *Osteoarthritis Cartilage.* 2005; 13:497-507.
14. Kizawa H, Kou I, Iida A, Sudo A, Miyamoto Y, Fukuda A, et al. An aspartic acid repeat polymorphism in asporin inhibits chondrogenesis and increases susceptibility to osteoarthritis. *Nat Genet.* 2005; 37: 138-44.
15. Mototani H, Mabuchi A, Saito S, Fujioka M, Iida A, Takatori Y, et al. A functional single nucleotide polymorphism in the core promoter region of CALM1 is associated with hip osteoarthritis in Japanese. *Hum Mol Genet.* 2005; 14:1009-17.
16. Corr M. Wnt-beta-catenin signaling in the pathogenesis of osteoarthritis. *Nat Clin Pract Rheumatol.* 2008; 4: 550-6.
17. Kim SJ, Im DS, Kim SH, Ryu JH, Hwang SG, Seong JK, et al. Beta-catenin regulates expression of cyclooxygenase-2 in articular chondrocytes. *Biochem Biophys Res Commun.* 2002; 296:221-6.
18. Diarra D, Stolina M, Polzer K, Zwerina J, Ominsky MS, Dwyer D, et al. Dickkopf-1 is a master regulator of joint remodeling. *Nat Med.* 2007; 13:156-63.
19. Loughlin J, Dowling B, Chapman K, Marcelline L, Mustafa Z, Southam L, et al. Functional variants within

- the secreted frizzled-related protein 3 gene are associated with hip osteoarthritis in females. *Proc Natl Acad Sci U S A*. 2004; 101:9757-62.
20. Urano T, Narusawa K, Shiraki M, Usui T, Sasaki N, Hosoi T, et al. Association of a single nucleotide polymorphism in the WISP1 gene with spinal osteoarthritis in postmenopausal Japanese women. *J Bone Miner Metab*. 2007; 25:253-8.
  21. Bergink AP, van Meurs JB, Loughlin J, Arp PP, Fang Y, Hofman A, et al. Estrogen receptor alpha gene haplotype is associated with radiographic osteoarthritis of the knee in elderly men and women. *Arthritis Rheum*. 2003; 48:1913-22.
  22. Jin SY, Hong SJ, Yang HI, Park SD, Yoo MC, Lee HJ, et al. Estrogen receptor-alpha gene haplotype is associated with primary knee osteoarthritis in Korean population. *Arthritis Res Ther*. 2004; 6:R415-21.
  23. Jordan KM, Syddall H, Dennison EM, Cooper C, Arden NK. Birthweight, vitamin D receptor gene polymorphism, and risk of lumbar spine osteoarthritis. *J Rheumatol*. 2005; 32:678-83.
  24. Lian K, Lui L, Zmuda JM, Nevitt MC, Hochberg MC, Lee JM, et al. Estrogen receptor alpha genotype is associated with a reduced prevalence of radiographic hip osteoarthritis in elderly Caucasian women. *Osteoarthritis Cartilage*. 2007; 15:972-8.
  25. Uitterlinden AG, Burger H, van Duijn CM, Huang Q, Hofman A, Birkenhager JC, et al. Adjacent genes, for COL2A1 and the vitamin D receptor, are associated with separate features of radiographic osteoarthritis of the knee. *Arthritis Rheum*. 2000; 43:1456-64.
  26. Valdes AM, Hart DJ, Jones KA, Surdulescu G, Swarbrick P, Doyle DV, et al. Association study of candidate genes for the prevalence and progression of knee osteoarthritis. *Arthritis Rheum*. 2004; 50: 2497-507.
  27. Bid HK, Mishra DK, Mittal RD. Vitamin-D receptor (VDR) gene (Fok-I, Taq-I and Apa-I) polymorphisms in healthy individuals from north Indian population. *Asian Pac J Cancer Prev*. 2005; 6:147-52.
  28. Huang J, Ushiyama T, Inoue K, Kawasaki T, Hukuda S. Vitamin D receptor gene polymorphisms and osteoarthritis of the hand, hip, and knee: a case-control study in Japan. *Rheumatology (Oxford)*. 2000; 39: 79-84.
  29. Lee YH, Woo JH, Choi SJ, Ji JD, Song GG. Vitamin D receptor TaqI, BsmI and ApaI polymorphisms and osteoarthritis susceptibility: A meta-analysis. *Joint Bone Spine*. 2009; 76:156-61.
  30. Wilkins JM, Southam L, Mustafa Z, Chapman K, Loughlin J. Association of a functional microsatellite within intron 1 of the BMP5 gene with susceptibility to osteoarthritis. *BMC Med Genet*. 2009; 10:141.
  31. Miyamoto Y, Mabuchi A, Shi D, Kubo T, Takatori Y, Saito S, et al. A functional polymorphism in the 5' UTR of GDF5 is associated with susceptibility to osteoarthritis. *Nat Genet*. 2007; 39:529-33.
  32. Southam L, Rodriguez-Lopez J, Wilkins JM, Pombo-Suarez M, Snelling S, Gomez-Reino JJ, et al. An SNP in the 5'-UTR of GDF5 is associated with osteoarthritis susceptibility in Europeans and with in vivo differences in allelic expression in articular cartilage. *Hum Mol Genet*. 2007; 16:2226-32.
  33. Honsawek S, Chayanupatkul M, Tanavalee A, Sakdinakittikoon M, Deepaisarnsakul B, Yuktanandana P, et al. Relationship of plasma and synovial fluid BMP-7 with disease severity in knee osteoarthritis patients: a pilot study. *Int Orthop*. 2009; 33:1171-5.
  34. Hayashi M, Muneta T, Ju YJ, Mochizuki T, Sekiya I. Weekly intra-articular injections of bone morphogenetic protein-7 inhibits osteoarthritis progression. *Arthritis Res Ther*. 2008; 10:R118.
  35. Meulenbelt I, Min JL, Bos S, Riyazi N, Houwing-Duistermaat JJ, van der Wijk HJ, et al. Identification of DIO2 as a new susceptibility locus for symptomatic osteoarthritis. *Hum Mol Genet*. 2008; 17:1867-75.
  36. Kerna I, Kisand K, Tamm AE, Lintrop M, Veske K, Tamm AO. Missense single nucleotide polymorphism of the ADAM12 gene is associated with radiographic knee osteoarthritis in middle-aged Estonian cohort. *Osteoarthritis Cartilage*. 2009; 17:1093-8.
  37. Rodriguez-Lopez J, Pombo-Suarez M, Loughlin J, Tsezou A, Blanco FJ, Meulenbelt I, et al. Association of a nsSNP in ADAMTS14 to some osteoarthritis phenotypes. *Osteoarthritis Cartilage*. 2009; 17:321-7.
  38. Barlas IO, Sezgin M, Erdal ME, Sahin G, Ankarali HC, Altintas ZM, et al. Association of (-1,607) 1G/2G polymorphism of matrix metalloproteinase-1 gene with knee osteoarthritis in the Turkish population (knee osteoarthritis and MMPs gene polymorphisms). *Rheumatol Int*. 2009; 29:383-8.
  39. Lories RJ, Peeters J, Bakker A, Tylzanowski P, Derese I, Schrooten J, et al. Articular cartilage and biomechanical properties of the long bones in Frzb-knockout mice. *Arthritis Rheum*. 2007; 56:4095-103.
  40. Roach HI, Aigner T. DNA methylation in osteoarthritic chondrocytes: a new molecular target. *Osteoarthritis Cartilage*. 2007; 15:128-37.
  41. Iliopoulos D, Malizos KN, Tsezou A. Epigenetic

- regulation of leptin affects MMP-13 expression in osteoarthritic chondrocytes: possible molecular target for osteoarthritis therapeutic intervention. *Ann Rheum Dis*. 2007; 66:1616-21.
42. Iliopoulos D, Malizos KN, Oikonomou P, Tsezou A. Integrative microRNA and proteomic approaches identify novel osteoarthritis genes and their collaborative metabolic and inflammatory networks. *PLoS ONE*. 2008; 3:e3740.
  43. Loughlin J, Sinsheimer JS, Carr A, Chapman K. The CALM1 core promoter polymorphism is not associated with hip osteoarthritis in a United Kingdom Caucasian population. *Osteoarthritis Cartilage*. 2006; 14:295-8.
  44. Shi D, Ni H, Dai J, Qin J, Xu Y, Zhu L, et al. Lack of association between the CALM1 core promoter polymorphism (-16C/T) and susceptibility to knee osteoarthritis in a Chinese Han population. *BMC Med Genet*. 2008; 9:91.
  45. Tsezou A, Satra M, Oikonomou P, Bargiotas K, Malizos KN. The growth differentiation factor 5 (GDF5) core promoter polymorphism is not associated with knee osteoarthritis in the Greek population. *J Orthop Res*. 2008; 26:136-40.
  46. Kochi Y, Suzuki A, Yamada R, Yamamoto K. Genetics of rheumatoid arthritis: underlying evidence of ethnic differences. *J Autoimmun*. 2009; 32:158-62.
  47. Glasson SS, Askew R, Sheppard B, Carito B, Blanchet T, Ma HL, et al. Deletion of active ADAMTS5 prevents cartilage degradation in a murine model of osteoarthritis. *Nature*. 2005; 434:644-8.
  48. Ikegawa S. New gene associations in osteoarthritis: what do they provide, and where are we going? *Curr Opin Rheumatol*. 2007; 19:429-34.
  49. Honsawek S, Tanavalee A, Sakdinakittikoon M, Chayanupatkul M, Yuktanandana P. Correlation of plasma and synovial fluid osteopontin with disease severity in knee osteoarthritis. *Clin Biochem*. 2009; 42: 808-12.
  50. Honsawek S, Tanavalee A, Yuktanandana P. Elevated circulating and synovial fluid endoglin are associated with primary knee osteoarthritis severity. *Arch Med Res*. 2009; 40:590-4.
  51. Li WW, Nemirovskiy O, Fountain S, Rodney Mathews W, Szekely-Klepser G. Clinical validation of an immunoaffinity LC-MS/MS assay for the quantification of a collagen type II neoepitope peptide: A biomarker of matrix metalloproteinase activity and osteoarthritis in human urine. *Anal Biochem*. 2007; 369:41-53.
  52. Sharif M, Granell R, Johansen J, Clarke S, Elson C, Kirwan JR. Serum cartilage oligomeric matrix protein and other biomarker profiles in tibiofemoral and patellofemoral osteoarthritis of the knee. *Rheumatology (Oxford)*. 2006; 45:522-6.
  53. Zivanovic S, Rackov LP, Vojvodic D, Vucetic D. Human cartilage glycoprotein 39—biomarker of joint damage in knee osteoarthritis. *Int Orthop*. 2009; 33:1165-70.
  54. Aspden RM. Osteoarthritis: a problem of growth not decay? *Rheumatology (Oxford)*. 2008; 47:1452-60.
  55. Ministry of Public Health of Thailand. Thai seniors encounter degenerative diseases. (online). 2010 [cited 15 June 2010]. Available from: [http://www.moph.go.th/show\\_hotnew.php?idHot\\_new=30798](http://www.moph.go.th/show_hotnew.php?idHot_new=30798).
  56. Sanchez-Sabate E, Alvarez L, Gil-Garay E, Munuera L, Vilaboa N. Identification of differentially expressed genes in trabecular bone from the iliac crest of osteoarthritic patients. *Osteoarthritis Cartilage*. 2009; 17:1106-14.
  57. Park HJ, Yoon SH, Zheng LT, Lee KH, Kim JW, Chung JH, et al. Association of the -2510A/G chemokine (C-C motif) ligand 2 polymorphism with knee osteoarthritis in a Korean population. *Scand J Rheumatol*. 2007; 36: 299-306.
  58. Valdes AM, Loughlin J, Timms KM, van Meurs JJ, Southam L, Wilson SG, et al. Genome-wide association scan identifies a prostaglandin-endoperoxide synthase 2 variant involved in risk of knee osteoarthritis. *Am J Hum Genet*. 2008; 82:1231-40.
  59. Schneider EM, Du W, Fiedler J, Hogel J, Gunther KP, Brenner H, et al. The (-765 G->C) promoter variant of the COX-2/PTGS2 gene is associated with a lower risk for end-stage hip and knee osteoarthritis. *Ann Rheum Dis (online)*. 2010 Apr 8 [cited 15 June 2010]. Available from: <http://ard.bmj.com/content/early/2010/03/02/ard.2009.124040.long>.
  60. Attur M, Wang HY, Byers Kraus V, Bukowski JF, Aziz N, Krasnokutsky S, et al. Radiographic severity of knee osteoarthritis is conditional on interleukin-1 receptor antagonist gene variations. *Ann Rheum Dis*. 2010; 69: 856-61.
  61. Moxley G, Meulenbelt I, Chapman K, van Duijn CM, Eline Slagboom P, Neale MC, et al. Interleukin-1 region meta-analysis with osteoarthritis phenotypes. *Osteoarthritis Cartilage*. 2010; 18:200-7.
  62. Sezgin M, Erdal ME, Altintas ZM, Ankarali HC, Barlas IO, Turkmen E, et al. Lack of association polymorphisms of the IL1RN, IL1A, and IL1B genes with knee osteoarthritis in Turkish patients. *Clin Invest Med*. 2007; 30:E86-92.
  63. Livshits G, Zhai G, Hart DJ, Kato BS, Wang H, Williams



- FM, et al. Interleukin-6 is a significant predictor of radiographic knee osteoarthritis: The Chingford Study. *Arthritis Rheum.* 2009; 60:2037-45.
64. Sakao K, Takahashi KA, Arai Y, Saito M, Honjo K, Hiraoka N, et al. Osteoblasts derived from osteophytes produce interleukin-6, interleukin-8, and matrix metalloproteinase-13 in osteoarthritis. *J Bone Miner Metab.* 2009; 27:412-23.
65. Gordon A, Kiss-Toth E, Stockley I, Eastell R, Wilkinson JM. Polymorphisms in the interleukin-1 receptor antagonist and interleukin-6 genes affect risk of osteolysis in patients with total hip arthroplasty. *Arthritis Rheum.* 2008; 58:3157-65.
66. Kamarainen OP, Solovieva S, Vehmas T, Luoma K, Riihimäki H, Ala-Kokko L, et al. Common interleukin-6 promoter variants associate with the more severe forms of distal interphalangeal osteoarthritis. *Arthritis Res Ther.* 2008; 10:R21.
67. Joosten LA, Lubberts E, Durez P, Helsen MM, Jacobs MJ, Goldman M, et al. Role of interleukin-4 and interleukin-10 in murine collagen-induced arthritis. Protective effect of interleukin-4 and interleukin-10 treatment on cartilage destruction. *Arthritis Rheum.* 1997; 40:249-60.
68. Fytili P, Giannatou E, Karachalios T, Malizos K, Tsezou A. Interleukin-10G and interleukin-10R microsatellite polymorphisms and osteoarthritis of the knee. *Clin Exp Rheumatol.* 2005; 23:621-7.
69. Riyazi N, Kurreeman FA, Huizinga TW, Dekker FW, Stoeken-Rijsbergen G, Kloppenburg M. The role of interleukin 10 promoter polymorphisms in the susceptibility of distal interphalangeal osteoarthritis. *J Rheumatol.* 2005; 32:1571-5.
70. Rovetta G, Buffrini L, Monteforte P, Grignolo MC, Molfetta L. HLA-DRB1 alleles and osteoarthritis in a group of patients living in Liguria-Italy. *Minerva Med.* 2006; 97:271-5.
71. Moos V, Menard J, Sieper J, Sparmann M, Muller B. Association of HLA-DRB1\*02 with osteoarthritis in a cohort of 106 patients. *Rheumatology (Oxford).* 2002; 41:666-9.
72. Riyazi N, Spee J, Huizinga TW, Schreuder GM, de Vries RR, Dekker FW, et al. HLA class II is associated with distal interphalangeal osteoarthritis. *Ann Rheum Dis.* 2003; 62:227-30.
73. Wakitani S, Imoto K, Mazuka T, Kim S, Murata N, Yoneda M. Japanese generalised osteoarthritis was associated with HLA class I—a study of HLA-A, B, Cw, DQ, DR in 72 patients. *Clin Rheumatol.* 2001; 20:417-9.
74. Jiang Q, Shi D, Yi L, Ikegawa S, Wang Y, Nakamura T, et al. Replication of the association of the aspartic acid repeat polymorphism in the asporin gene with knee-osteoarthritis susceptibility in Han Chinese. *J Hum Genet.* 2006; 51:1068-72.
75. Kaliakatsos M, Tzetzis M, Kanavakis E, Fytili P, Chouliaras G, Karachalios T, et al. Asporin and knee osteoarthritis in patients of Greek origin. *Osteoarthritis Cartilage.* 2006; 14:609-11.
76. Song JH, Lee HS, Kim CJ, Cho YG, Park YG, Nam SW, et al. Aspartic acid repeat polymorphism of the asporin gene with susceptibility to osteoarthritis of the knee in a Korean population. *Knee.* 2008; 15:191-5.
77. Nakamura T, Shi D, Tzetzis M, Rodriguez-Lopez J, Miyamoto Y, Tsezou A, et al. Meta-analysis of association between the ASPN D-repeat and osteoarthritis. *Hum Mol Genet.* 2007; 16:1676-81.
78. Rodriguez-Lopez J, Pombo-Suarez M, Liz M, Gomez-Reino JJ, Gonzalez A. Lack of association of a variable number of aspartic acid residues in the asporin gene with osteoarthritis susceptibility: case-control studies in Spanish Caucasians. *Arthritis Res Ther.* 2006; 8:R55.
79. Shi D, Nakamura T, Dai J, Yi L, Qin J, Chen D, et al. Association of the aspartic acid-repeat polymorphism in the asporin gene with age at onset of knee osteoarthritis in Han Chinese population. *J Hum Genet.* 2007; 52:664-7.
80. Meulenbelt I, Bijkerk C, De Wildt SC, Miedema HS, Breedveld FC, Pols HA, et al. Haplotype analysis of three polymorphisms of the COL2A1 gene and associations with generalised radiological osteoarthritis. *Ann Hum Genet.* 1999; 63:393-400.
81. Ikeda T, Mabuchi A, Fukuda A, Kawakami A, Ryo Y, Yamamoto S, et al. Association analysis of single nucleotide polymorphisms in cartilage-specific collagen genes with knee and hip osteoarthritis in the Japanese population. *J Bone Miner Res.* 2002; 17:1290-6.
82. Chen HC, Shah SH, Li YJ, Stabler TV, Jordan JM, Kraus VB. Inverse association of general joint hypermobility with hand and knee osteoarthritis and serum cartilage oligomeric matrix protein levels. *Arthritis Rheum.* 2008; 58:3854-64.
83. Pullig O, Weseloh G, Klatt AR, Wagener R, Swoboda B. Matrilin-3 in human articular cartilage: increased expression in osteoarthritis. *Osteoarthritis Cartilage.* 2002; 10:253-63.
84. Min JL, Meulenbelt I, Riyazi N, Kloppenburg M, Houwing-Duistermaat JJ, Seymour AB, et al. Association of matrilin-3 polymorphisms with spinal disc degeneration and osteoarthritis of the first



- carpometacarpal joint of the hand. *Ann Rheum Dis.* 2006; 65:1060-6.
85. Pullig O, Tagariello A, Schweizer A, Swoboda B, Schaller P, Winterpacht A. MATN3 (matrilin-3) sequence variation (pT303M) is a risk factor for osteoarthritis of the CMC1 joint of the hand, but not for knee osteoarthritis. *Ann Rheum Dis.* 2007; 66: 279-80.
  86. Klatt AR, Klinger G, Paul-Klausch B, Kuhn G, Renno JH, Wagener R, et al. Matrilin-3 activates the expression of osteoarthritis-associated genes in primary human chondrocytes. *FEBS Lett.* 2009; 583:3611-7.
  87. Pullig O, Weseloh G, Gauer S, Swoboda B. Osteopontin is expressed by adult human osteoarthritic chondrocytes: protein and mRNA analysis of normal and osteoarthritic cartilage. *Matrix Biol.* 2000; 19: 245-55.
  88. Gao SG, Li KH, Zeng KB, Tu M, Xu M, Lei GH. Elevated osteopontin level of synovial fluid and articular cartilage is associated with disease severity in knee osteoarthritis patients. *Osteoarthritis Cartilage.* 2010; 18, 82-7.
  89. Szekanecz Z, Haines GK, Harlow LA, Shah MR, Fong TW, Fu R, et al. Increased synovial expression of transforming growth factor (TGF)-beta receptor endoglin and TGF-beta 1 in rheumatoid arthritis: possible interactions in the pathogenesis of the disease. *Clin Immunol Immunopathol.* 1995; 76:187-94.
  90. Lane NE, Lian K, Nevitt MC, Zmuda JM, Lui L, Li J, et al. Frizzled-related protein variants are risk factors for hip osteoarthritis. *Arthritis Rheum.* 2006; 54: 1246-54.
  91. Min JL, Meulenbelt I, Riyazi N, Kloppenburg M, Houwing-Duistermaat JJ, Seymour AB, et al. Association of the Frizzled-related protein gene with symptomatic osteoarthritis at multiple sites. *Arthritis Rheum.* 2005; 52:1077-80.
  92. Rodriguez-Lopez J, Pombo-Suarez M, Liz M, Gomez-Reino JJ, Gonzalez A. Further evidence of the role of frizzled-related protein gene polymorphisms in osteoarthritis. *Ann Rheum Dis.* 2007; 66:1052-5.
  93. Blom AB, Brockbank SM, van Lent PL, van Beuningen HM, Geurts J, Takahashi N, et al. Involvement of the Wnt signaling pathway in experimental and human osteoarthritis: prominent role of Wnt-induced signaling protein 1. *Arthritis Rheum.* 2009; 60:501-12.
  94. Nakase T, Miyaji T, Tomita T, Kaneko M, Kuriyama K, Myoui A, et al. Localization of bone morphogenetic protein-2 in human osteoarthritic cartilage and osteophyte. *Osteoarthritis Cartilage.* 2003; 11:278-84.
  95. Southam L, Dowling B, Ferreira A, Marcelline L, Mustafa Z, Chapman K, et al. Microsatellite association mapping of a primary osteoarthritis susceptibility locus on chromosome 6p12.3-q13. *Arthritis Rheum.* 2004; 50: 3910-4.
  96. Chubinskaya S, Frank BS, Michalska M, Kumar B, Merrihew CA, Thonar EJ, et al. Osteogenic protein 1 in synovial fluid from patients with rheumatoid arthritis or osteoarthritis: relationship with disease and levels of hyaluronan and antigenic keratan sulfate. *Arthritis Res Ther.* 2006; 8:R73.
  97. Settle SH, Jr., Rountree RB, Sinha A, Thacker A, Higgins K, Kingsley DM. Multiple joint and skeletal patterning defects caused by single and double mutations in the mouse *Gdf6* and *Gdf5* genes. *Dev Biol.* 2003; 254: 116-30.
  98. Solovieva S, Hirvonen A, Siivola P, Vehmas T, Luoma K, Riihimäki H, et al. Vitamin D receptor gene polymorphisms and susceptibility of hand osteoarthritis in Finnish women. *Arthritis Res Ther.* 2006; 8:R20.
  99. Hulejova H, Baresova V, Klezl Z, Polanska M, Adam M, Senolt L. Increased level of cytokines and matrix metalloproteinases in osteoarthritic subchondral bone. *Cytokine.* 2007; 38:151-6.